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(21) International Application Number: PCT/US87/00987 (22) International Filing Date: 30 April 1987 (30.04.87) (31) Priority Application Number: 858,354 (32) Priority Date: 30 April 1986 (30.04.86) (33) Priority Country: US (60) Parent Application or Grant (63) Related by Continuation US 858,354 (CIP) Filed on 30 April 1986 (30.04.86) (71) Applicant (for all designated States except US): IGEN, INC. [US/US]; 1530 East Jefferson Street, Rockville, MD 20852 (US). (72) Inventors; and (75) Inventors/Applicants (for US only): MASSEY, Richard, J. [US/US]; 10201 Grosvenor Place, Rockville, MD 20852 (US). POWELL, Michael, J. [GB/US]; 5 War Admiral Way, Gaithersburg, MD 20878 (US). MIED, Paul, A. [US/US]; 3713 Buffalo Road, New Windsor, MD 21776 (US). FENG, Peter [CN/US]; 245 Rollins Avenue, Rockville, MD 20852 (US). DELLA CIANA, Leopoldo [IT/US];		5511 Halpine Place, Rockville, MD 20851 (US). DRESSICK, Walter, J. [US/US]; 12905 Crookston Lane, Rockville, MD 20851 (US). POONIAN, Mohindar, S. [US/US]; 224 High Timber Court, Gaithersburg, MD 20879 (US). (74) Agent: WHITE, John, P.; Cooper, Dunham, Griffin & Moran, 30 Rockefeller Plaza, New York, NY 10112 (US). (81) Designated States: AT (European patent), AU, BE (European patent), CH (European patent), DE (European patent), DK, FI, FR (European patent), GB (European patent), IT (European patent), JP, KR, LU (European patent), NL (European patent), NO, SE (European patent), US. Published With international search report. Before the expiration of the time limit for amending the claims and to be republished in the event of the receipt of amendments.																									
(54) Title: ELECTROCHEMILUMINESCENT ASSAYS																											
(57) Abstract																											
<p>Qualitative and quantitative electrochemiluminescent assays for analytes of interest present in multicomponent liquids. These methods comprise contacting a sample with a reagent labeled with an electrochemiluminescent chemical moiety and capable of combining with the analyte of interest, exposing the resulting sample to electrochemical energy and detecting electromagnetic radiation emitted by the electrochemiluminescent chemical moiety. Further provided are methods for detecting and identifying the presence of a multiplicity of analytes in a liquid food or food homogenate. These methods comprise immersing a diagnostic reagent holder, provided with a multiplicity of reagents, into the food or food homogenate, removing the diagnostic reagent holder from the liquid food or food homogenate, and detecting and identifying the presence of a multiplicity of analytes of interest bound to the diagnostic reagent holder, thereby detecting and identifying the presence of a multiplicity of analytes of interest in the food or food homogenate. The invention further provides an enzyme immunoassay for coliform bacteria. This assay comprises inoculating a sample into a suitable medium for coliform reproduction, immobilizing coliforms present in the medium to a suitable surface, treating the surface with an antibody directed to the immobilized coliforms and detecting the presence of the immobilized coliforms immobilized to a suitable surface.</p>																											
		<table border="1"> <caption>Estimated data from the graph</caption> <thead> <tr> <th>Theophylline Concentration (mg/ml)</th> <th>Luminescent Counts (thousands) - Curve 1 (Squares)</th> <th>Luminescent Counts (thousands) - Curve 2 (Circles)</th> <th>Luminescent Counts (thousands) - Curve 3 (Triangles)</th> </tr> </thead> <tbody> <tr> <td>0</td> <td>10.0</td> <td>10.0</td> <td>10.0</td> </tr> <tr> <td>10</td> <td>18.0</td> <td>16.0</td> <td>15.0</td> </tr> <tr> <td>20</td> <td>25.0</td> <td>22.0</td> <td>21.0</td> </tr> <tr> <td>30</td> <td>28.0</td> <td>24.0</td> <td>23.0</td> </tr> <tr> <td>40</td> <td>29.0</td> <td>25.0</td> <td>24.0</td> </tr> </tbody> </table>		Theophylline Concentration (mg/ml)	Luminescent Counts (thousands) - Curve 1 (Squares)	Luminescent Counts (thousands) - Curve 2 (Circles)	Luminescent Counts (thousands) - Curve 3 (Triangles)	0	10.0	10.0	10.0	10	18.0	16.0	15.0	20	25.0	22.0	21.0	30	28.0	24.0	23.0	40	29.0	25.0	24.0
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ELECTROCHEMILUMINESCENT ASSAYSBackground of the Invention

5 Within this application several publications are referenced by Arabic numerals within parentheses. Full citations for these references may be found at the end of the specification immediately preceding the claims. The disclosures of these publications in their entirety are hereby incorporated by reference into this application in order to more fully describe the state of the art to which this invention pertains.

15 There is a continuous and expanding need for rapid, highly specific methods of detecting and quantifying chemical, biochemical, and biological substances. Of particular value are methods for measuring small quantities of pharmaceuticals, metabolites, microorganisms and other materials of diagnostic value. Examples of such materials include narcotics and poisons, drugs administered for therapeutic purposes, hormones, pathogenic microorganisms and viruses, antibodies, metabolites, enzymes and nucleic acids.

25 The presence of these materials can often be determined by binding methods which exploit the high degree of specificity which characterizes many biochemical and biological systems. Frequently used methods are based on, for example, antigen-antibody systems, nucleic acid hybridization techniques, and protein-ligand systems. In these methods, the existence of a complex of diagnostic value is typically indicated by the presence or absence of an observable "label" which has been attached to one or more of the complexing materials.

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The specific labeling method chosen often dictates the usefulness and versatility of a particular system for detecting a material of interest. A preferred label should be inexpensive, safe, and capable of being attached efficiently to a wide variety of chemical, biochemical, and biological materials without changing the important binding characteristics of those materials.

5 The label should give a highly characteristic signal, and should be rarely, and preferably never, found in nature. The label should be stable and detectable in aqueous systems over periods of time ranging up to months. Detection of the label should be rapid, sensitive, and reproducible without the need for expensive, specialized facilities or personnel. Quantification of the label should be relatively independent of variables such as temperature and the composition of the mixture to be assayed. Most advantageous are labels

10 which can be used in homogeneous systems, i.e., systems in which separation of the complexed and uncomplexed labeled material is not necessary. This is possible if the detectability of the label is modulated when the labeled material is incorporated into a specific complex.

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A wide variety of labels have been developed, each with particular advantages and disadvantages. For example, radioactive labels are quite versatile, and can be

25 detected at very low concentrations. However, they are expensive, hazardous, and their use requires sophisticated equipment and trained personnel. Furthermore, the sensitivity of radioactive labels is limited by the fact that the detectable event can, in its essential

30 nature, occur only once per radioactive atom in the labeled material. Moreover, radioactive labels cannot be used in homogeneous methods.

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Thus, there is wide interest in non-radioactive labels. These include molecules observable by spectrophotometric, spin resonance, and luminescence techniques, as well as enzymes which produce such molecules. Among the useful non-radioactive labeling materials are organometallic compounds. Because of the rarity of some metals in biological systems, methods which specifically assay the metal component of the organometallic compounds can be successfully exploited. For example, Cais, U.S. Patent No. 4,205,952 (1980) discloses the use of immunochemically active materials labeled with certain organometallic compounds for use in quantitating specific antigens. Any general method of detecting the chosen metals can be used with these labels, including emission, absorption and fluorescence spectroscopy, atomic absorption, and neutron activation. These methods often suffer from lack of sensitivity, can seldom be adapted to a homogeneous system, and as with atomic absorption, sometimes entail destruction of the sample.

Of particular interest are labels which can be made to luminesce through photochemical, chemical, and electrochemical means. "Photoluminescence" is the process whereby a material is induced to luminesce when it absorbs electromagnetic radiation. Fluorescence and phosphorescence are types of photoluminescence. "Chemiluminescent" processes entail the creation of the luminescent species by a chemical transfer of energy. "Electrochemiluminescence" entails the creation of the luminescent species electrochemically.

These luminescent systems are of increasing importance. For example, Mandle, U.S. Patent No. 4,372,745 (1983)

discloses the use of chemiluminescent labels in immunochemical applications. In the disclosed systems, the labels are excited into a luminescent state by chemical means such as by reaction of the label with H_2O_2 and an oxalate. In these systems, H_2O_2 oxidatively converts the oxalate into a high energy derivative, which then excites the label. This system will, in principle, work with any luminescent material that is stable in the oxidizing conditions of the assay and can be excited by the high energy oxalate derivative. Unfortunately, this very versatility is the source of a major limitation of the technique: typical biological fluids containing the analyte of interest also contain a large number of potentially luminescent substances that can cause high background levels of luminescence.

Another example of the immunochemical use of chemiluminescence which suffers from the same disadvantages is Oberhardt et al., U.S. Patent No. 4,280,815, (1981) who disclose the in situ electrochemical generation of an oxidant (e.g., H_2O_2) in close proximity to an immunoreactant labeled with a chemiluminescent species. The electrogenerated oxidant diffuses to the chemiluminescent species and chemically oxidizes it, resulting in the net transfer of one or more electrons to the electrogenerated oxidant. Upon oxidation, the chemiluminescent species emits a photon. In contrast, the subject invention requires the direct transfer of electrons from a source of electrochemical energy to a chemiluminescent species which is capable of repeatedly emitting photons.

The present invention is concerned with electrochemiluminescent labels. Suitable labels comprise electrochemiluminescent compounds, including organic compounds

and organometallic compounds. Electrochemiluminescent methods of determining the presence of labeled materials are preferred over other methods for many reasons. They are highly diagnostic of the presence of a particular label, sensitive, nonhazardous, inexpensive, and can be used in a wide variety of applications. Organic compounds which are suitable electrochemical labels include, for example, rubrene and 9,10-diphenyl anthracene. Many organometallic compounds are suitable electrochemical labels, but of particular use are Ru-containing and Os-containing compounds.

Thus, in one embodiment, the present invention is concerned with the use of Ru-containing and Os-containing labels which can be detected by a wide variety of methods. These labels are advantageous for many reasons that will be discussed herein.

Ru-containing and Os-containing organometallic compounds have been discussed in the literature. Cais discloses that any metal element or combination of metal elements, including noble metals from group VIII such as Ru, would be suitable components of organometallic labels detectable by atomic absorption methods. (Cais, column 11, line 20). However, ruthenium is not a preferred metal in Cais, osmium is not specifically mentioned, no data are presented on the efficiency of using Ru or Os in any of the methods disclosed, and the preferred method of detection, atomic absorption, entails destruction of the sample.

Weber, U.S. Patent No. 4,293,310 (1981), discloses the use of Ru-containing and Os-containing complexes as electrochemical labels for analytes in immunoassays. The disclosed complexes are linked to amino groups on

the analytes through a thiourea linkage. Weber also suggests the possibility of forming carboxylate esters between the labels and hydroxy groups on other analytes.

According to Weber, the presence of the labeled materials can be determined with an apparatus and method which comprises a quencher and an electrochemical flow cell with light means. The photoelectrochemically active label upon photoexcitation transfers an electron to a quencher molecule; the oxidized molecule is subsequently reduced with an electron from an electrode of the flow cell which is held at suitable potential. This electron is measured as photocurrent. The amount of free labelled analyte in the system is determined by the photocurrent signal. Note that this method is the reverse of electrochemiluminescent detection of luminescent materials.

In subsequent reports, Weber et al. discussed the problems associated with the use of this method to detect Ru-containing labels (1). In Table 2 of Weber et al. (1), the extrapolated detection limit for tris(bipyridyl)ruthenium(II) is 1.1×10^{-10} moles/L under optimal conditions. In anticipating that the actual use of these labels would entail measurements in the presence of complex mixtures, Weber et al. tested for potential interferences in their system. Table 3 of Weber et al. lists dimethylalkyl amines, EDTA, N-methylmorpholine, N,N'-dimethylpiperazine, hydroxide, oxalate, ascorbate, uric acid, and serum as interferences which would presumably raise the practical detection limit substantially above 1.1×10^{-10} moles/L.

These studies were performed with a simple Ru-containing compound. No studies were reported in Weber or Weber et al. regarding the limits of detection of complex substances labelled with Ru-containing labels, or whether the thiourea linkage between the labeled material and label is stable under conditions of the assay.

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The particular labels with which the present invention is concerned are electrochemiluminescent. They can often be excited to a luminescent state without their oxidation or reduction by exposing the compounds to electromagnetic radiation or to a chemical energy source such as that created by typical oxalate-H₂O₂ systems. In addition, luminescence of these compounds can be induced by electrochemical methods which do entail their oxidation and reduction.

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Extensive work has been reported on methods for detecting Ru(2,2'-bipyridine)₃²⁺ using photoluminescent, chemiluminescent, and electrochemiluminescent means (2, 3). This work demonstrates that bright orange chemiluminescence can be based on the aqueous reaction of chemically generated or electrogenerated Ru(bpy)₃³⁺ (where "bpy" represents a bipyridyl ligand) with strong reductants produced as intermediates in the oxidation of oxalate ions or other organic acids. Luminescence also can be achieved in organic solvent-H₂O solutions by the reaction of electrogenerated, or chemically generated, Ru(bpy)₃¹⁺ with strong oxidants generated during reduction of peroxydisulfate. A third mechanism for production of electrochemiluminescence from Ru(bpy)₃²⁺ involves the oscillation of an electrode potential between a potential sufficiently negative to produce Ru(bpy)₃¹⁺ and sufficiently positive to produce

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$\text{Ru}(\text{bpy})_3^{3+}$. These three methods are called, respectively, "oxidative-reduction," "reductive-oxidation," and "the $\text{Ru}(\text{bpy})_3^{3+/+}$ regenerative system".

The oxidative-reduction method can be performed in water, and produces an intense, efficient, stable luminescence, which is relatively insensitive to the presence of oxygen or impurities. This luminescence from $\text{Ru}(\text{bpy})_3^{2+}$ depends upon the presence of oxalate or other organic acids such as pyruvate, lactate, malonate, tartrate and citrate, and means of oxidatively producing $\text{Ru}(\text{bpy})_3^{3+}$ species. This oxidation can be performed chemically by such strong oxidants as PbO_2 or a Ce(IV) salt. It can be performed electrochemically by a sufficiently positive potential applied either continuously or intermittently. Suitable electrodes for the electrochemical oxidation of $\text{Ru}(\text{bpy})_3^{2+}$ are, for example, Pt, pyrolytic graphite, and glassy carbon. Although the oxalate or other organic acid is consumed during chemiluminescence, a strong, constant chemiluminescence for many hours can be achieved by the presence of an excess of the consumed material, or by a continuous supply of the consumed material to the reaction chamber.

The reductive-oxidation method can be performed in partially aqueous solutions containing an organic co-solvent such as, for example, acetonitrile. This luminescence depends upon the presence of peroxydisulfate and a means of reductively producing $\text{Ru}(\text{bpy})_3^{1+}$ species. The reduction can be performed chemically by strong reductants such as, for example, magnesium or other metals. It can be performed electrochemically by a sufficiently negative potential applied either continuously or intermittently. A suitable electrode for

the electrochemical reduction of $\text{Ru}(\text{bpy})_3^{2+}$ is, for example, a polished glassy-carbon electrode. As with the oxidative-reduction method, continuous, intense luminescence can be achieved for many hours by inclusion of excess reagents, or by continuous addition of the consumed reagents to the reaction mixture.

5 The $\text{Ru}(\text{bpy})_3^{3+/+}$ regenerative system can be performed in organic solvents such as acetonitrile or in partially aqueous systems, by pulsing an electrode potential between a potential sufficiently negative to reduce $\text{Ru}(\text{bpy})_3^{2+}$ and a potential sufficiently positive to
10 oxidize $\text{Ru}(\text{bpy})_3^{2+}$. A suitable electrode for such a regenerative system is, for example, a Pt electrode. This system does not consume chemical reagents and can proceed, in principle, for an unlimited duration.

15 These three methods of producing luminescent Ru-containing compounds have in common the repetitive oxidation-reduction or reduction-oxidation of the Ru-containing compound. The luminescence of solutions containing these compounds is therefore highly dependent
20 on the electric potential of the applied energy source, and is therefore highly diagnostic of the presence of the Ru-containing compound.

Mandle cites Curtis et al. (4) as a possible label in
25 chemiluminescent applications. Curtis et al. reports only unpublished observations that Ru complexes can be induced to emit light when chemically excited by an oxalate/ H_2O_2 system (Curtis et al. p. 350).

30 Neither Mandle nor Curtis recognized the exceptional utility of ruthenium and osmium complexes in chemiluminescent applications or the utility of electrochemi-

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luminescent systems. Sprintschnik, G. et al. (5) have described complexes of tris (2,2'-bipyridine)ruthenium(II) esterified with octadecanol or dehydrocholesterol, and have created monolayer films of these surfactant complexes. The complexes were photoluminescent. But when the films were exposed to water, and then to light, the Ru-complexes failed to photoluminesce. This was attributed to photohydrolysis of ester groups in the presence of light.

It has been discovered, and is disclosed herein, that a wide variety of analytes of interest and chemical moieties that bind to analytes of interest may be conveniently attached to Ru-containing or Os-containing labels through amide or amine linkages. The labeled materials may then be determined by any of a wide variety of means, but by far the most efficient, reliable, and sensitive means are photoluminescent, chemiluminescent, and electrochemiluminescent means. It is also disclosed herein that electrochemiluminescent labels, including Ru-containing and Os-containing labels and organic molecules such as rubrene and 9,10-diphenyl anthracene, are particularly versatile and advantageous. The great advantages of the use of these novel labeled materials, and of the methods of detecting them, are further discussed hereinbelow.

For many years the food industry has been concerned with the presence of biological and chemical contaminants in raw food components and processed foods. While technological advances have been made in reducing the occurrence of food contamination and food borne disease outbreaks resulting therefrom, little progress has been reported in developing rapid and sensitive methods for the detection and identification

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of food contaminants. Existing standard methods for the detection of harmful contaminants in foods are generally very time consuming, labor intensive, and technically difficult. While the analytical methods themselves for the most part are of adequate sensitivity, the lengthy sample preparation procedures prior to the performance of the detection method often result in low yield of the contaminant in question so that false negatives are frequently encountered.

Two examples which serve to illustrate these problems are the currently recognized standard methods for detecting the presence of Salmonella and Staphylococcal enterotoxins in foods. The detection of Salmonella in foods involves several enrichment stages due to the fact that these bacteria, when present in foods, are usually found in low numbers and are often sublethally injured. Therefore, detection methods for Salmonella must be sensitive and allow for the resuscitation and growth of injured cells.

Two methods for Salmonella detection are currently recommended by the U.S. Food and Drug Administration. These methods appear in The Bacteriological Analytical Manual for Foods (1984), 6th ed., Association of Official Analytical Chemists, Washington, DC. One method is a pure culture technique involving preenrichment, selective enrichment and selective plating, a procedure which requires 4 days to obtain presumptive results and 5 to 7 days to obtain complete results. The other method involves immunofluorescence after selective enrichment. This procedure is more rapid, however it can result in a high incidence of false-positive results due to problems of cross-reactivity of the polyvalent antisera used in the test (6, 7).

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The procedure recommended by the U.S. Food and Drug Administration for the detection of Staphylococcal enterotoxins in foods also appears in The Bacteriological Analytical Manual for Foods (1984), 6th ed. Association of Official Analytical Chemists, Washington, DC. This method involves the concentration of an extract of a large food sample, e.g. approximately 100 grams, to a small volume, e.g. approximately 0.2 ml, by several dialysis concentration steps and an ion exchange column purification of the sample extract in order to prepare the sample for the microslide double-immunodiffusion technique. This procedure generally requires more than a week to perform.

Tests which are more rapid have recently been developed for the detection of a variety of contaminants such as bacteria, toxins, antibiotics and pesticide residues in foods. In many cases however, sample preparation prior to running the assay continues to be laborious and time consuming. Radioimmunoassays (RIA) and enzyme-linked immunosorbent assays (ELISA) have shortened the hands-on time for the analytical method itself, however these methods are still labor intensive and far from simple to perform. In addition, these methods are usually based on the use of polyclonal antisera, which is variable in specificity and sensitivity, and is generally in short supply for testing for a given food contaminant. ELISA methods have been developed for the analysis of food samples which employ monoclonal antibodies rather than polyclonal antisera. The use of monoclonal antibodies in an assay system for a food contaminant assures the constant supply of a reagent which imparts unchanging

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specificity and sensitivity to the test itself. Monoclonal antibodies have been used in ELISA systems to test for food contaminants such as *Salmonella* (8) and *Staphylococcal* enterotoxins (9). Commercially available products for *Salmonella* detection which employ EIA methodology (Bio-Enzabead Screen Kit, Litton Bionetics) and DNA probe technology (Gene-Trak, Integrated Genetics) are time consuming and labor intensive. Commercially available tests for detection of *Staphylococcal* enterotoxin in foods which employ reversed passive latex agglutination (SET-RPLA, Denka Seiken Co.) and EIA methodology (SET-EIA, Dr. W. Bommeli Laboratories) suffer from the same limitations.

For the past 100 years the bacterium *Escherichia coli* and the coliform group have been commonly used as indicators to monitor water quality and incidences of sewage contamination.

Current detection methodologies for *E. coli* and/or coliforms are based on the properties of acid or gas production from the fermentation of lactose. The most widely used methods are: the Most Probable Number (MPN) assay and the Membrane Filtration (MF) test. Both techniques are approved by the Environmental Protection Agency (EPA) and the American Public Health Association (APHA) for the microbiological examination of water and waste water (10), and also by the Food and Drug Administration (FDA) for the bacteriological examination of milk and foods (11).

The MPN method is actually comprised of three (12) separate assays (10). In the Presumptive test, a nonselective medium such as Lauryl Sulfate Tryptose (LST) broth or Lactose broth is used to check for gas

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production from the fermentation of lactose. Gas positive tubes are then subcultured into a more selective medium, Brilliant Green Lactose Bile (BGLB) broth for coliforms and E. coli (EC) broth for fecal coliforms, and again checked for gas production (confirmed test). Samples from positive Confirmatory tests are required to be tested further by plating on a selective and differential medium like Eosin Methylene Blue (EMB) agar or Endos agar, followed by Gram stain and some biochemical tests to firmly establish the presence of the indicator bacteria (Completed test). The entire MPN assay may require up to five (5) days for completion; therefore, for routine water analysis, most laboratories use only the Presumptive and the Confirmed portions of the MPN assay, which still requires 48 hours to 72 hours to complete. In addition to being time consuming and cost ineffective in terms of materials, incidences of both false positive and false negative reactions have also been commonly encountered in the MPN assays (15, 16, 20).

The MF technique for the bacteriological examination of water was introduced in the early 1950's (12). Unlike the MPN assay, which was tedious and time consuming, MF analysis could be completed in 24 hours without the need for further confirmations. The basic MF procedure is as follows: A volume of water sample, usually 100 ml is filtered through a 0.45 um pore diameter filter, and then incubated on a sterile pad saturated with selective medium. The two media most often used are the mEndo broth, selective for coliforms at 35°C, and the mFC broth, selective for fecal coliforms at 44.5°C (10). Since the introduction of the media, numerous authors have reported that both the mEndo and the mFC broths tend to underestimate the actual numbers of

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indicator bacteria present, due either to the selectivity of the medium or the high temperature used for incubation (44.5°C) (21, 22). Such incidences of false negatives have been especially prevalent when the organisms in the sample have been sublethally injured by environmental factors and/or chemicals (17, 18). Recently, modifications have been proposed by the EPA to follow up the MF test by a confirmatory procedure, whereby at least ten colonies on each filter need to be checked for gas production using the LST broth followed by BGLB broth as in the MPN assay (14). Such modifications although would reduce the incidences of both false negative and false positive reactions, it would also increase material cost as well as triple the MF assay time from 24 hours to 72 hours.

In 1982, Feng and Hartman introduced a fluorogenic assay for the detection of E. coli using the substrate 4-methyl umbelliferone glucuronide (MUG) (13). E. coli cells produced the enzyme beta-glucuronidase which would cleave the substrate releasing the fluorogenic 4-methylumbelliferone radical (19). By incorporating the compound MUG into the Presumptive LST medium, a single tube of LST-MUG medium provided both the Presumptive data (gas production) and the Confirmed data (fluorescence) for fecal coliforms within 24 hours. Although the MUG assay was rapid and simple, only 85% to 95% of the E. coli (depending on source) produced this enzyme, hence the test was not 100% reliable. Also the system was not applicable to the coliform group.

Currently, no suitable assay exists for the detection and enumeration of coliforms and fecal coliforms in a sample. The development of a simple, rapid, and reliable detection assay would not only decrease cost and

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time, but also greatly increase the efficiency of monitoring water sanitation and food processing and handling.

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SUMMARY OF THE INVENTION

The present invention provides a method of detecting in a predetermined volume of a multicomponent, liquid sample an analyte of interest present in the sample at a concentration below about 10^{-3} molar which comprises:

5 a) contacting a sample with a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation and (ii) capable of combining with the analyte of interest, the contact

10 being effected under appropriate conditions such that the analyte and the reagent combine; b) exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being

15 effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and c) detecting electromagnetic radiation so emitted and thereby detecting the presence of the analyte of interest in the sample.

20 The present invention also provides a competitive method for detecting in a predetermined volume of a multicomponent, liquid sample an analyte of interest present in the sample at a concentration below about

25 10^{-3} molar which comprises: a) contacting the sample with a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly

30 emit radiation and (ii) capable of competing with the analyte of interest for binding sites on a complementary material not normally present in the

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sample, and with the complementary material, the contact being effected under appropriate conditions such that the analyte of interest and the reagent competitively bind to the complementary material; b) exposing the resulting sample to an amount of electrochemical energy from a suitable source effected to induce the reagent to repeatedly emit radiation, the exposure being effective under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and c) detecting electromagnetic radiation so emitted and thereby detecting the presence of the analyte of interest in the sample.

Additionally provided is a method for quantitatively determining in a predetermined volume of a multicomponent, liquid sample, the amount of an analyte of interest present in the sample which comprises: a) contacting the sample with a known amount of a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation and (ii) capable of combining with the analyte of interest, the contact being effected under appropriate conditions such that the analyte and reagent combine; b) exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and c) quantitatively determining the amount of radiation so emitted and thereby quantitatively determining the amount of the analyte of interest present in the sample.

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The invention further provides a competitive method for quantitatively determining in a predetermined volume of a multicomponent, liquid sample the amount of an analyte of interest present in the sample. This method comprises: a) contacting the sample with a known amount of a reagent (i) capable of being induced to repeatedly
5 emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation and (ii) capable of competing with the analyte of interest for binding sites on a
10 complementary material not normally present in the sample, and with a known amount of the complementary material, the contact being effected under appropriate conditions such that the analyte of interest and the reagent competitively bind to the complementary
15 material; b) exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly
20 emit electromagnetic radiation; and c) quantitatively determining the amount of radiation so emitted and thereby quantitatively determining the amount of the analyte of interest present in the sample.

25 Also provided is a method for detecting and identifying the presence of a multiplicity of analytes of interest in a liquid food or food homogenate. This method comprises: a) immersing into the liquid food or food homogenate a portion of a diagnostic reagent holder
30 suitable for immersing into a liquid or solid suspension and having immobilized to it a multiplicity of reagents, each reagent being immobilized to the

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diagnostic reagent holder in distinct, identifiable regions and capable of forming a complex with a single analyte of interest so as to allow the formation of immobilized reagent-analyte of interest complexes; b) removing the diagnostic reagent holder from the liquid food or food homogenate; c) rinsing the diagnostic reagent holder with a suitable rinsing solution; d) immersing the portion of the diagnostic reagent holder which contains the immobilized reagent-analyte of interest complexes into a detection solution which contains at least one detection reagent capable of forming complexes with the immobilized reagent-analyte of interest complexes so as to allow the formation of immobilized reagent-analyte of interest detection reagent complexes; and e) detecting the presence on the identifiable regions of the diagnostic reagent holder to which reagents are immobilized of immobilized reagent-analyte of interest-detection reagent complexes, thereby detecting and identifying the presence of a multiplicity of analytes of interest in the liquid food or food homogenate.

The present invention also provides a kit useful for detecting and identifying the presence of a multiplicity of enterotoxins in a sample. This kit comprises: a) a diagnostic reagent holder provided with a handle connected to a surface area suitable for immersing into a liquid or solid suspension, said surface area having at least one nitrocellulose membrane attached to it, said nitrocellulose membrane having a multiplicity of monoclonal antibodies separately and distinctly immobilized to identifiable regions on it each immobilized monoclonal antibody being specific for an antigenic determinant on one enterotoxin; b) a first rinsing solution which

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comprises a buffered-aqueous solution containing a surfactant; c) a detection which comprises at least one monoclonal antibody enzyme conjugate, the monoclonal antibody of which is specific for antigenic determinants different from but located on each enterotoxin for which the monoclonal antibodies immobilize to the nitrocellulose membrane attached to the diagnostic reagent holder are specific; d) a second rinsing solution which comprises a buffered-aqueous solution; e) an enzyme substrate capable of reacting with the enzyme conjugated to the detection solution monoclonal antibody; and f) a colorless dye.

A method for detecting the presence of at least one species of bacteria in a sample is also provided by the present invention. This method comprises: a) innoculating the sample into at least one receptacle which is provided with an open end and which contains a suitable medium for supporting the growth of the species of bacteria; b) coupling a cap to the end of the receptacle, the face of the cap which when coupled to the receptacle, is exposed to the interior of the receptacle being provided with a surface suitable for immobilizing at least a component of the bacteria; c) incubating the innoculated media under conditions so as to allow bacteria innoculated into the media to reproduce; d) turning each receptacle upside-down for a suitable length of time so as to allow components of the bacterium present in the medium to become immobilized to the surface of the cap which is exposed to the interior of the receptacle; e) turning the upside-down receptacle rightside-up; f) uncoupling the cap from the receptacle; g) contacting the surface of the cap which has components of the bacteria immobilized to it with a reagent capable of forming a

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complex with the immobilized components of the bacteria under conditions so as to allow the formation of immobilized bacterial component-reagent complexes; and h) detecting immobilized bacterial component-reagent complexes, thereby detecting the presence of the species of bacteria within the sample.

A method for detecting the presence of coliform bacteria in a sample is further provided by the invention. This method comprises: a) inoculating the sample into at least one receptacle which has an open end and contains a non-selective lactose medium and an inverted Durham; b) coupling a cap provided with a polystyrene insert to the open end of each receptacle; c) incubating the inoculated medium for at least 2 hours at 37°C; d) determining whether each receptacle has gas produced by bacterial fermentation trapped within the inverted Durham vial; e) turning receptacles which have gas trapped within the inverted Durham vial upside-down for a suitable length of time so as to coliform bacteria present in the medium to form coliform-polystyrene complexes; f) turning the upside-down receptacles rightside-up; g) uncoupling the caps from the receptacles; h) treating the polystyrene inserts of the uncoupled caps with a substance suitable for blocking unbound sites; i) treating the polystyrene insert of the caps which had been treated with a substance suitable for blocking unbound sites with at least one anti-coliform bacteria antibody labeled with a detectable marker and capable of binding to coliform bacteria under conditions so as to allow the formation of antibody-coliform-polystyrene complexes; and j) detecting the presence of antibody-coliform-polystyrene complexes on the polystyrene insert, thereby detecting the presence of coliform bacteria in the sample.

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BRIEF DESCRIPTION OF FIGURES

5 Figure 1 depicts electrochemiluminescent measurements made for a homogeneous immunoassay for the determination of the concentration of an antigen in solution.

Figure 2 graphically depicts the results of a homogeneous ECL theophylline assay.

10 Figure 3 graphically depicts the results of a homogeneous theophylline assay in various sera.

- 15
- = Normal sera
 - = Hemolyzed sera
 - ◆ = Lipemic sera
 - ◻ = Icteric sera

20 Figure 4 graphically depicts the results of an ECL theophylline assay compared to the results of a fluorescence polarization theophylline assay.

- 25
- A. Normal sera: $n = 4$; slope = .986; $r = 1.00$.
 - B. Hemolyzed sera: $n = 3$; slope = .878; $r = 1.00$
 - C. Lipemic sera: $n = 5$; slope = .872; $r = 0.99$
 - 30 D. Icteric sera: $n = 4$; slope = 2.14; $r = 1.00$

35 Figure 5 graphically depicts the results of an ECL theophylline assay compared to the results from a high pressure liquid chromatography assay.

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$n = 9$; slope = 1.197; $r = 0.98$.

Figure 6 graphically depicts the modulation of an ECL signal generated in an ECL digoxin immunoassay.

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Figure 7 graphically depicts the results of an ECL digoxin immunoassay.

■ = Blank

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● = Digoxin

Figure 8 graphically depicts the ECL signal generated by various concentrations of MBI 38-Compound I.

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Figure 9 shows the results of a Hybridization/Sensitivity Study of MBI 38-Compound I.

TAG = Compound I

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Figure 10 shows the results of a Specificity Study of MBI 38-Compound I.

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DETAILED DESCRIPTION OF THE INVENTION

The present invention provides a method of detecting in a predetermined volume of a multicomponent, liquid sample an analyte of interest present in the sample at a concentration below about 10^{-3} molar which comprises:

5 a) contacting a sample with a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the

10 reagent to repeatedly emit radiation and (ii) capable of combining with the analyte of interest, the contact being effected under appropriate conditions such that the analyte and the reagent combine; b) exposing the

15 resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation;

20 and c) detecting electromagnetic radiation so emitted and thereby detecting the presence of the analyte of interest in the sample.

Within this application "molar" means the concentration of an analyte in solution in moles per liter or the

25 amount of particulate matter present in a liquid sample in particles or units per liter. For example, 1×10^{23} particles per liter may be expressed as 1 molar.

The methods provided by the present invention may be performed as heterogeneous assays, i.e. assays in which unbound labeled reagent is separated from bound labeled reagent prior to exposure of the bound labeled reagent to electrochemical energy, and homogeneous assays, i.e.

30 assays in which unbound labeled reagent and bound

35 labeled reagent are exposed to electrochemical energy

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together. In the homogeneous assays of the present invention the electromagnetic radiation emitted by the bound labeled reagent is distinguishable from the electromagnetic radiation emitted by the unbound labeled reagent, either as an increase or as a decrease in the amount of electromagnetic radiation emitted by the bound labeled reagent in comparison to the unbound labeled reagent, or as electromagnetic radiation of different wavelength. Accordingly, in one embodiment of the invention any reagent which is not combined with the analyte of interest is separated from the sample, which had been contacted with the reagent, prior to exposure of the sample to electrochemical energy. In another embodiment of the invention, prior to contacting the sample with the reagent, the sample is treated so as to immobilize the analyte of interest. Means for immobilizing analytes of interest are well known within the art and include contacting the sample with a polystyrene, nitrocellulose or nylon surface, or a surface coated with whole cells, subcellular particles, viruses, prions, viroids, lipids, fatty acids, nucleic acids, polysaccharides, proteins, lipoproteins, lipopolysaccharides, glycoproteins, peptides, cellular metabolites, hormones, pharmacological agents, tranquilizers, barbiturates, alkaloids, steroids, vitamins, amino acids, sugars, nonbiological polymers, synthetic organic molecules, organometallic molecules or inorganic molecules. Additionally, the analyte of interest may be any of these substances. In one embodiment of the invention, the analyte of interest is theophylline. In another embodiment of the invention, the analyte of interest is digoxin. In still another embodiment of the invention, the analyte of interest is human chorionic gonadotropin (hCG). Furthermore, the analyte of interest may be a

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whole cell, subcellular particle, virus, prion, viroid, nucleic acid, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, hormone, pharmacological agent, nonbiological polymer, synthetic organic molecule, organometallic molecule or an inorganic molecule present in the sample at a concentration below about 10^{-12} molar. Moreover the analyte of interest may be a whole cell, subcellular particle, virus, prion, viroid or nucleic acid present in the sample at a concentration below about 10^{-15} molar.

The reagent which is contacted with the sample may comprise an electrochemiluminescent chemical moiety conjugated to a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, nonbiological polymer, synthetic organic molecule, organometallic molecule, inorganic molecule, biotin, avidin or streptavidin. In one embodiment of the invention the agent is an electrochemiluminescent moiety conjugated to an antibody, antigen, nucleic acid, hapten, ligand or enzyme, or biotin avidin or streptavidin.

The electrochemiluminescent chemical moiety may comprise a metal-containing organic compound wherein the metal is selected from the group consisting of ruthenium, osmium, rhenium, iridium, rhodium, platinum, palladium, molybdenum and technetium. In one embodiment of the invention the metal is ruthenium or osmium. In another embodiment of the invention the electrochemiluminescent chemical moiety is rubrene or

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9, 10-diphenylanthracene. In still another embodiment of the invention, the electrochemiluminescent chemical moiety is bis[(4,4'-carbomethoxy)-2,2'-bipyridine] 2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-dioxolane ruthenium (II). In yet another embodiment of the invention, the electrochemiluminescent chemical moiety is bis (2,2'-bipyridine) [4-(butan-1-yl)-4'-methyl-2,2'-bipyridine] ruthenium (II). In a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis (2,2'-bipyridine) [4-(4'-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II). In still another embodiment of the invention, the electrochemiluminescent chemical moiety is (2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene]{2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane}osmium (II). In yet a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) [4-(4'-methyl-2,2'-bipyridine)-butylamine] ruthenium (II). In yet another embodiment of the invention the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[1-bromo-4(4'-methyl-2,2'-bipyridine-4-yl)butane] ruthenium (II). In still a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)maleimidoheptanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II).

The sample may be derived from a solid, emulsion, suspension, liquid or gas. Furthermore, the sample may be derived from water, food, blood, serum, urine, feces, tissue, saliva, oils, organic solvents or air. Moreover, the sample may comprise acetonitrile, dimethylsulfoxide, dimethylformamide, n-methylpyrrolidinone or tert-butyl alcohol. The sample may comprise a reducing agent or an oxidizing agent.

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5 The present invention also provides a competitive method for detecting in a predetermined volume of a multicomponent, liquid sample an analyte of interest present in the sample at a concentration below about 10^{-3} molar which comprises: a) contacting the sample with a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly
10 emit radiation and (ii) capable of competing with the analyte of interest for binding sites on a complementary material not normally present in the sample, and with the complementary material, the contact being effected under appropriate conditions such that the analyte of interest and the reagent competitively bind to the complementary material; b)
15 exposing the resulting sample to an amount of electrochemical energy from a suitable source effected to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and c) detecting electromagnetic radiation so emitted and thereby detecting the presence of the analyte of interest in
20 the sample.

25 The reagent may be the analyte of interest conjugated to an electrochemiluminescent chemical moiety or an analogue of the analyte of interest conjugated to an electrochemiluminescent moiety. Additionally, the analyte of interest may be theophylline, digoxin or hCG. Moreover, the electrochemiluminescent chemical moiety may be bis[(4,4'-carbomethoxy)-2,2'-bipyridine]
30 2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-

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dioxolane ruthenium (II). In yet another embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(butan-1-yl)-4'-methyl-2,2'-bipyridine] ruthenium (II). In a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II). In still another embodiment of the invention, the electrochemiluminescent chemical moiety is (2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene]{2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane}osmium (II). In yet a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(4'-methyl-2,2'-bipyridine)-butylamine], ruthenium (II). In yet another embodiment of the invention the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[1-bromo-4(4'-methyl-2,2'-bipyridine-4'-yl)butane] ruthenium (II). In still a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)maleimidoheptanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II).

The complementary material may be a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule.

It is within the scope of this application that the methods provided herein may be performed so as to

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quantify an analyte of interest. Accordingly the present invention provides a method for quantitatively determining in a predetermined volume of a multicomponent, liquid sample, the amount of an analyte of interest present in the sample which comprises: a) contacting the sample with a known amount of a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation and (ii) capable of combining with the analyte of interest, the contact being effected under appropriate conditions such that the analyte and reagent combine; b) exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and c) quantitatively determining the amount of radiation so emitted and thereby quantitatively determining the amount of the analyte of interest present in the sample.

This method may be performed as a heterogeneous assay or as a homogeneous assay. In one embodiment of the invention any reagent which is not combined with the analyte of interest is separated from the sample, which had been contacted with a known amount of the reagent, prior to the exposure of the sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation. In yet another embodiment of the invention, prior to contacting the sample with the reagent, the sample is treated so as to immobilize the analyte of interest.

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The analyte of interest may be a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule. In one embodiment of the invention, the analyte of interest is theophylline. In another embodiment of the invention, the analyte of interest is digoxin. In yet another embodiment of the invention, the analyte of interest is hCG.

The reagent with which the sample is contacted may be an electrochemiluminescent chemical moiety conjugated to a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule.

In one embodiment of the invention the reagent is a electrochemiluminescent chemical moiety conjugated to an antibody, antigen, nucleic acid, hapten, ligand or enzyme, or biotin, avidin or streptavidin.

The electrochemiluminescent moiety may be a metal-containing organic compound wherein the metal is selected from the group consisting of ruthenium, osmium, rhenium, iridium, rhodium, platinum, palladium,

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molybdenum and technetium. In one embodiment of the invention the metal is ruthenium or osmium. In another embodiment of the invention the electrochemiluminescent chemical moiety is rubrene or 9, 10-diphenylanthracene. In still a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis[(4,4'-carbomethoxy)-2,2'-bipyridine] 2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-dioxolane ruthenium (II). In yet another embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(butan-1-yl)-4'-methyl-2,2'-bipyridine] ruthenium (II). In a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) [4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II). In still another embodiment of the invention, the electrochemiluminescent chemical moiety is (2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene]{2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane}osmium (II). In yet a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) [4-(4'-methyl-2,2'-bipyridine)-butylamine] ruthenium (II). In yet another embodiment of the invention the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[1-bromo-4(4'-methyl-2,2'-bipyridine-4-yl)butane] ruthenium (II). In still a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)maleimido-hexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II).

The sample may be derived from a solid, emulsion, suspension, liquid or gas. Samples which comprise the analyte of interest may be derived from water, food, blood, serum, urine, feces, tissue, saliva, oils,

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organic solvents or air. Additionally, samples may comprise acetonitrile, dimethylsulfoxide, dimethylformamide, n-methylpyrrolidinone or tert-butyl alcohol. Furthermore, the sample may comprise a reducing agent or an oxidizing agent.

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The invention also provides a competitive method for quantitatively determining in a predetermined volume of a multicomponent, liquid sample the amount of an analyte of interest present in the sample. This method comprises: a) contacting the sample with a known amount of a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation and (ii) capable of competing with the analyte of interest for binding sites on a complementary material not normally present in the sample, and with a known amount of the complementary material, the contact being effected under appropriate conditions such that the analyte of interest and the reagent competitively bind to the complementary material; b) exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and c) quantitatively determining the amount of radiation so emitted and thereby quantitatively determining the amount of the analyte of interest present in the sample.

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The analyte of interest may be theophylline, digoxin or hCG.

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In one embodiment of the invention, the reagent is the analyte of interest conjugated to an electrochemiluminescent chemical moiety or an analogue of the analyte of interest conjugated to an electrochemiluminescent chemical moiety. The electrochemiluminescent chemical moiety may be bis[(4,4'-carbomethoxy)-2,2'-bipyridine] 2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-dioxolane ruthenium (II). In another embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(butan-1-yl)-4'-methyl-2,2'-bipyridine] ruthenium (II). In a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II). In yet a further embodiment of the invention, the electrochemiluminescent chemical moiety is (2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene]{2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane}osmium (II). In yet a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(4'-methyl-2,2'-bipyridine)butylamine] ruthenium II. In yet another embodiment of the invention the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[1-bromo-4(4'-methyl-2,2'-bipyridine-4-yl)butane] ruthenium (II). In still a further embodiment of the invention, the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)maleimido-hexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II).

The complementary material may be a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide,

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cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule.

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The present invention further provides a method for detecting and identifying the presence of a multiplicity of analytes of interest in a liquid food or food homogenate. This method comprises: a) 10 immersing into the liquid food or food homogenate a portion of a diagnostic reagent holder suitable for immersing into a liquid or solid suspension and having immobilized to it a multiplicity of reagents, each reagent being immobilized to the diagnostic reagent 15 holder in distinct, identifiable regions and capable of forming a complex with a single analyte of interest so as to allow the formation of immobilized reagent-analyte of interest complexes; b) removing the diagnostic reagent holder from the liquid food or food 20 homogenate; c) rinsing the diagnostic reagent holder with a suitable rinsing solution; d) immersing the portion of the diagnostic reagent holder which contains the immobilized reagent-analyte of interest complexes into a detection solution which contains at least one 25 detection reagent capable of forming complexes with the immobilized reagent-analyte of interest complexes so as to allow the formation of immobilized reagent-analyte of interest-detection reagent complexes; and e) detecting the presence on the identifiable regions of 30 the diagnostic reagent holder to which reagents are immobilized of immobilized reagent-analyte of interest-detection reagent complexes, thereby detecting and identifying the presence of a multiplicity of analytes of interest in the liquid food or food homogenate.

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The analytes of interest may be microorganisms. The microorganisms may be viable or nonviable. Additionally, the microorganisms may be bacteria. Examples of bacteria which may be detected by this method include, but are not limited to, Salmonella, Campylobacter, Escherichia, Yersinia, Bacillus, Vibrio, Legionella, Clostridium, Streptococcus or Staphylococcus.

Additionally, the analytes of interest may be antigens. Such antigens include, but are not limited to, enterotoxins and aflatoxins.

The immobilized reagents and the detection reagents may be polyclonal antibodies, monoclonal antibodies, mixtures of monoclonal antibodies, or mixtures of polyclonal and monoclonal antibodies.

The detection reagents may be labeled with a detectable marker. In one embodiment of the invention the detection reagents are each labeled with the same detectable marker. Such markers are well known in the art and may comprise an enzyme, e.g. alkaline phosphatase, horseradish peroxidase, glucose oxidase, beta-galactosidase or urease, a fluorescent moiety, e.g. fluorescein isothiocyanate, tetramethylrhodamine isothiocyanate, dichlorotriazinylamino fluorescein or Texas Red or a chemiluminescent moiety, e.g. luminol, isoluminol or an acridinium ester. Moreover, the detectable marker may be an electrochemiluminescent chemical moiety capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the moiety to emit radiation. This

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electrochemiluminescent chemical moiety may comprise ruthenium or osmium.

5 The immobilized reagents may be immobilized to a nitrocellulose membrane which is attached to the diagnostic reagent holder.

10 The present invention also provides a kit useful for detecting and identifying the presence of a multiplicity of enterotoxins in a sample. This kit comprises: a) a diagnostic reagent holder provided with a handle connected to a surface area suitable for immersing into a liquid or solid suspension, said surface area having at least one nitrocellulose membrane attached to it, said nitrocellulose membrane
15 having a multiplicity of monoclonal antibodies separately and distinctly immobilized to identifiable regions on it, each immobilized monoclonal antibody being specific for an antigenic determinant on one enterotoxin; b) a first rinsing solution which
20 comprises a buffered-aqueous solution containing a surfactant; c) a detection which comprises at least one monoclonal antibody enzyme conjugate, the monoclonal antibody of which is specific for antigenic determinants different from but located on each
25 enterotoxin for which the monoclonal antibodies immobilize to the nitrocellulose membrane attached to the diagnostic reagent holder are specific; d) a second rinsing solution which comprises a buffered-aqueous solution; e) an enzyme substrate capable of reacting
30 with the enzyme conjugated to the detection solution monoclonal antibody; and f) a colorless dye.

35 In one embodiment of the invention the nitrocellulose membrane is provided with monoclonal antibodies which

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are separately and distinctly immobilized to it and which are specific for antigenic determinants on a multiplicity of Staphylococcal enterotoxins. In another embodiment of the invention the nitrocellulose membrane is provided with four monoclonal antibodies which are separately and distinctly immobilized to it and which are specific for Staphylococcal enterotoxins A, B, D and E. Furthermore, this nitrocellulose membrane may be additionally provided with a monoclonal antibody which is separately and distinctly immobilized to it and which is specific for an antigenic determinant on more than one Staphylococcal enterotoxin. This enterotoxin may be specific for antigenic determinants on Staphylococcal enterotoxins C₁, C₂ and C₃. The enzyme of this kit which is conjugated to the monoclonal antibody which is specific for antigenic determinant different from but located on each Staphylococcal enterotoxin for which the nitrocellulose immobilized monoclonal antibodies are specific may be alkaline phosphatase, the enzyme substrate which is capable of reacting with the enzyme is 5-bromo, 4-chloro indolyl phosphate and the colorless dye is nitroblue tetrazolium.

The invention further provides a method for detecting and identifying the presence of a multiplicity of Staphylococcal enterotoxins in a liquid or solid suspension. This method comprises: a) immersing a diagnostic reagent holder provided with a multiplicity of monoclonal antibodies specific for Staphylococcal enterotoxins and which are separately and distinctly immobilized to it into the liquid or solid suspension for a suitable length of time so as to allow the formation of immobilized monoclonal antibody-Staphylococcal enterotoxin complexes; b) removing the

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diagnostic reagent holder from the sample; c) immersing the diagnostic holder into a buffered-aqueous solution which comprises a surfactant; d) removing the diagnostic reagent holder from the buffered-aqueous solution which contains a surfactant; e) immersing the diagnostic reagent holder into a detection solution which comprises a monoclonal antibody-alkaline phosphatase conjugate for a suitable length of time so as to allow the formation of immobilized monoclonal antibody-Staphylococcal enterotoxin-monoclonal antibody-alkaline phosphatase complexes; f) removing the diagnostic reagent holder from the detection solution; g) immersing the diagnostic reagent holder into the a buffered-aqueous solution; h) removing the diagnostic reagent holder from the buffered-aqueous solution; i) immersing the diagnostic reagent holder into a solution which comprises 5-bromo, 4-chloro indolyl phosphate and nitroblue tetrazolium; j) visibly detecting the identifiable regions of the nitrocellulose membrane onto which a blue precipitate accumulates; k) correlating the identifiable regions of the nitrocellulose membrane onto which a blue percipitate accumulates with the Staphylococcal enterotoxin for which the monoclonal antibody immobilized to the region is specific, thereby indicating the presence and the identity of a multiplicity of Staphylococcal enterotoxins in the sample.

Also provided is a method for detecting the presence of at least one species of bacteria in a sample. This method comprises: a) inoculating the sample into at least one receptacle which is provided with an open end and which contains a suitable medium for supporting the growth of the species of bacteria; b) coupling a cap to the end of the receptacle, the face of the cap which

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when coupled to the receptacle, is exposed to the interior of the receptacle being provided with a surface suitable for immobilizing at least a component of the bacteria; c) incubating the inoculated media under conditions so as to allow bacteria inoculated into the media to reproduce; d) turning each receptacle upside-down for a suitable length of time so as to allow components of the bacterium present in the medium to become immobilized to the surface of the cap which is exposed to the interior of the receptacle; e) turning the upside-down receptacle rightside-up; f) uncoupling the cap from the receptacle; g) contacting the surface of the cap which has components of the bacteria immobilized to it with a reagent capable of forming a complex with the immobilized components of the bacteria under conditions so as to allow the formation of immobilized bacterial component-reagent complexes; and h) detecting immobilized bacterial component-reagent complexes, thereby detecting the presence of the species of bacteria within the sample.

Within this application "component of bacteria" includes, but is not limited to, a whole cell, cell wall component, cell membrane component and flagellar component.

The surface of the cap which is suitable for immobilizing at least a component of the bacteria may be a polystyrene insert, a nitrocellulose membrane or a nylon membrane. Furthermore, the surface may be coated with a polyclonal antibody, a monoclonal antibody, a mixture of monoclonal antibodies or a mixture of polyclonal and monoclonal antibodies.

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The reagent capable of forming a complex with the immobilized bacterial components may be a polyclonal antibody, a monoclonal antibody, a mixture of monoclonal antibodies or a mixture of polyclonal and monoclonal antibodies. Furthermore, the reagent may be labeled with a detectable marker, for example a detectable enzyme, a fluorescent moiety or a chemiluminescent moiety. Additionally, the detectable marker may be an electrochemiluminescent moiety. In one embodiment of the invention, the electrochemiluminescent moiety comprises ruthenium or osmium.

In yet another embodiment of the invention, the immobilized bacterial component-reagent complexes may be detected with a detectably marked second reagent capable of forming a complex with the immobilized bacterial component-reagent complexes. In still another embodiment of the invention the reagent may be a polyclonal antibody, a monoclonal antibody, a mixture of monoclonal antibodies or a mixture of polyclonal and monoclonal antibodies and the second reagent may be a detectably marked anti-antibody directed to the reagent.

Samples in which bacteria may be detected include water and food.

In one embodiment of the invention the medium suitable for supporting the growth of the species of bacteria is a non-selective lactose medium. In one embodiment of the invention the medium is a nonselective lactose medium and the species of bacteria which is detected is at least one coliform.

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A method for detecting the presence of coliform bacteria in a sample is also provided by the invention. This method comprises: a) inoculating the sample into at least one receptacle which has an open end and contains a non-selective lactose medium and an inverted Durham; b) coupling a cap provided with a polystyrene insert to the open end of each receptacle; c) incubating the inoculated medium for at least 2 hours at 37°C; d) determining whether each receptacle has gas produced by bacterial fermentation trapped within the inverted Durham vial; e) turning receptacles which have gas trapped within the inverted Durham vial upside-down for a suitable length of time so as to allow coliform bacteria present in the medium to form coliform-polystyrene complexes; f) turning the upside-down receptacles rightside-up; g) uncoupling the caps from the receptacles; h) treating the polystyrene inserts of the uncoupled caps with a substance suitable for blocking unbound sites; i) treating the polystyrene insert of the caps which had been treated with a substance suitable for blocking unbound sites with at least one anti-coliform bacteria antibody labeled with a detectable marker and capable of binding to coliform bacteria under conditions so as to allow the formation of antibody-coliform-polystyrene complexes; and j) detecting the presence of antibody-coliform-polystyrene complexes on the polystyrene insert, thereby detecting the presence of coliform bacteria in the sample.

In one embodiment of the invention the non-selective lactose medium is phenol red broth.

The substance suitable for blocking unbound sites on the polystyrene insert may be bovine serum albumin.

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The anti-coliform bacteria antibody labeled with a detectable marker may be a monoclonal antibody labeled with a detectable marker. The detectable marker may be an enzyme conjugated to the monoclonal antibody. In one embodiment of the invention the enzyme is calf intestinal alkaline phosphatase. In another embodiment of the invention the detectable marker is a fluorescent moiety conjugated to the monoclonal antibody. In still another embodiment of the invention the detectable marker is an electrochemiluminescent moiety conjugated to the monoclonal antibody. The electrochemiluminescent moiety may comprise ruthenium or osmium.

In one embodiment of the invention the sample is water. In another embodiment of the invention the sample is food.

The receptacle which has an open end may be a vial.

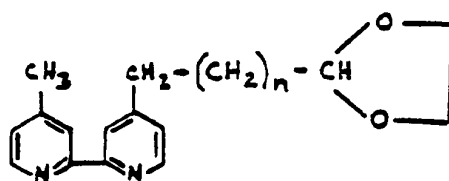
Also provided is a kit for detecting in water samples the presence of coliform bacteria. This kit comprises: a) at least one vial containing a non-selective lactose medium; b) at least one Durham vial; c) at least one vial cap provided with a polystyrene insert; d) a solution of bovine serum albumin; e) an anti-coliform bacteria monoclonal antibody-enzyme conjugate; f) a buffered-aqueous wash solution; and g) a solution of enzyme substrate. In one embodiment of the invention the enzyme conjugated to the anti-coliform bacteria monoclonal antibody is calf intestinal alkaline phosphatase and the enzyme substrate is p-nitrophenyl phosphate disodium.

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The invention also provides a compound having the structure

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wherein n is an integer. In one embodiment of the invention, n is 2.

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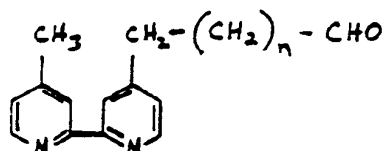
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-46-

Further, the invention provides a compound having the structure

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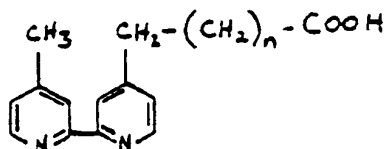
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wherein n is an integer. In one embodiment of the invention, n is 2.

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The invention also provides a compound having the structure

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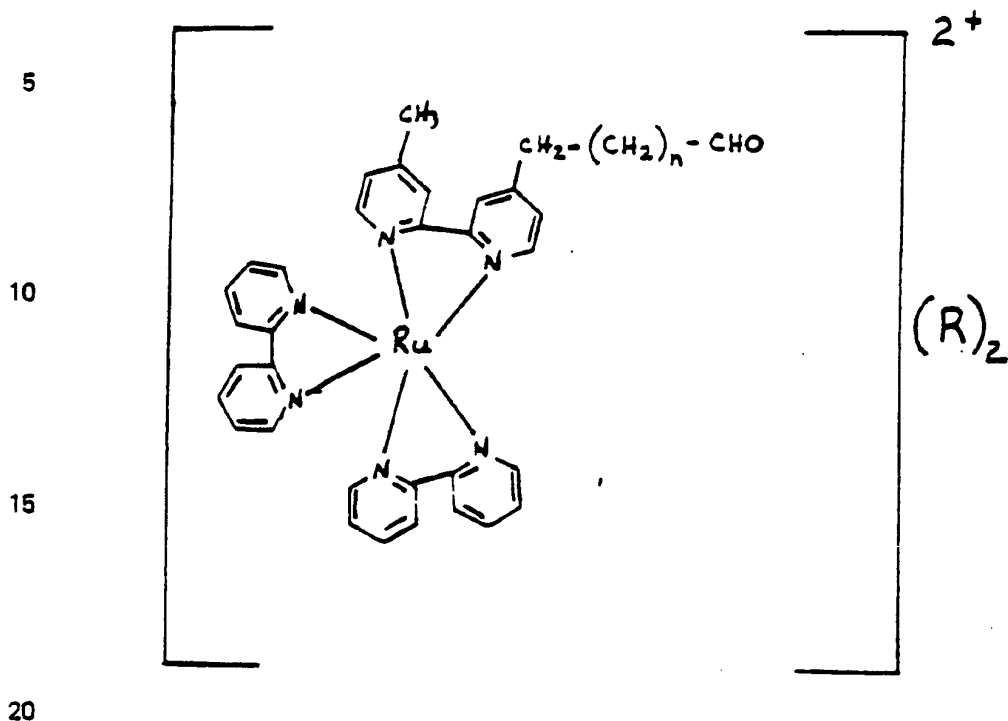
wherein n is an integer. In one embodiment of the invention, n is 2.

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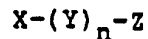
-47-

Also, the invention provides a compound having the structure



wherein R is an anion and n is an integer. In one embodiment of the invention, n is 2.

25 This compound may comprise a composition of matter having the structure

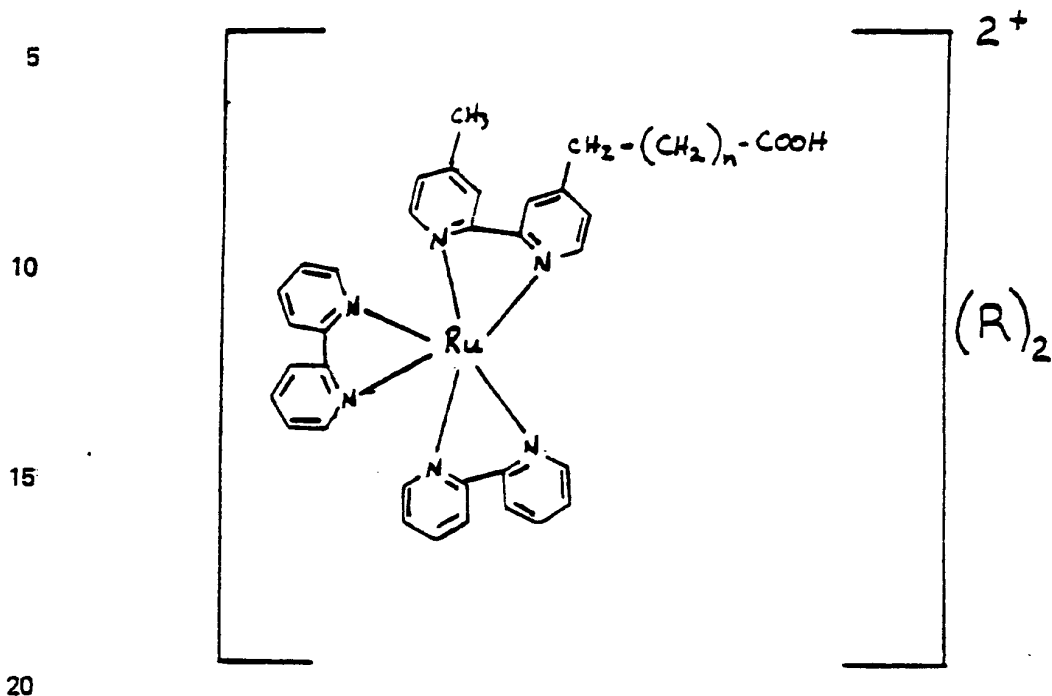


30 wherein X represents one or more nucleotides which may be the same or different, one or more amino acids which may be the same or different, an antibody, an analyte of interest or an analogue of an analyte of interest, n represents an integer, and Z represents the compound

35 provided by this invention.

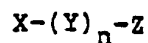
-48-

The invention further provides a compound having the structure



wherein R is an anion and n is an integer. In one embodiment of the invention, n is 2.

25 This compound may comprise a composition of matter having the structure

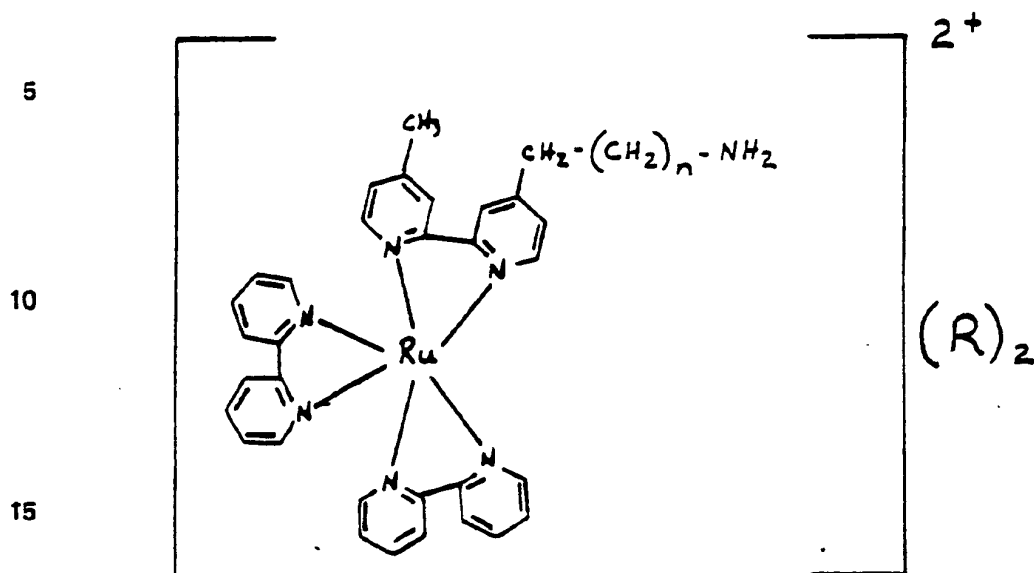


30 wherein X represents one or more nucleotides which may be the same or different, one or more amino acids which may be the same or different, an antibody, an analyte of interest or an analogue of an analyte of interest, n represents an integer, and Z represents the compound provided by this invention.

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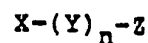
-49-

Also provided is a compound having the structure



wherein R is an anion and n is an integer. In one embodiment of the invention, n is 3.

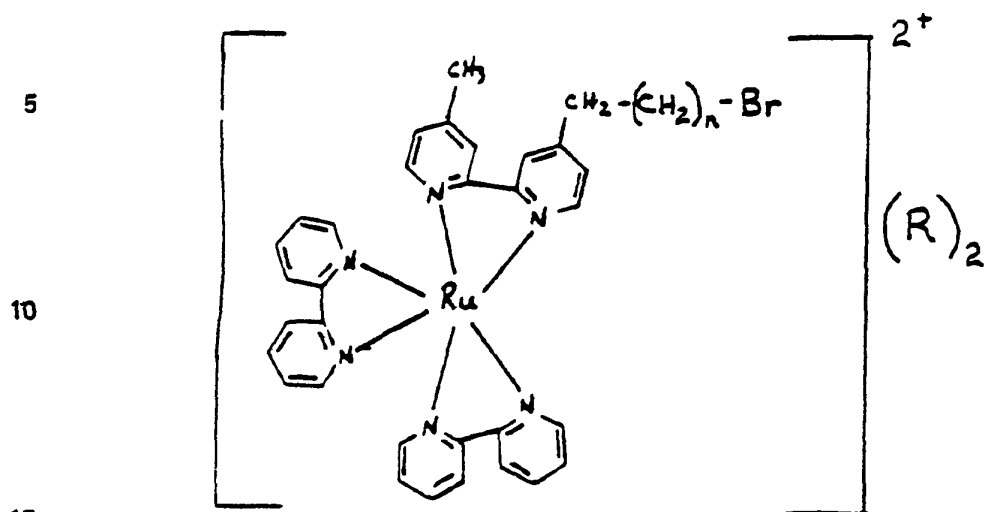
This compound may comprise a composition of matter having the structure



wherein X represents one or more nucleotides which may be the same or different, one or more amino acids which may be the same or different, an antibody, an analyte of interest or an analogue of an analyte of interest, n represents an integer, and Z represents the compound provided by this invention.

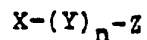
-50-

The invention further provides a compound having the structure



wherein R is an anion and n is an integer. In one embodiment of the invention, n is 3.

20 This compound may comprise a composition of matter having the structure



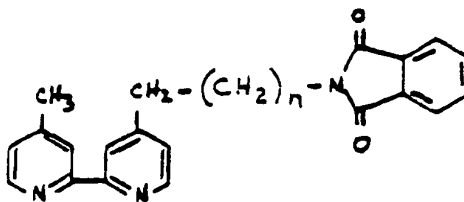
25 wherein X represents one or more nucleotides which may be the same or different, one or more nucleotides which may be the same or different, an antibody, an analyte of interest or an analogue of an analyte of interest, n represents an integer, and Z represents the compound

30 provided by this invention.

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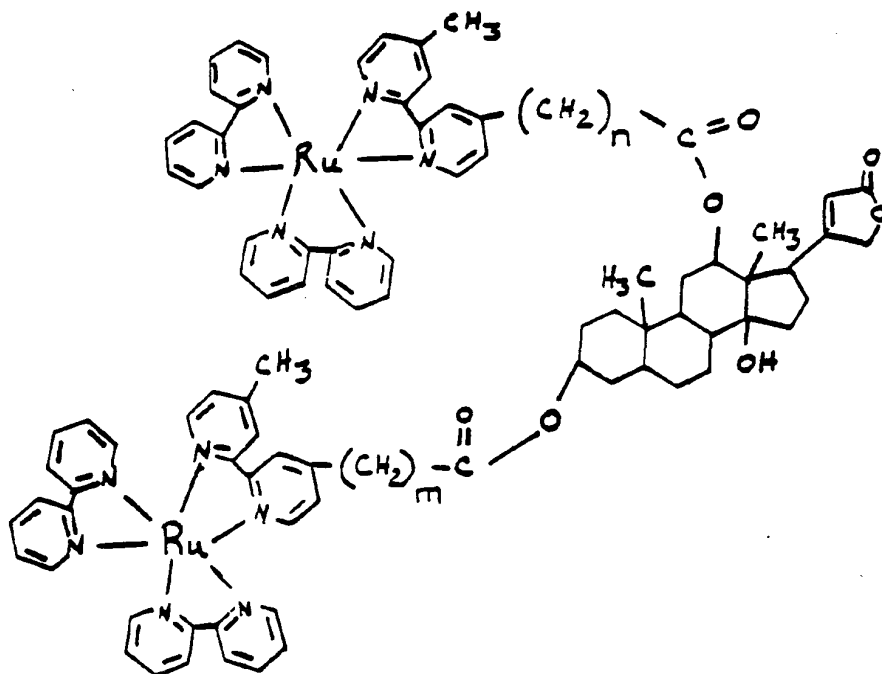
-51-

The invention also provides a compound having the structure



wherein n is an integer. In one embodiment of the invention, n is 3.

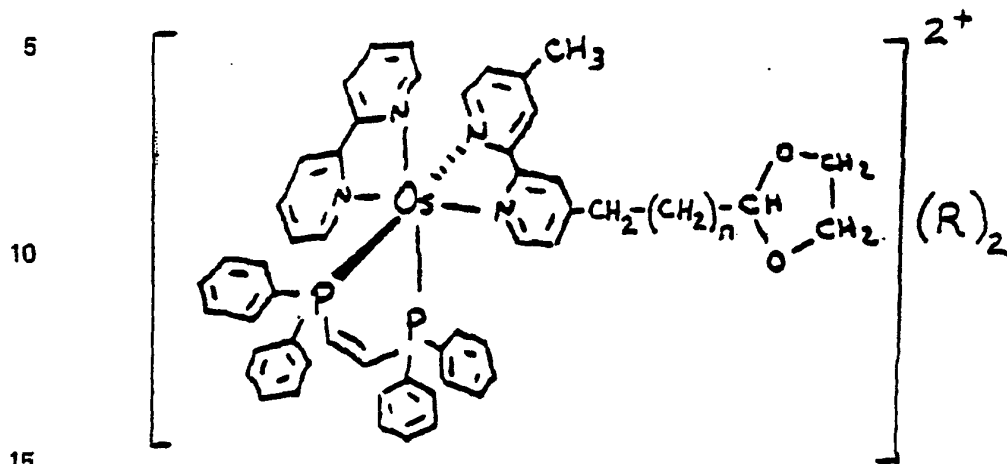
Also, the invention provides a compound having the structure



wherein m and n are each integers which may be the same or different. In one embodiment of the invention, m and n are both 3.

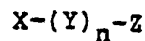
-52-

Further the invention provides a compound having the structure



wherein R is an anion and n is an integer. In one embodiment of the invention, n is 2.

20 This compound may comprise a composition of matter having the structure



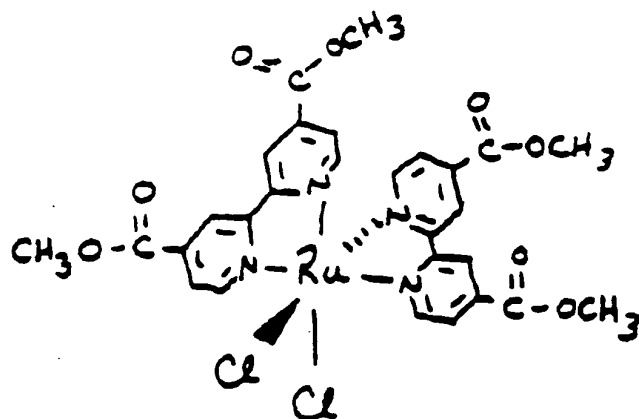
25 wherein X represents one or more nucleotides which may be the same or different, one or more amino acids which may be the same or different, an antibody, an analyte of interest or an analogue of an analyte of interest, n represents an integer, and Z represents the compound

30 provided by this invention.

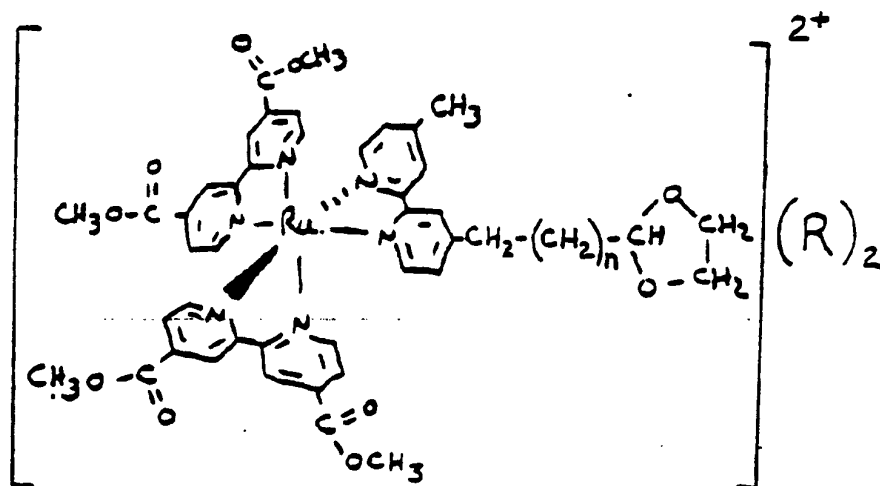
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The invention further provides a compound having the structure



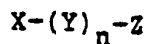
Also, the invention provides a compound having the structure



wherein R is an anion and n is an integer. In one embodiment of the invention, n is 2.

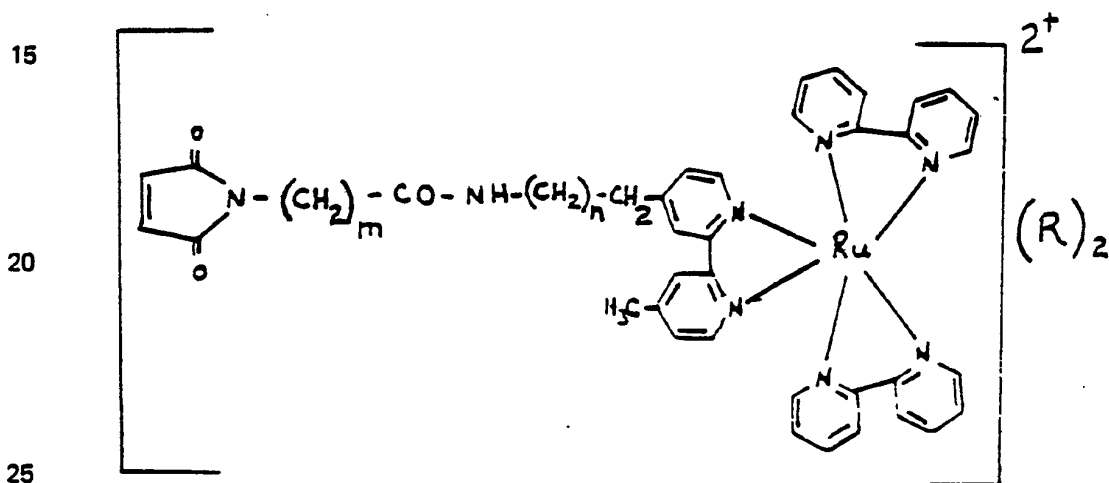
-54-

This compound may comprise a composition of matter having the structure



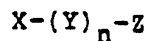
wherein X represents one or more nucleotides which may be the same or different, one or more amino acids which may be the same or different, an antibody, an analyte of interest or an analogue of an analyte of interest, n represents an integer, and Z represents the compound provided by this invention.

The invention further provides a compound having the structure



wherein R is an anion and m and n are integers. In one embodiment of the invention, m is 5 and n is 3.

This compound may comprise a composition of matter having the structure

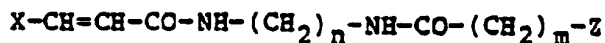


wherein X represents one or more nucleotides which may be the same or different, one or more amino acids which

-55-

may be the same or different, an antibody, an analyte of interest or an analogue of an analyte of interest, n represents an integer, and Z represents the compound provided by this invention. In one embodiment of the invention, X is theophylline. In another embodiment of the invention, X is digoxigenin. In a further embodiment of the invention, X is a peptide derived from hCG.

Also provided by the invention is a composition of matter having the structure



wherein:

X represents one or more nucleotides which may be the same or different;

Z represents an electrochemiluminescent chemical moiety;

n represents an integer greater than or equal to 1; and

m represents an integer greater than or equal to 1.

In one embodiment of the invention, X is thymidine attached to CH at carbon 5, n is 7 and m is 3.

In another embodiment of the invention Z is bis (2,2' - bipyridine) [4 -(butan-1-yl)-4' methyl-2,2'-bipyridine] ruthenium (II).

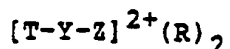
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In yet another embodiment of the invention, the thymidine nucleotide is a 3' terminal nucleotide attached to the nucleotide sequence

TCACCAATAAACOGCAAACACCATCCCGTCCTGCCAG

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Also provided is a composition of matter having the structure



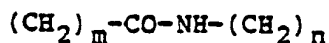
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wherein T represents theophylline, Y represents a linker group attaching T to Z, Z represents bis-(2,2'-bipyridine) [4-methyl-2,2'-bipyridine-4'-yl] ruthenium (II) and R represents an anion.

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In one embodiment of the invention, Y is attached to the carbon at position 8 of T. In another embodiment of the invention, Y has the structure

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wherein m and n represent an integer, which may be the same or different, greater than or equal to 1. In another embodiment of the invention, m is 3 and n is 4. In another embodiment of the invention, m and n are both 3.

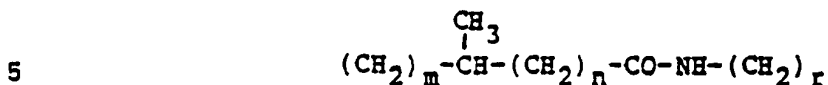
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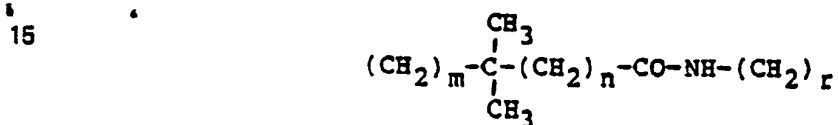
In yet another embodiment of the invention, Y has the structure



wherein m, n and r represent an integer, which may be the same or different, greater than or equal to 1. In one embodiment of the invention, m is 1, n is 1 and r is 4.

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In still a further embodiment of the invention, Y has the structure

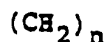


wherein m, n and r represent an integer, which may be the same or different, greater than or equal to 1. In one embodiment of the invention, m is 1, n is 1 and r is 4.

20

In yet another embodiment of the invention, Y is attached to the nitrogen at position 7 of T. In one embodiment of the invention, Y has the structure

25



wherein n is an integer greater than or equal to 1. In still another embodiment of the invention, n is 4.

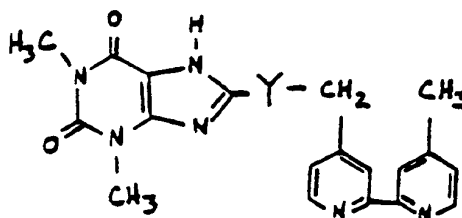
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The invention further provides a compound having the structure

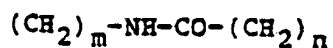
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wherein Y is a linker arm. In one embodiment of the invention, Y has the structure

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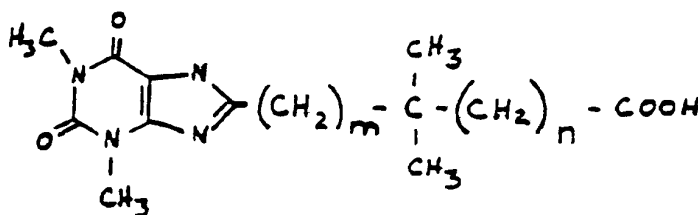


wherein m and n are integers, which may be the same or different, greater than or equal to 1. In one embodiment of the invention, m is 3 and n is 2.

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The invention also provides a compound having the structure

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-59-

wherein m and n are integers which may be the same or different. In one embodiment of the invention, m and n are both 1.

Further provided is a composition of matter having the structure

X-Z

wherein X represents one or more amino acids which may be the same or different, comprising at least one amino acid which is cysteine or methionine, and Z is bis (2,2'-bipyridine) maleimido hexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II) attached by the carbon at position 3 or 4 of the maleimide to a sulfur substituent of cysteine or methionine.

The following examples are provided to illustrate, but in no way limit, the present invention, which is defined by the claims which follow the specification.

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Example 1 - Electrochemiluminescence in Various Organic Solvents

5 The electrochemiluminescence of tris (2,2'-bipyridyl) ruthenium (II) chloride hexahydrate was measured in a 15 ml three-neck, round bottom flask containing 10 ml of a solution prepared as described below; a 1.5mm x 10 mm magnetic stir bar; a 1.0mm diameter silver wire quasi-reference electrode; a combination 28 gauge 10~ platinum wire counter electrode; and a working electrode consisting of a 22 gauge platinum wire welded to a 1 cm x 1 cm square piece of 0.1 mm thick, highly polished platinum foil. (The working platinum foil electrode was shaped into a 3/16 of an inch diameter 15~ semi-circle surrounding the 28 gauge platinum wire counter electrode by 3/32 of an inch equidistantly.)

20 The silver wire was connected to the EG&G Model 178 electrometer probe of the EG&G Model 173 potentiostat/galvanostat. The platinum wire counter electrode and the platinum working electrode were connected to the anode and cathode respectively of the EG&G Model 173 potentiostat. The device was grounded.

25~ Cyclic voltammetry was performed with the EG&G Model 173 potentiostat to which an EG&G Model 175 universal programmer was attached. The programmer was set for 30 100 mV/second sweeps between +1.75 volt anodic and -1.80 volt cathodic potentials. Electrochemiluminescence was detected using a Hamamatsu R928 photomultiplier tube, set inside a Products for Research Model PR1402RF photomultiplier tube housing which was fitted with a Kodak #23A gelatin (red) filter. The photo-

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multiplier tube housing was connected to an Oriel Model 7070 photomultiplier detection system. The cyclic voltammogram was recorded on a Houston Instruments Model 200 X-Y recorder.

5 Cyclic voltammograms were generated for 1mM tris (2, 2' - bipyridyl) ruthenium (II) chloride hexahydrate (Aldrich Chemical Company), 0.1M tetrabutylammonium tetrafluoroborate (TBABF₄) (Aldrich Chemical Company) solutions prepared with the following organic
10 solvents: acetonitrile; n-dimethylformamide; dimethylsulfoxide and 1-methyl, 2-pyrrolidinone (Aldrich Chemical Company). Tert-butyl alcohol and deionized, distilled water (1:1, v/v) also was used to make a solution containing 1 mM tris (2, 2'-bipyridyl)
15 ruthenium II chloride hexahydrate and 0.1 M TBABF₄. The resulting voltammograms did not indicate any change in the redox potential of the tris (2,2'-bipyridyl) ruthenium (II) chloride hexahydrate upon variation of the organic solvent.

20 For visual determination of electrochemiluminescence, solutions were prepared as follows: sufficient amounts of tris (2,2'-bipyridyl) ruthenium (II) chloride hexahydrate and TBABF₄ were dissolved in the spectroscopic grade organic solvents (Aldrich Chemical
25 Company) described above to provide final concentrations of 1mM and 0.1M, respectively. 10ml of the resulting solution was then added to the 15 ml three-neck round bottom flask. The electrodes were immersed in the solution and the working electrode
30 pulsed between a +1.75 and -1.45 volt potential to generate electrochemiluminescence. Electrochemiluminescence was visually observed in each of the solutions described above.

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For quantitative measurements of the effect of solvent variation on electrochemiluminescence, solutions were prepared as follows: sufficient amounts of tris (2, 2'-bipyridyl) ruthenium (II) chloride hexahydrate and TBABF₄ were added to the organic solvents described above to provide final concentrations of 2 mM and 0.2 M respectively. To an aliquot of this solution was added an equal volume of deionized, distilled water containing a strong oxidizing ammonium persulfate, at a concentration of 36 mM. Control solutions that did not contain the tris (2, 2'-bipyridyl) ruthenium (II) chloride hexahydrate were prepared. 10ml of the resulting solution was then added to the 15 ml three-neck round bottom flask. Electrochemiluminescence was accomplished by pulsing for one second intervals, between zero and -2.0 volts cathodic potential.

Electrochemiluminescent measurements were performed by integrating the resulting electrochemiluminescent photomultiplier tube signal using an. integrator connected to a Micronta Model 22191 digital multimeter. The electrochemiluminescent signal was integrated for 10 seconds during the pulsing and recorded in millivolts. The results are shown in Table I and indicate that variation of solvents effects quantum efficiency of the ruthenium (II) chloride.

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TABLE I

	<u>Organic Solvent</u>	<u>Tris RuBiPy (10⁻⁶M)</u>	<u>Control</u>	<u>Δ</u>
5	Acetonitrile	2,540*	104	2,436
	tert-butyl alcohol	1,280	0	1,280
	N, N dimethyl- formamide	2,390	143	2,247
10	Dimethylsulfoxide	2,760	29	2,731
	1-methyl-2- pyrrolidinone	1,630	0	1,630

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*all measurements in millivolts.

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Example 2 - Sensitivity of Detection of
Electrochemiluminescence of Ruthenium- Labeled Rabbit
Anti-Mouse Immunoglobulin G (IgG) Antibody

5 The electrochemiluminescence of rabbit anti-mouse IgG
antibody labeled with 4,4' - (dichloromethyl) - 2,2' -
bipyridyl, bis(2,2' - bipyridyl) ruthenium (II) (ruthen-
10 nium-labeled rabbit anti-mouse IgG antibody) was mea-
sured in a 15 ml three-neck, round bottom flask con-
taining 10 ml of a solution prepared as described
below; a 1.5mm x 10mm magnetic stir bar; a 1.0 mm diam-
eter silver wire quasi-reference electrode; a combina-
15 tion 28 gauge platinum wire counter electrode, and a
working electrode consisting of a 22 gauge platinum
wire welded to a 1 cm x 1 cm square piece of 0.1 mm
thick, highly polished platinum foil. (The working
platinum foil electrode was shaped into a 3/16 of an
20 inch diameter semi-circle surrounding the 28 gauge
platinum wire counter electrode by 3/32 of an inch
equidistantly.)

The silver wire was connected to the EG&G Model 178
electrometer probe of the EG&G Model 173 potent-
25 iostat/galvanostat. The platinum wire counter electrode
and the platinum working electrode were connected to
the anode and cathode respectively of the EG&G Model
173 potentiostat. The device was grounded.

The electrochemiluminescence emitted from the rutheni-
30 um-labeled rabbit anti-mouse IgG antibody solution was
detected using an Hamamatsu R928 photomultiplier tube,
set inside a Products for Research Model PRL402RF
photomultiplier tube housing which was fitted with a
Kodak #23A gelatin (red) filter. The photomultiplier

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-65-

tube housing was connected to an Oriel Model 7070 photomultiplier detection system.

Electrochemiluminescence was induced by pulsing for one second intervals, between zero and -2.0 volts cathodic potential. Electrochemiluminescent measurements were performed by integrating the resulting electrochemiluminescent photomultiplier tube signal using an integrator connected to a Micronta Model 22191 digital multimeter. The electrochemiluminescent signal was integrated for 10 seconds during the pulsing and recorded in millivolts.

A stock solution of $1.25 \times 10^{-7} \text{M}$ ruthenium-labeled rabbit anti-mouse IgG antibody was prepared from a concentrated solution (2 mg/ml, 7.5 Ru/antibody) of the labeled antibody by dilution in phosphate-buffered saline (PBS). An aliquot of this solution (80 microliters) was added to 10 ml of dimethylsulfoxide (DMSO)/deionized, distilled water (1:1) containing 0.1 M tetrabutylammonium tetrafluoroborate (TBABF_4) and 18 mM ammonium persulfate in the reaction vessel. The final ruthenium-labeled antibody concentration was $1 \times 10^{-9} \text{M}$. Electrochemiluminescence was measured as described above.

Additional solutions representing various dilutions of the ruthenium-labeled rabbit anti-mouse IgG antibody stock solution were made and aliquots (80 microliters) of these solutions were added to the same solution of ruthenium-labeled antibody in the reaction vessel in increments which resulted in the following concentrations of labeled antibody: $5 \times 10^{-9} \text{M}$, $1 \times 10^{-8} \text{M}$, and $5 \times 10^{-8} \text{M}$. Electrochemiluminescence measurements were made for each solution as described. These measure-

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-66-

ments are listed in Table II below. These results indicate the sensitivity of electrochemiluminescent detection of labeled antibody ($1 \times 10^{-9} \text{M}$), and the dependence of the intensity of electrochemiluminescence on the concentration of the ruthenium-labeled anti-mouse IgG antibody.

TABLE II

ELECTROCHEMILUMINESCENCE (ECL) OF RUTHENIUM-LABELED
RABBIT ANTI-MOUSE IMMUNOGLOBULIN G (IgG) ANTIBODY

10	Concentration of Ruthenium-Labeled <u>Anti-Mouse IgG Antibody</u>	<u>ECL (mV)</u>
	$5 \times 10^{-8} \text{M}$	1610
	$1 \times 10^{-8} \text{M}$	892
15	$5 \times 10^{-9} \text{M}$	418
	$1 \times 10^{-9} \text{M}$	72
	0	0

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Example 3 - Immunological Reactivity of Ruthenium-Labeled Bovine Serum Albumin (BSA) In a Solid Phase Enzyme Linked-Immunosorbent Assay (ELISA)

5 The wells of a polystyrene microtiter plate were coated with a saturating concentration of either bovine serum albumin labeled with 4,4' - (dichloromethyl) - 2,2' -
10 bipyridyl, bis(2,2' - bipyridyl) ruthenium (II), i.e. ruthenium-labeled bovine serum albumin, (6 Ru/BSA, 20 micrograms/ml in PBS buffer, 50 microliters/well) or
unlabeled BSA (20 micrograms/ml in PBS buffer, 50 microliters/well) and incubated for one hour at room temperature. After this incubation period the plate
15 was washed three times with PBS, 5 minutes per wash. A solution containing 6 mg/ml rabbit anti-BSA antibody was diluted 1:20,000, 1:30,000, 1:40,000, 1:50,000, and 1:60,000 in PBS, and the dilutions were added in duplicate to the wells coated with ruthenium-labeled BSA or
20 unlabeled BSA, and the plate was incubated for one hour at room temperature. After three washes with PBS as before, the presence of bound rabbit anti-BSA antibody was determined by adding goat anti-rabbit IgG-peroxidase conjugate (1:1000 dilution in PBS of a 0.5 mg/ml solution, 50 microliters/well) to each well and incubating the plate for one hour at room temperature.
25 After washing the plate twice with PBS, 0.5% Tween-20, and twice with PBS as before, hydrogen peroxide (30%) and 2,2'-azino-di-[3-ethyl-benzthiazoline sulfonate] (KPL, Gaithersburg, MD) were mixed in equal volumes and 200 microliters were added to each well of the plate.
30 After a 30 minute incubation at room temperature, the plate was read spectrophotometrically at 414 nm. The mean background absorbance in the control wells was subtracted from the mean value of duplicate readings

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for each dilution of the rabbit anti-BSA antibody that was added to the wells coated with ruthenium-labeled BSA or unlabeled BSA. These corrected absorbance values are shown in Table V.

5 The curves obtained for the unlabeled BSA and ruthenium-labeled BSA were parallel, with corrected absorbance values at each point on the two curves at a constant ratio, approximately 0.6. Identical results were obtained for two other lots of ruthenium-labeled BSA which were made using the same activated ruthenium complex as described previously, and which had similar Ru/BSA labeling ratios. These results indicate that the ruthenium-labeled BSA is immunologically reactive and that it retains approximately 60% of its immunoreactivity when labeled with ruthenium in comparison to unlabeled BSA.

TABLE III

ABSORBANCE AT 414 nm

Rabbit Anti-BSA Antibody Dilution	Unlabeled BSA	Ruthenium- labeled BSA	Relative Immunoreactivity
20,000	1.06	0.66	62%
25 30,000	0.83	0.50	60%
40,000	0.67	0.40	60%
50,000	0.56	0.33	59%
60,000	0.47	0.28	60%

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Example 4 - Immunological Reactivity of Ruthenium-Labeled Rabbit Anti-Mouse Immunoglobulin (IgG) Antibody by a Competitive Solid Phase Enzyme Linked-Immunosorbent Assay (ELISA)

5 Rabbit anti-mouse IgG antibody labeled with 4,4' -
(dichloromethyl) - 2,2' - bipyridyl, bis(2,2' -
10 bipyridyl) ruthenium (II) (ruthenium-labeled rabbit
anti-mouse IgG antibody) was compared with unlabeled
rabbit anti-mouse IgG antibody with respect to its
ability to compete with enzyme-labeled, anti-mouse IgG
antibody for binding to mouse IgG. The wells of a 96-
well polystyrene microtiter plate were coated with a
15 solution of mouse IgG (5 micrograms/ml in PBS buffer),
incubated for 60 minutes at room temperature and washed
three times, 5 minutes per wash, with PBS. Two solu-
tions were prepared, one containing a mixture of rabbit
anti-mouse IgG-alkaline phosphatase conjugate and rab-
bit anti-mouse IgG (1 mg/ml), and the other a mixture
20 of rabbit anti-mouse IgG-alkaline phosphatase conjugate
and ruthenium-labeled rabbit anti-mouse IgG (1 mg/ml,
7.5 Ru/antibody). These two solutions, and a third
containing rabbit anti-mouse IgG-alkaline phosphatase
conjugate, were diluted 1:6000, 1:7000, 1:8000, 1:9000,
25 1:10,000, 1:12,000, 1:14,000 and 1:16,000 in PBS
containing 0.5% Tween-20, and added (50
microliters/well) to separate rows of the plate
containing bound mouse IgG. The plate was incubated
for 60 minutes at room temperature and washed twice
30 with PBS-Tween-20 and twice with PBS, 5 minutes per
wash. The enzyme substrate p-nitrophenyl phosphate
(1.5 mg/ml in 10% diethanolamine buffer, pH 9.6) was
added to each well (200 microliters/well); the plate
was incubated for 30 minutes at room temperature and

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read spectrophotometrically at 405 nm. The mean background absorbance in the control wells was subtracted from the mean value of duplicate readings for each of the three solutions at each dilution. These absorbance values are shown in Table VI.

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Three parallel curves were obtained, the top curve representing the uninhibited binding of the enzyme conjugate, and the two lower curves representing inhibition by ruthenium - labeled anti-mouse IgG and unlabeled anti-mouse IgG. The ruthenium-labeled, anti-mouse IgG curve, on a point-by-point comparison, is approximately 81% as low as the unlabeled anti-mouse IgG curve in comparison to the enzyme conjugate curve. These results indicate that the ruthenium-labeled, anti-mouse IgG antibody is immunologically reactive for its antigen (mouse IgG), and is approximately 81% as effective as unlabeled anti-mouse IgG antibody in competing with enzyme-labeled, anti-mouse IgG antibody for binding to mouse IgG.

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TABLE IV

ABSORBANCE AT 405 nm

		<u>A</u>	<u>B</u>	<u>C</u>	
5		Anti-Mouse IgG Alkaline Phosphatase (Enzyme Conjugate)	Enzyme Conjugate+ Ruthenium Labeled Anti-Mouse IgG	Enzyme Conjugate+ Unlabeled Anti-Mouse IgG	Comparative* Degree of Inhibition
	<u>Dilution</u>		<u>IgG</u>	<u>IgG</u>	
10	6,000	1.48	0.94	0.79	77%
	7,000	1.33	0.82	0.69	79%
	8,000	1.21	0.72	0.62	82%
	9,000	1.10	0.65	0.56	84%
15	10,000	1.01	0.60	0.51	82%
	12,000	0.87	0.51	0.43	80%
	14,000	0.77	0.45	0.37	81%
	16,000	0.68	0.40	0.33	80%

*(A-B/A-C) x 100%

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Example - 5 Electrochemiluminescence of Ruthenium-Labeled Bovine Serum Albumin (BSA)

5 A solution containing 7.8×10^{-6} M bovine serum albumin (BSA) labeled with 4,4' - (dichloromethyl) - 2,2'-bipyridyl, bis(2,2' bipyridyl) ruthenium (II) (ruthenium-labeled bovine serum albumin) was prepared from a stock solution of ruthenium-labeled BSA (2.6 mg/ml, 6 Ru/BSA) by dilution in phosphate-buffered saline. 26 microliters of this solution were added to 10 ml of DMSO/deionized, distilled water (1:1) containing 0.1 M TBABF₄ and 18 mM ammonium persulfate in the reaction vessel. The final ruthenium-labeled BSA concentration was 2×10^{-8} M. Electrochemiluminescence was measured as described in Example V.

20 In an analogous manner, a solution containing 7.8×10^{-6} M unlabeled BSA was prepared and added to the reaction vessel to give a final unlabeled BSA concentration of 2×10^{-8} M. The electrochemiluminescence of this solution and of a similar solution without BSA was measured. Electrochemiluminescence measurements are shown in Table V for covalently coupled, ruthenium-labeled BSA and unlabeled BSA.

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TABLE VELECTROCHEMILUMINESCENCE (ECL) OF RUTHENIUM-LABELED BSA

	<u>Solution</u>	<u>ECL (mV)</u>
5	2×10^{-8} M Ruthenium-Labeled BSA	730
	2×10^{-8} M BSA	100
10	DMSO: H ₂ O (1:1)	0
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20		
25		
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Example 6 - Electrochemiluminescence of Ruthenium-Labeled Rabbit Anti-Mouse Immunoglobulin G (IgG) Antibody

5 A solution containing 1.25×10^{-6} M rabbit anti-mouse IgG antibody labeled with 4,4' - (dichloromethyl) - 2,2' - bipyridyl, bis(2,2' - bipyridyl) ruthenium (II) (ruthenium-labeled, rabbit anti-mouse IgG antibody) was prepared from a stock solution of ruthenium-labeled, rabbit anti-mouse IgG antibody (2 mg/ml, 7.5 Ru/antibody) by dilution in phosphate-buffered saline. 80 microliters of this solution were added to 10 ml of DMSO/deionized, distilled water (1:1) containing 0.1M TBAPF₄ and 18 mM ammonium persulfate in the reaction vessel. The final ruthenium-labeled antibody concentration was 1×10^{-8} M. Electrochemiluminescence was measured as described in Example 2.

20 In an analogous manner, a solution containing 1.25×10^{-6} M unlabeled, rabbit anti-mouse IgG antibody was prepared and added to the reaction vessel to give a final unlabeled antibody concentration of 1×10^{-8} M. The electrochemiluminescence of this solution and of the solution without added antibody was also measured as described. Electrochemiluminescent measurements are shown in Table VIII for covalently-coupled, ruthenium-labeled rabbit anti-mouse IgG antibody and unlabeled rabbit anti-mouse IgG antibody.

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TABLE VI

ELECTROCHEMILUMINESCENCE (ECL) OF RUTHENIUM-
LABELED RABBIT ANTI-MOUSE IMMUNOGLOBULIN
G (IgG) ANTIBODY

	<u>Solution</u>	<u>ECL (mV)</u>
5	1×10^{-8} M Ruthenium - Labeled Rabbit Anti-Mouse IgG Antibody	892
10	1×10^{-8} M Rabbit Anti-Mouse IgG Antibody	0
	DMSO: H ₂ O (1:1)	0
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20		
25		
30		
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Example 7 - Homogeneous Electrochemiluminescent
Immunoassay for Antibody to Bovine Serum Albumin

5 A solution containing 7.8×10^{-6} M bovine serum albumin (BSA) labeled with 4,4' - (dichloromethyl-) - 2,2' bipyridyl, bis (2,2' - bipyridyl) ruthenium (II) (ruthenium-labeled bovine serum albumin) was prepared from a stock solution of ruthenium-labeled BSA (2.5 mg/ml, 6 Ru/BSA) by dilution in phosphate-buffered saline (PBS). 10 26 microliters of this solution were added to 10 ml of DMSO/deionized, distilled water (1:1) containing 0.1M TBABF₄ and 18 mM ammonium persulfate in the reaction vessel. The final ruthenium-labeled BSA concentration was 2×10^{-8} M. Electrochemiluminescence 15 was measured as described in Example 2.

In an analogous manner, a solution containing 7.8×10^{-6} M unlabeled BSA was prepared and added to the reaction vessel to give a final unlabeled BSA concentration of 5×10^{-8} M. 20 The electrochemiluminescence of this solution and of a similar solution without BSA were measured.

A solution containing 3.75×10^{-5} M rabbit anti-BSA antibody was prepared from a stock solution of rabbit 25 anti-BSA antibody (6.0 mg/ml) by dilution in PBS, and an aliquot (26 microliters) was added to the solution of ruthenium-labeled BSA in the reaction vessel to give a final rabbit anti-BSA antibody concentration of 1×10^{-7} M.

30 The electrochemiluminescence of the resulting mixture of ruthenium-labeled BSA antigen and antibody (rabbit anti-BSA) was measured. The results shown in Table VII indicate a reduction in the electro-

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chemiluminescence of the ruthenium-labeled BSA upon addition of rabbit anti-BSA antibody and demonstrate that a homogeneous electrochemiluminescent detection of antibody to BSA may be achieved. Based upon these results one skilled in the art would know that a homogeneous electrochemiluminescent immunoassay for detecting other analytes of interest may be developed.

TABLE VII

REDUCTION OF ELECTROCHEMILUMINESCENCE (ECL) OF
RUTHENIUM-LABELED BOVINE SERUM ALBUMIN UPON
BINDING OF ANTIBODY

	UNLABELED BSA (CONTROL)	RUTHENIUM- LABELED BSA (ANTIGEN)	RABBIT ANTI-BSA (ANTIBODY)	ECL (mV)
15	0	$2 \times 10^{-8}M$	0	727
	0	$2 \times 10^{-8}M$	$1 \times 10^{-7}M$	92
	$2 \times 10^{-8}M$	0	0	94

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Example 8 - Homogeneous Electrochemiluminescent
Immunoassay for Mouse Immunoglobulin G (IgG)

5 A solution containing $6.25 \times 10^{-6} \text{M}$ rabbit anti-mouse
IgG antibody labeled with 4,4' - (dichloromethyl) -
2,2' - bipyridyl, bis (2,2' - bipyridyl) ruthenium
(II) (ruthenium-labeled rabbit anti-mouse IgG antibody)
was prepared from a stock solution of ruthenium-labeled
10 rabbit anti-mouse IgG antibody (2 mg/ml, 7.5
Ru/antibody) by dilution in phosphate-buffered saline
(PBS). 80 microliters of this solution were added to
10 ml of DMSO/deionized, distilled water (1:1)
containing 0.1M TBABF₄ and 18mM ammonium persulfate in
the reaction vessel. The final ruthenium-labeled
15 antibody concentration was $5 \times 10^{-8} \text{M}$.
Electrochemiluminescence was measured as described in
Example 2.

20 In an analogous manner, a solution containing $6.25 \times$
 10^{-6}M unlabeled rabbit, anti-mouse IgG antibody was
prepared and added to the reaction vessel to give a
final unlabeled antibody concentration of $5 \times 10^{-8} \text{M}$.
The electrochemiluminescence of this solution and of a
similar solution without antibody were measured.

25 A solution containing $2.5 \times 10^{-5} \text{M}$ mouse IgG was
prepared from a stock solution of mouse IgG (4.0 mg/ml)
by dilution in PBS, and different aliquots (20
microliters and 40 microliters) of this solution were
30 added to the solution of ruthenium-labeled, anti-mouse
IgG antibody in the reaction vessel to give final
mouse IgG concentrations of $5 \times 10^{-8} \text{M}$ and $1 \times 10^{-7} \text{M}$,
respectively.

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The electrochemiluminescence of the resulting mixture of ruthenium-labeled, anti-mouse IgG antibody and the antigen (mouse IgG) was measured. The results are shown in Table VIII. The dependence of the electrochemiluminescence measurements upon the concentration of the mouse IgG antigen is shown in Figure 1. These results demonstrate a reduction in the electrochemiluminescence of the ruthenium-labeled antibody upon addition of antigen. Based upon these results one skilled in the art would know that a homogeneous electrochemiluminescent immunoassay for determining the concentration of other analytes of interest may be developed.

TABLE VIII

REDUCTION OF ELECTROCHEMILUMINESCENCE (ECL)
OF RUTHENIUM-LABELED ANTIBODY UPON BINDING OF ANTIGEN

UNLABELED ANTI-MOUSE IgG (CONTROL)	RUTHENIUM- LABELED ANTI-MOUSE IgG (ANTIBODY)	MOUSE IgG (ANTIGEN)	ECL (MV)
0	$5 \times 10^{-8}M$	0	1610
0	$5 \times 10^{-8}M$	$5 \times 10^{-8}M$	1360
0	$5 \times 10^{-8}M$	$1 \times 10^{-7}M$	1240
$5 \times 10^{-8}M$	0	0	0

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Example 9 - Heterogeneous Electrochemiluminescent Immunoassay for Legionella Using a Mouse Anti-Legionella Immunoglobulin G (IgG) Antibody and Ruthenium-Labeled Rabbit Anti-Mouse Immunoglobulin G (IgG) Antibody

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A formalinized suspension of the bacterium Legionella micdadei was adjusted to an optical density (at 425nm) of 1.00 by dilution with PBS buffer. Approximately 3×10^9 cells were added to a conical microcentrifuge tube. The cells were centrifuged (10 minutes, 10,000 RPM), the supernatant decanted, and the cells resuspended in a 1:50 dilution of a mouse monoclonal IgG antibody, (1.45 mg/ml) specific for Legionella micdadei, in PBS (1 ml). After incubation at room temperature for 1 hour, the cells were centrifuged, the supernatant decanted, the cells resuspended in PBS buffer and centrifuged again. Following decantation of the supernatant, the cells were resuspended in a 1:50 dilution (in PBS) of rabbit anti-mouse IgG antibody labeled with 4,4' - (dichloromethyl) - 2,2' - bipyridyl, bis (2,2' - bipyridyl) ruthenium (II) i.e. ruthenium-labeled rabbit anti-mouse IgG antibody, (2mg/ml, 7.5 Ru/antibody). After incubation at room temperature for 1 hour, the cells were centrifuged, the supernatant decanted, and the cells resuspended in PBS and washed twice, with centrifugation, as before. Following the last wash the cells were resuspended in 200 microliters of PBS. 100 microliters of the cell suspension was added to the reaction vessel containing 10 ml of DMSO/deionized, distilled water (1:1) containing 0.1 M TBABF₄ and 18 mM ammonium persulfate and transferred to the reaction vessel. The electrochemiluminescence was measured for the cell suspension. Another 100

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microliters of the cell suspension was added to the reaction vessel and electrochemiluminescence measured. Electrochemiluminescence was measured for the solution without cells as a control according to the method described in Example 2. The results shown in Table IX indicate a heterogeneous electrochemiluminescent immunoassay for Legionella using ruthenium-labeled rabbit anti-mouse IgG antibody has been successfully carried out.

TABLE IX

HETEROGENEOUS ELECTROCHEMILUMINESCENT (ECL) IMMUNOASSAY
FOR LEGIONELLA MICDADEI

	<u>Sample</u>	<u>ECL (mV)</u>
15	<u>Legionella micdadei</u> cell suspension, 1.9×10^9 cells in DMSO/H ₂ O (1:1)	160
	<u>Legionella micdadei</u> cell suspension, 9.3×10^8 cells in DMSO/H ₂ O (1:1)	90
20	DMSO: H ₂ O (1:1)	0

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Example 10 - Homogeneous Electrochemiluminescent Immunoassay for Legionella Using a Mouse Anti-Legionella Immunoglobulin G (IgG) Antibody and Ruthenium-Labeled Rabbit Anti-Mouse Immunoglobulin G (IgG) Antibody

5

A suspension of the bacterium Legionella micdadei was prepared and incubated with a mouse monoclonal IgG antibody specific for Legionella as described in Example 9. The cells were centrifuged, washed, and resuspended in 0.2 ml of PBS. An aliquot (80 microliters) of rabbit, anti-mouse IgG antibody labeled with 4,4' - (dichloromethyl) - 2,2' - bipyridyl, bis (2,2' - bipyridyl) ruthenium (II), i.e. ruthenium-labeled rabbit anti-mouse IgG antibody ($1.25 \times 10^{-6} \text{M}$) was added to the cell suspension, and the mixture was incubated for 2 hours at room temperature. As a control, an identical dilution of ruthenium-labeled, rabbit anti-mouse IgG antibody was incubated in the same way in the absence of the cell suspension. After the incubation period, the solution of labeled antibody was added to 10 ml of DMSO/deionized, distilled water (1:1) containing 0.1 M TBABF₄ and 18mM ammonium persulfate in the reaction vessel to give a final ruthenium-labeled, rabbit anti-mouse IgG antibody concentration of $1 \times 10^{-8} \text{M}$. The electrochemiluminescence was measured as described in Example 2. The same procedure was followed for the cell suspension with added ruthenium-labeled rabbit anti-mouse IgG antibody. The results, shown in Table X, indicate a reduction of the electrochemiluminescent emission upon the interaction of ruthenium-labeled, anti-mouse IgG antibody with mouse monoclonal antibody bound to Legionella and that a homogeneous

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electrochemiluminescent immunoassay for Legionella micdadei has been successfully carried out.

TABLE X

HOMOGENEOUS ELECTROCHEMILUMINESCENT (ECL) IMMUNOASSAY
FOR LEGIONELLA MICDADEI

5

<u>SAMPLE</u>	<u>ECL (mV)</u>
1 x 10 ⁻⁸ M Ruthenium - Labeled Anti-mouse IgG Antibody	976

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1 x 10 ⁻⁸ M Ruthenium - Labeled Anti-Mouse IgG Antibody + Monoclonal Antibody Bound To <u>Legionella micdadei</u>	803
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DMSO: H ₂ O (1:1)	0
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Example 11 - Increase in Electrochemiluminescence Upon Release of A Ruthenium-Labeled Antibody Bound to Bacteria

5 A formalinized suspension of the bacterium *Legionella micdadei* was adjusted to an optical density (at 425nm) of 1.00 by dilution with PBS buffer and 2 ml of this suspension were added to a conical microcentrifuge tube. The cells were centrifuged (10 minutes, 10,000
10 RPM), the supernatant decanted, and the cells were resuspended in a 1:10 dilution in FBS (0.5 ml) of a mouse monoclonal IgG antibody, (1.45 mg/ml) specific for *Legionella micdadei*. After incubation at room temperature for 1 hour, the cells were centrifuged as before, the supernatant was decanted, the cells were
15 resuspended in PBS buffer and centrifuged again. Following decantation of the supernatant, the cells were resuspended in a 1:50 dilution (in PBS) of ruthenium-labeled, rabbit anti-mouse IgG antibody (1ml, 7.5 Ru/antibody). After incubation at room temperature for 1 hour, the cells were centrifuged as before, the supernatant was decanted, and the cells resuspended in PBS and washed twice, with centrifugation as before. Following the last wash the cells were resuspended in
20 100 microliters of either PBS or 1.0M acetic acid - 0.9%NaCl (normal saline) solution and incubated at room temperature for 40 minutes. After centrifugation, 100 microliters of the cell supernatant fluid was transferred into the reaction vessel along with 10 ml of DMSO-deionized, distilled water (1:1) containing 0.1
25 M TBABF₄ and 18 mM ammonium persulfate. The electrochemiluminescence of the acetic acid/normal saline cell supernatant fluid and for the supernatant fluid from the PBS washed cells was measured according

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to the method described in Example 2. The electrochemiluminescence measurements are shown in Table XI, and demonstrate that the elution of the ruthenium-labeled, rabbit anti-mouse IgG from the monoclonal antibody coated Legionella bacteria by treating the cells with 1.0M acetic acid-normal saline (Ref.23) results in an increase in the electrochemiluminescence generated by the unbound ruthenium-labeled antibody. These results also show that the ruthenium labeled antibody is bound to the monoclonal antibody-coated Legionella, and that the PBS wash did not result in an increase in ECL in comparison to the background signal.

TABLE XI

ELECTROCHEMILUMINESCENCE (ECL) OF SUPERNATANT FLUIDS OF CELLS COATED WITH RUTHENIUM-LABELED ANTI-MOUSE IMMUNOGLOBULIN(IgG)

<u>Solution</u>	<u>ECL (mV)</u>
Background control:	
100 microliters 1.0M acetic acid-normal saline	134
100 microliters of PBS cell wash supernatant fluid	121
100 microliters of 1.0M acetic acid-normal saline solution cell wash supernatant fluid	214

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Example 12 - Homogeneous Competitive Immunoassay For
Pregnane - diol - 3 Glucuronide (PD3G) In Urine

5 Pregnane-diol-3-glucuronide (PD3G) may be detected and
quantified in urine by contacting a sample of urine
with a known amount of PD3G labeled with an
electrochemiluminescent moiety and a known amount of an
anti-PD3G antibody under conditions such that the PD3G
10 present in the sample and the PD3G-
electrochemiluminescent moiety compete for binding
sites on the antibody. After a suitable time the
resulting sample may be induced to repeatedly emit
electromagnetic radiation upon direct exposure to an
15 electrochemical energy source effective for inducing
the electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
quantified, and the amount of PD3G present in the
sample determined therefrom.

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Example 13 - Labeling Nucleic Acids with a tris-Ruthenium bipyridyl-n-hydroxysuccinimide ester

Nucleic acid samples (5 to 25 micrograms) in 10mM tris hydrochloride (pH 8.0)-1mM EDTA may be heat denatured, cooled and modified by bisulphite catalysed transamination of cytosine residues with ethylenediamine for three hours at 42°C. After overnight dialysis against three changes of 5mM sodium phosphate buffer, pH 8.5, the samples may be concentrated to 100 microliters by ultrafiltration. These modified nucleic acids (1 to 10 micrograms) may be diluted in 100 microliters with 0.1 M sodium phosphate buffer, pH 8.5.

A tris-ruthenium bipyridyl-N-hydroxysuccinimide ester derivative may be prepared as a 0.2M stock solution in N, N' dimethylformamide (DMF) by methods known in the art. 5 microliters of this ester solution may be added to the modified nucleic acid solution in 0.1 M sodium phosphate buffer, pH 8.5 and incubated at room temperature for 1 hour. Labeled nucleic acid probes may be purified by dialysis against at least three changes of 150mM sodium chloride - 10mM sodium phosphate buffer (pH 7.0) stored at 4°C until used.

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Example 14 - Nucleic Acid Hybridization Assay for Human
T-Cell Leukemia III Virus (HTLV-III) in Blood

5 HTLV-III virus may be detected in whole blood by
treating the blood to release the RNA from virus
particles. A sample containing the HTLV-III RNA is
contacted with a single-stranded oligonucleotide probe
complementary to the HTLV-III RNA and labeled with an
10 electrochemiluminescent moiety. After a suitable time
the resulting sample may be induced to repeatedly emit
electromagnetic radiation upon direct exposure to an
electrochemical energy source effective for inducing
the electrochemiluminescent moiety to repeatedly emit
15 electromagnetic radiation. Emitted radiation may be
quantified, and the amount of HTLV-III RNA present in
the sample determined therefrom.

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Example 15 - Homogeneous Nucleic Acid Hybridization
Assay for Legionella Bacteria in a Clinical Sample

Legionella bacteria may be detected in a clinical
5 sample such as sputum by treating the sputum to release
the ribosomal RNA from the bacteria. The resulting
sample is contacted with a single-stranded
oligonucleotide probe that is complementary to
10 sequences in the ribosomal RNA, specific for
Legionella, and labeled with an electrochemiluminescent
moiety. After a suitable time the resulting sample may
be induced to repeatedly emit electromagnetic radiation
upon direct exposure to an electrochemical energy
15 source effective for inducing the
electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
quantified, and the amount of Legionella Bacteria
present in the sample determined therefrom.

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Example 16 - Hybridization Assay for the Detection of
Cytomegalovirus DNA Integrated into Genomic DNA by
Strand Displacements Method

5 Cytomegalovirus (CMV) DNA may be detected in human
genomic DNA by treating a tissue sample, for example
lymphocytes, to release the DNA and cleaving the DNA
with restriction enzymes. The resulting sample
10 containing double-stranded fragments of CMV is
contacted with a single-stranded oligonucleotide probe
that is complementary to the CMV DNA, labeled with a
electrochemiluminescent moiety and capable of
displacing one of the strands of the DNA duplex. After
a suitable time the resulting sample may be induced to
15 repeatedly emit electromagnetic radiation upon direct
exposure to an electrochemical energy source effective
for inducing the electrochemiluminescent moiety to
repeatedly emit electromagnetic radiation. Emitted
radiation may be quantified, and the amount of
20 Cytomegalovirus RNA present in the sample determined
therefrom.

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Example 17 - Hydrization Assay for the Detection of the
Ras-Oncogene in Human Bladder Carcinoma Cells using a
Heterogeneous Method

5 The ras-oncogene may be detected in human bladder
carcinoma cells by treating a tissue sample to release
the DNA, cleaving the DNA with restriction enyzmes,
melting the DNA to single strands and binding to
10 nitrocellulose paper as previously described in
Maniatis, T. et al. (24). The filter paper with bound
single-stranded DNA is contacted with an
oligonucleotide probe specific for the ras-oncogene,
and labeled with a electrochemiluminescent moiety.
After a suitable reaction time the resulting sample
15 may be induced to repeatedly emit electromagnetic
radiation upon direct exposure to an electrochemical
energy source effective for inducing the
electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
20 detected and the presence of ras-oncogene RNA in the
sample determined.

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Example 18 - An Electrochemiluminescent Polymer - Based
Enzyme Assay

5 A macromolecular enzyme substrate, for example starch,
dextran or a synthetically prepared macromolecular
polymer, may be labeled with an electrochemiluminescent
moiety. The labeled substrate may be used to determine
and quantify the presence of an enzyme present in a
10 multicomponent system. The labeled substrate may also
be used to quantify an amount of enzyme linked to an
antibody, DNA probe, RNA probe, streptavidin, avidin,
or biotin. The labeled substrate may be cleaved into
smaller fragments selectively by an appropriate enzyme.
Any enzyme-substrate combination may be used. Examples
15 of enzymes include glycosidases, glucosidases,
amylases, proteases, endonucleases, lyases and
dextranases. After a suitable time the resulting
sample may be induced to repeatedly emit
electromagnetic radiation upon direct exposure to an
20 electrochemical energy source effective for inducing
the electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
quantified, and the amount of enzyme present in the
sample determined therefrom. The advantage is that an
25 amplification of the electrochemiluminescent signal
will result from the cleaved fragments each labeled
with electrochemiluminescent moiety.

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Example 19 - Detection of Streptococci by an
Electrochemiluminescent Enzyme Assay

5 A sample containing streptococci may be contacted with
a reagent mixture containing a synthetic peptide
labeled with an electrochemiluminescent moiety. The
synthetic peptide being a substrate specific for a
peptidase produce by the bacteria. Action of the
10 peptidase on the synthetic peptide results in the
production of fragments labeled with the
electrochemiluminescent moiety. After a suitable time
the resulting sample may be induced to repeatedly emit
electromagnetic radiation upon direct exposure to an
electrochemical energy source effective for inducing
15 the electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
quantified, and the amount of streptococci present in
the sample determined therefrom.

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Example 20 - Enzyme Immunoassay for Hepatitis B Surface
Antigen Based On Electrochemiluminescence

5 A blood sample containing Hepatitis B surface antigen
(HBsag) is contacted for a suitable time with an
antibody specific for the HBsag and labeled with
dextranase. The antibody not bound to HBsag is removed
and the antigen-antibody complex is contacted with a
10 dextran polymer labeled with an electrochemiluminescent
moiety. Action of the dextranase on the labeled
polymers will result in the production of fragments
each labeled with the electrochemiluminescent moiety.
After a suitable time the resulting sample may be
15 induced to repeatedly emit electromagnetic radiation
upon direct exposure to an electrochemical energy
source effective for inducing the
electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
20 quantified, and the amount of Hepatitis B surface
antigen present in the sample determined therefrom.

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Example 21 - Homogeneous Assay for Bradykinin receptors
using an Electrochemiluminescent-labeled Bradykinin

5 A tissue sample containing bradykinin receptors is
suitably prepared and contacted with bradykinin labeled
with an electrochemiluminescent moiety. After a
suitable amount of time the resulting sample may be
induced to repeatedly emit electromagnetic radiation
10 upon direct exposure to an electrochemical energy
source effective for inducing the
electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
quantified, and the amount of bradykinin receptor
present in the sample determined therefrom. This assay
15 could also be used for measuring other ligand-receptor
interactions such as estrogen-estrogen receptor. In
addition this assay may be used to determine and
quantify the amount of unlabeled ligands using a
competitive binding assay format.

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Example 22 - Use of an Electrochemiluminescent moiety
for Detecting Nucleic Acid Hybrids

5 A sample containing nucleic acid hybrids, such as
double-stranded DNA, DNA-RNA duplexes, or double-
stranded RNA is contacted with an
electrochemiluminescent moiety that specifically
intercalates into nucleic acid hybrids. After a
suitable amount of time the resulting sample may be
10 induced to repeatedly emit electromagnetic radiation
upon direct exposure to an electrochemical energy
source effective for inducing the
electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
15 quantified, and the amount of nucleic acid hybrids
present in the sample determined therefrom.

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Example 23 - Detection of Human T-Cell Leukemia Virus
III (HTLV-III) Antigen Complexed to Antibody in Saliva
by a Homogeneous Immunoassay using
Electrochemiluminescence

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A saliva sample containing HTLV-III complexed to
antibody is contacted with a solution containing a
chaotropic agent to disrupt the antigen-antibody
complexes. This solution is then contacted with an
10 antibody specific for HTLV-III and labeled with an
electrochemiluminescent moiety. The chaotropic agent is
removed allowing the labeled and unlabeled antibodies
to recombine with antigen. After a suitable amount of
time the resulting sample may be induced to repeatedly
15 emit electromagnetic radiation upon direct exposure to
an electrochemical energy source effective for inducing
the electrochemiluminescent moiety to repeatedly emit
electromagnetic radiation. Emitted radiation may be
quantified, and the amount of human T-cell leukemia
20 virus III (HTLV-III) antigen present in the sample
determined therefrom. These methods are applicable
to samples containing other types of antigen-antibody
complexes such as hepatitis antigen-antibody complexes,
cytomegalavirus-antibody complexes, and non-A, non-B
25 hepatitis-antibody complexes in serum.

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Example 24 - Immunoassay for the Detection and Identification of a Multiplicity of Staphylococcal Enterotoxins

5 Monoclonal antibodies specific for each of the Staphylococcal enterotoxins A, B, C, D, and E and monoclonal antibodies which are cross-reactive for these enterotoxins were purified. Each antibody was purified from ascitic fluid by passage of the ascites
10 through a column of Staphylococcal protein A coupled to an agarose gel support matrix which is provided with binding, elution, and regeneration buffers as part of the Monoclonal Antibody Purification System (Bio-Rad Laboratories, Inc.). In the purification procedure, 2
15 ml of ascitic fluid typically containing 5-15 mg of monoclonal antibody per milliliter, were prefiltered by placing a Metricell membrane filter (Gelman Sciences, Inc.) between two F-13 analytical papers (Schleicher and Schuell, Inc.) in the bottom of a 10 ml syringe
20 (Becton Dickenson, Inc.), introducing the ascitic fluid into the syringe, inserting the plunger, and filtering the ascitic fluid into a collection vessel. The filtrate was mixed with an equal volume of the binding buffer and applied to a 5 ml protein A-agarose column.
25 The column reagent reservoir was then filled with the binding buffer to begin the elution process. The effluent was monitored for absorbance at 280 nm (A_{280}), and 1 ml fractions were collected. When the A_{280} had returned to a stable baseline value, the column was
30 washed with 5 bed volumes of binding buffer, and the elution buffer was then used to elute the purified immunoglobulin from the column. To neutralize the immunoglobulin, the eluate was collected in 0.3 ml of 1 M Tris-HCl, pH 9.0. When the A_{280} had again returned

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to a stable baseline value, the column was washed with 5 bed volumes of elution buffer, followed by 10 bed volumes of regeneration buffer and 5 bed volumes of binding buffer so that the column was ready for the next purification cycle. The fractions containing the purified immunoglobulin were pooled, and concentrated using a stirred ultrafiltration cell (Amicon Corp.). The final concentration of purified immunoglobulin was greater than 5 mg/ml as determined by the Lowry assay for total protein. The purified antibody was dialyzed against two changes of phosphate-buffered saline (PBS) containing 0.1% sodium azide for 48 hours at 4°C.

The purified monoclonal antibodies were titrated in a 96-well microtiter plate ELISA system to provide a measure of their immunological reactivity for specific enterotoxin. The end point titers for the antibodies which were obtained were compared to reference titer values for previously acceptable lots of each antibody. Antibodies of sufficient titer were accepted as immunologically functional coating antibodies for immobilization to a diagnostic reagent holder, e.g. dipstick or for conjugation to an enzyme for use as probes in the immunoassay.

Diagnostic reagent holders (dipsticks) with nitrocellulose membranes attached were prepared for immobilization of antibodies to the membrane surface by first immersing the membranes in phosphate-buffered saline for one hour at room temperature to improve the wettability of the membrane. The sticks were removed from the solution and the membranes were allowed to air dry. Solutions of the purified monoclonal antibodies, each of which is specific for one of the Staphylococcal enterotoxins A, B, C, D or E, and a non-specific

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control mouse immunoglobulin (Jackson Immuno Research Laboratories, Inc.) were applied to different regions of the membrane by spotting 2 microliters of antibody solution directly onto the membrane surface. The concentration of each antibody used was one which was known to saturate the binding sites of the nitrocellulose: 250 micrograms/ml of 3A antibody (specific for A toxin), 50 micrograms/ml of 2B antibody (specific for B toxin), 300 micrograms/ml of 1C3 antibody (specific for C1, C2, and C3 toxins) 200 micrograms/ml of 3D antibody (specific for D toxin) 250 micrograms/ml of 4E antibody (specific for E toxin), and 200 micrograms/ml of non-specific control mouse immunoglobulin. The dipsticks were allowed to air dry at room temperature, and the remaining protein binding sites on the membrane were blocked by immersing the dipsticks in a solution of phosphate-buffered saline containing 0.5% Tween 20 and 3% bovine serum albumin. Following an overnight incubation at room temperature in this blocking solution, the sticks were allowed to air dry.

Purified monoclonal antibodies 2A (specific for A and E toxins), 6B (specific for B, C1, C2, and C3 toxins), and 1D (specific for D toxin) were covalently linked to the enzyme alkaline phosphatase for use as conjugated probes to determine the presence of A, B, C1, C2, C3, D, and E toxins bound to antibodies immobilized to the membrane surface of the diagnostic reagent holder dipstick. Each conjugate was prepared by a stoichiometrically controlled two step procedure which utilizes glutaraldehyde as the bifunctional cross-linking agent. In this procedure 0.47 ml of EIA grade alkaline phosphatase (2500 U/mg, Boehringer Mannheim Corp.) was added to 1.2 ml of 50 mM potassium phosphate

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buffer pH 7.2, containing 13.4 microliters of aqueous glutaraldehyde solution (25%, Sigma Chemical Co.). The mixture was stirred at room temperature for 90 minutes to allow attachment of glutaraldehyde to free amino groups of the enzyme molecule. After this incubation period, a 2 ml aliquot of purified monoclonal antibody at a concentration of 1 mg/ml was added, the mixture stirred for an additional 90 minutes, and placed on ice to stop the conjugation reaction. The conjugate was dialyzed against two changes of phosphate-buffered saline containing 0.1% sodium azide for 48 hours at 4°C. Bovine serum albumin was added to the dialysate to a final concentration of 3% for stabilization during prolonged storage at 4°C. The antibody-enzyme conjugates were titrated in a 96-well microliter plate ELISA system to provide a measure of their immunological reactivity for specific enterotoxin. The end point titers for the conjugates which were obtained were compared to reference titer values for previously acceptable lots of each conjugate. Conjugates of sufficient titer were accepted for use as probes in the dipstick assay for Staphylococcal enterotoxins.

Solid foods such as ham, sausage, noodles and cheese, which represent the types of foods most commonly associated with outbreaks of Staphylococcal food poisoning, were converted to a homogeneous liquid suspension in order to perform the assay for Staphylococcal enterotoxins. In a typical extraction, 20 milliliters of water were added to 20 grams of food, and the mixture was homogenized for 30 seconds using a stomacher lab-blender (Tekmar Co.). For more viscous foods, twice the volume of water was used. Staphylococcal enterotoxin was added to the food either

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before or after homogenization. The food homogenate was tested directly using the diagnostic reagent holder dipstick described above. Positive results were obtained with food samples to which low levels of enterotoxin were added. Additional extraction steps were carried out on the food homogenates to improve the sensitivity of Staphylococcal enterotoxin detection. The homogenate was typically adjusted to pH 4.5 with 6 N HCl and centrifuged for 20 minutes at 20,000 x g, and the supernatant was adjusted to pH 7.5 with 5 N NaOH, centrifuged again 20 minutes at 20,000 x g, and the supernatant was used directly in the dipstick assay. For food extracts prepared in this way the sensitivity of enterotoxin detection was 1 nanogram per milliliter of extract for each toxin in each food tested.

Liquid foods such as milk were tested directly by immersing the diagnostic reagent holder dipstick in the liquid food and running the test in an identical manner as for the solid food homogenates. In testing these foods, it was also determined that sensitivity may be improved to 1 nanogram per milliliter by carrying out the additional extraction steps described above.

Diagnostic reagent holders (dipsticks) provided with a nitrocellulose membrane having immobilized to it monoclonal antibodies specific for the individual Staphylococcal enterotoxins were immersed in a tube containing 1 ml of a liquid food, food homogenate, or food extract prepared as described above, or 1 ml of FBS. Each of these solutions contained 1 ng/ml of an added Staphylococcal enterotoxin or a mixture of enterotoxins, each at 1 ng/ml concentration. The dipstick remained immersed in these sample solutions for a 1 hour incubation period at room temperature with

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shaking on a rotary shaker. The dipsticks were then removed from the samples and washed with shaking for 5 minutes in PBS with 0.5% Tween-20, then twice more with shaking in PBS alone, 5 minutes per wash. A mixture of monoclonal antibody-alkaline phosphatase conjugates prepared as described was made as follows: 1:1000 dilutions of 2A conjugate (specific for A and E toxins), 6B conjugate (specific for B, C1, C2 and C3 toxins), and 1D conjugate (specific for D toxin) were made in the same solution (30 microliters of each conjugate in 30 ml of PBS containing 0.5% Tween-20 and 3% BSA). The dipsticks were immersed in this conjugate mixture, 2 ml per stick, and incubated for 30 minutes at room temperature with shaking. The dipsticks were washed twice in PBS-Tween, then three times with shaking in PBS, 5 minutes per wash, to remove unbound monoclonal antibody-alkaline phosphatase conjugate from the membrane surface.

Following the removal of unbound monoclonal antibody-enzyme conjugate from the diagnostic reagent holder dipstick, the presence of bound conjugate was indicated by immersing the dipstick in a solution of 5-bromo 4-chloro indolyl phosphate (BCIP, Sigma Chemical Co.) and nitroblue tetrazolium (NBT, Sigma Chemical Co.). The BCIP and NBT solutions were prepared individually as follows: 3.2 mg of BCIP was dissolved in 10 ml of 0.1M Tris-HCl, 0.1M NaCl, 5mM MgCl₂, and 8.8 mg of NBT was dissolved in 10 ml of the same solution. These solutions were mixed together immediately prior to use. The dipsticks were immersed in the substrate mixture for 30 minutes at room temperature with shaking, then rinsed with water and air dried, providing a permanent record of the assay result. The entire assay procedure requires few to none pipetting steps, minimal hands-on

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time, and can be performed in approximately two hours. The substrate of the enzyme conjugate, BCIP, was hydrolyzed by the conjugate bound to the toxin on the membrane to yield a reaction product which reduces the NBT to an insoluble blue product that binds to the membrane on the dipstick. The presence of Staphylococcal enterotoxin in the sample which was bound by the immobilized first monoclonal antibody on the membrane and detected by the binding of the second monoclonal antibody-enzyme conjugate, was indicated by the presence of a blue spot on the nitrocellulose membrane. When no Staphylococcal enterotoxin was present in the sample the nitrocellulose membrane remained white. The area of the membrane to which the control mouse immunoglobulin was immobilized remained white in each case. The identification of the particular Staphylococcal enterotoxin that was present, that is, A, B, C, D or E, was indicated by the presence of a blue spot on the distinct and identifiable area of the membrane to which monoclonal antibody specific for that toxin was immobilized. This method, which employs a diagnostic reagent holder dipstick, is capable of detecting one nanogram per milliliter of one or a multiplicity of Staphylococcal enterotoxins in a liquid food, food homogenate or food extract.

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Example 25 - Enzyme Immunoassay (EIA) for Coliform Bacteria

5 Experiments were performed to compare the detection efficiency for fecal coliforms between the method of the present invention (CAP-EIA MPN) and the EPA approved MPN confirmatory procedure using the EC broth at 44.5⁰C.

10 The experiments were performed as a standard 5-vial MPN assay with the following modifications. Instead of the lauryl sulfate tryptose (LST; Difco Labs, Detroit, MI) broth, or lactose broth (Difco Labs, Detroit, MI) phenol red lactose broth (PRLB; Difco Labs, Detroit, MI) containing 80 micrograms/ml of 4-methylumbelliferone glucuronide (MUG; Sigma Chemical Co., St. Louis, MO) was used as the presumptive medium. PRLB was chosen because the phenol red dye allowed the detection of acid production from the fermentation of lactose. Gas which evolved from lactose fermentation was trapped by the inverted Durham vials which were inserted inside each of the vials. MUG was included as an option to provide a preliminary confirmation for the presence of E. coli. E. coli, the principal fecal coliform, is the only organism prevalent in water which can cleave MUG to release a fluorogenic radical visible under UV light (4, 10).

30 The CAP-EIA MPN assay format provides 3 types of data: 1) acid and gas production from the fermentation of lactose (presumptive coliform analysis); 2) fluorescence from the cleavage of MUG (presumptive confirmation for E. coli or fecal coliforms); and 3) CAP-enzyme immunoassay confirmation (confirmatory test

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based on monoclonal antibodies). The results of the MUG and CAP-EIA reactions were then compared to the results obtained by Standard Methods (1) based on lactose fermentation at the elevated temperature of 44.5°C in EC broth (Difco Labs, Detroit, MI).

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The experiment was performed as follows: to obtain the three 10-fold dilution series as required for an MPN assay, the following volumes of water sample collected from a nearby creek were used to inoculate the vials containing 10 ml of PRLB-MUG and equipped with a polystyrene well inside the vial cap.

10

Series A - 5 vials, 1.00ml inoculum

Series B - 5 vials, 0.10ml inoculum

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Series C - 5 vials, 0.01ml inoculum

All vials were then incubated overnight for approximately 18- 20 hours at 35°C.

The following day, all the vials were examined for acid and gas production as well as fluorescence under UV light. All vials which showed turbidity (growth), regardless of the acid or gas reactions, were inverted to allow the cell-medium suspension to fill the polystyrene well inside the cap. The inverted vials were held at 35°C for 1 hour to allow bacteria (antigen) to attach to the polystyrene cap-wells. The antigen bound cap-wells were then removed from the vials to continue with the antibody based confirmatory tests.

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To perform the EIA (enzyme immunoassay) portion of the assay, the wells were treated for 30 minutes at 35°C with a solution of 3% bovine serum albumin (BSA; Sigma

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Chemical Co., St. Louis, MO) in phosphate-buffered saline (PBS; NaCl, 8.5 g; Na_2HPO_4 , 1.02 g; $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$, 0.386 g; distilled water, 1000 ml, pH 7.2) to block out any unbound sites on the polystyrene. Once the BSA solution was decanted, the wells were washed twice with PBS, then 50 microliters of a hybridoma cell supernatant which contained monoclonal antibodies directed to E. coli cells were added to each well and incubated for 1 hour at 35°C to allow the antibody to interact with the specific antigens (E. coli cells) bound to the polystyrene. After decanting the antibody supernatant, traces of any unbound antibody were removed by washing the wells 3 times using a PBS - Tween 20 solution (PBS + 0.05% Tween 20, J.T. Baker Chemical Co., Phillipsburg, NJ). An affinity-purified alkaline phosphatase-labeled goat antibody to mouse IgG (conjugate; KPL, Gaithersburg, MD) was diluted 1:1000 in PBS, and 50 microliter aliquots were added to each well. Following a 1 hour incubation at 35°C , any unbound conjugate in the wells was removed by 4 washings with PBS-Tween 20 solution. The alkaline phosphatase substrate used for detection was sodium para-nitrophenyl phosphate or Sigma-104 substrate (Sigma Chemical Co., St. Louis, MO), prepared at a final concentration of 1 mg/ml in 10% diethanol-amine buffer (diethanolamine, 97 ml; NaN_3 , 0.2 g; $\text{MgCl}_2 \cdot 6\text{H}_2\text{O}$, 100 mg; and distilled water, 800 ml, pH 9.8). The substrate was added in 100 microliter aliquots per well, and following 30 minutes incubation at 35°C , the reaction in the wells was observed visually. For the purpose of quantitation in this example, the reacted substrate solution was transferred into a microtiter plate and determined instrumentally with Titertek Multiscan (Flow Laboratories, Mcleans, VA) equipped with a 405 nm filter.

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To perform the conventional confirmatory test for fecal coliforms using EC broth (Difco Labs, Detroit, MI), a loopful of growth from each of the overnight PRLB was aseptically transferred into a tube containing 10 ml of EC broth and equipped with an inverted Durham vial. The tubes were then incubated for 24-48 hours at 44.5°C as specified by Standard Methods (1) then examined for evidence of lactose fermentation (trapped gas inside the Durham vials). Any trace of gas bubbles within the vials was considered a positive reaction.

The summarized data for each experiment are shown on Table XII and XIII.

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The results of experiment 1 indicate that, based on gas production from lactose, a positive combination of 551 is obtained, which according to the statistical MPN table is 35 coliforms/ml (presumptive test). Of the gas positive (+) vials, only 4 (A2-A5) were positive by the MUG assay, and therefore most likely contained E. coli (presumptive confirmation for E. coli or fecal coliforms). Among the confirmatory data, the Conventional EC test showed 5 tubes (A1-A5) to be positive for fecal coliforms, whereas by the CAP-EIA method, only 4 vials, (A2-A5) showed positive reactivity. The positive EIA readings ranged from 0.55 up to 1.06, with the negative reaction reading around 0.20. The positive EIA data are strongly supported by the positive MUG data in that both of these tests indicated the presence of fecal coliforms in the same vials, A2 - A5. The fact that vial A1 was negative on MUG and EIA but positive by EC was not surprising because of the incidence of false (+) and false (-) reactions commonly associated with the Conventional MPN Methods (15, 16, 20). The selectivity of the EC medium along with the elevated temperature of incubation has been known to affect the recovery of fecal coliforms. Also, about 7% of the fecal coliforms (E. coli) will not produce gas on EC (false negatives), and 8% of the nonfecal coliforms have been found to be able to grow and produce gas in EC broth (false positives). The EC reaction observed in tube A1 therefore may be a false positive reaction, which would account for its discrepancy with the MUG and EIA results for tube A1. With regard to the other vials (B1 - B5, C1 - C5) in the same experiment, excellent correlation was observed between all the tests. In other words, vials which were MUG (-) were also EC (-) and EIA (-), thus confirming the absence of fecal coliforms in these vials.

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In experiment 2, the water sample used to inoculate the vials was more heavily contaminated, because all presumptive tubes were positive for gas production. Based on the statistical MPN table, a positive combination of 5 5 5 would indicate the presence of greater than 240 coliforms/ml. In comparing the confirmatory test, excellent correlation was obtained between MUG, EC and the antibody based EIA. Every vial that showed fluorescence (MUG +) was also positive by EC as well as EIA. The EIA reactions ranged from slightly weak positive readings of 0.46 (B2) up to very strong reactions of 2.15 observed for C4.

The results of these two experiments indicate that the antibody based CAP - EIA test of the present invention is as efficient as the EPA approved EC test in confirming the presence of fecal coliforms. Additionally, as shown in the case of vial A1 in experiment 1, the CAP-EIA test is less susceptible to false positive or negative reactions due to the specific nature of the antibody-antigen reactions. Furthermore, the CAP-EIA test offers several distinct advantages over the conventional MPN test, namely: 1) simplicity - both the presumptive and the confirmatory test can be done using the same vial without the need for additional transfers or medium; 2) speed - the CAP-EIA test can be completed in 1/3 to 1/2 the time required for a conventional test; and 3) specificity - antibodies are highly specific for their antigenic targets.

These advantages plus the unique cap-well design of the CAP-EIA, which can provide 4 separate pieces of data (acid, gas, fluorescence, and EIA) from the same vial,

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makes it a far superior alternative to the conventional MPN test for analyzing coliforms and fecal coliforms in water and/or food samples.

The concept of doing EIA in a vial or tube cap containing a polystyrene insert is not limited to coliform or fecal coliform assay, and should be equally applicable to the detection of other bacteria.

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Example 26 - Preparation of 2-[3-(4-methyl-2,2'-bipyridine-4'-yl)-propyl]-1,3-dioxolane

5 Under an inert atmosphere of argon, 30 ml of dry tetrahydrofuran (THF) and 7.65 ml of dry diisopropylamine (54.6 mmol) were added to a 3-neck, 600 ml flask via syringe with stirring. The solution was cooled to -78°C by immersing the flask in a mixture of dry ice-isopropanol in a low form beaker. 21.6 ml of 10 2.5M n-butyl lithium (54 mmol) were slowly added to the flask. The resulting solution was stirred for 15 min and a solution of 9.58 g of 4,4'-dimethyl-2,2'-bipyridine (52 mmol) dissolved in 300 ml of dry THF was added dropwise by cannula with stirring over 1 hr.

15 The resulting brown mixture was further stirred at -78°C for 2 hrs, 10 g of 2-(2-bromoethyl)-1,3-dioxolane (55 mmol) were added by syringe and the resulting mixture stirred at -78°C for 5 min. The reaction 20 vessel was then placed in an ice bath (10°C) and after 30 min began to change color (after 1 hr the color was dark violet; after 2 hrs the color was blue; after 2.5 hrs the color was green; and after 3.25 hrs the color was lemon yellow.

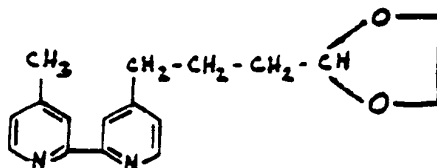
25 The reaction mixture was quenched with 30 ml of saturated NaCl followed by 10 ml of water and 50 ml of ether. The aqueous phase was extracted twice with 300 ml of ether and the combined ether phases were back-extracted with 100 ml of water and dried over anhydrous 30 sodium sulphate.

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To purify the reaction product, the sample was separated on alumina (Merck) 90, activity III, neutral. The eluants used were petroleum ether/diethyl ether (2:1) followed by petroleum ether/diethyl ether (1:1) (the starting material elutes completely with petroleum ether/diethyl ether (2:1), followed by the product).

Proton NMR analysis confirmed that the structure of the isolated reaction product is



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Example 27 -Preparation and Purification of 4-(butan-1-al)-4'-methyl-2,2' bipyridine

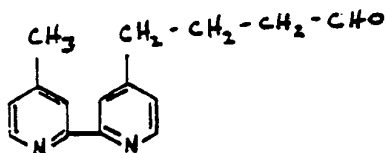
5 2 g of 2-[3-(4-methyl-2,2'-bipyridine-4'-yl)-propyl]-1,3 dioxolane were dissolved in 50 ml of 1N HCl and heated for 2 hrs at 50°C. The solution was cooled, adjusted to between pH 7 and 8 with sodium bicarbonate and extracted twice with 100 ml of chloroform.

10 The combined chloroform phases were washed with a small amount of water, dried over sodium sulfate, filtered and rotoevaporated to yield a yellow oil.

15 The yellow oil was purified on a silica gel column using ethyl acetate/toluene (1:1) as the eluant, the impurity being eluted with methanol.

20 Proton NMR analysis [δ 1.96-2.11 (m, 2H); 2.43 (s, 3H); 2.46-2.50 (t, 2H); 2.53-2.80 (m, 2H); 7.12-7.14 (m, 2H); 8.17-8.21 (br. s, 2H); 8.52-8.58 (m, 2H); 9.89 (s, 1H)] confirmed that the structure of the reaction product is

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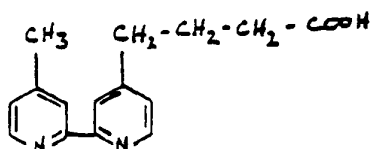
-115-

Example 28 - Preparation of 4-(4-methyl-2,2'-bipyridine-4'-yl) butyric acid

0.5 g of 4-(butan-1-yl)-4'-methyl-2,2'-bipyridine (2.0 mmol) were dissolved in 10 ml absolute acetone. 225 mg of finely powdered potassium permanganate (KMnO_4 ; 1.42 mmol) were added in portions to the solution with stirring. The reaction was followed by thin layer chromatography, (silica; ethyl acetate/toluene 50:50), which indicated that while the aldehyde gradually disappeared a bipyridine of low R_f was formed.

After the reaction reached completion, water was added and the MnO_2 was filtered and washed with small portions of $\text{Na}_2\text{CO}_3(\text{aq.})$. The acetone was rotoevaporated and the residue extracted with CH_2Cl_2 to remove non-acidic bipyridines. The aqueous solution was made acidic by careful addition of 1.0N HCl to pH 4.8. The solution became partially cloudy upon reaching this pH, the suspension redissolving at lower pH. The mixture was extracted five times with equal volumes of CH_2Cl_2 , dried over Na_2SO_4 and rotoevaporated to an oil which promptly solidified in vacuo. The crude solid was recrystallized from chloroform: petroleum ether to obtain white crystals.

Melting point: 103.5°C - 105.5°C ; IR: 1704 cm^{-1} . Proton NMR analysis was consistent with the following structure



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Example 29 - Preparation of bis(2,2'-bipyridine)[4-(butan-1-yl)-4'-methyl-2,2'-bipyridine]ruthenium (II) diperchlorate: Compound I

5 250 mg of ruthenium bipyridyl dichloride dihydrate (0.48 mmol) (Strem) in 50 ml of ethylene glycol were quickly heated to boiling and then immersed in a silicone oil bath (130°C). To the resulting purple-orange solution were added 150 mg of 2-[3-(4-methyl-2,2'-
10 bipyridine-4'-yl)propyl]-1,3-dioxolane (0.53 mmol) in 10 ml of ethylene glycol. The resulting orange solution was stirred at 130°C for 30 min, cooled to room temperature and diluted 1:1 with distilled water.

15 A concentrated solution of sodium perchlorate in water was added to the solution, causing the appearance of a very fine orange precipitate. The mixture was refrigerated overnight, filtered and the precipitate washed with water.

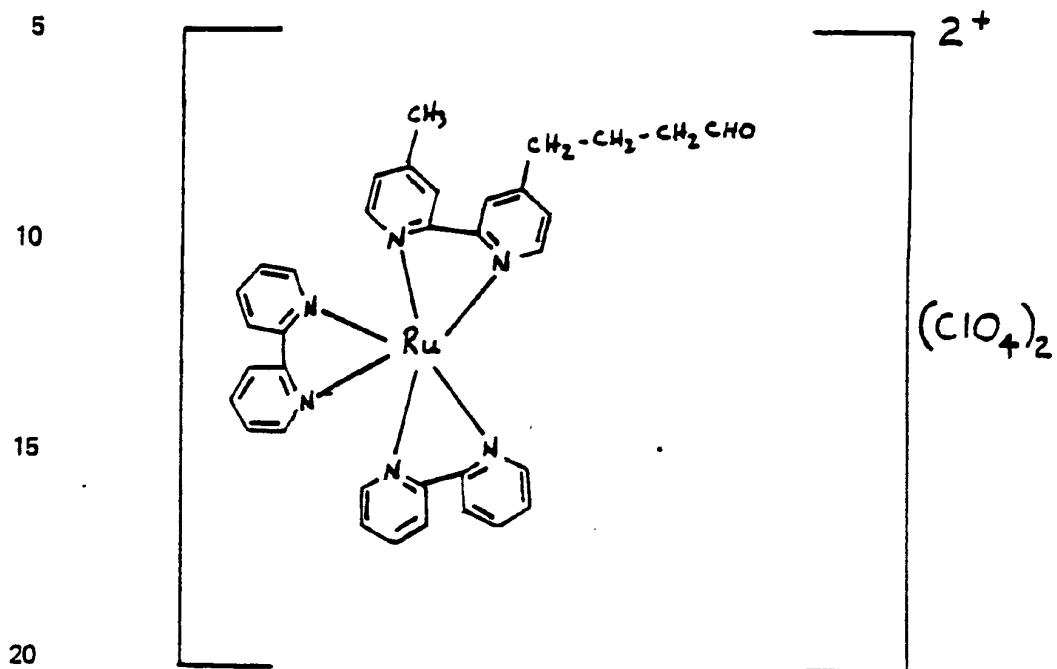
20 The precipitate was dissolved in hot water and recrystallized by adding perchloric acid to the solution to produce bright orange crystals which were then filtered, washed with cold water and dried. This
25 recrystallization procedure was repeated, yielding a total of 150 mg of bright orange crystals.

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NMR analysis indicated the following structure



I

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Within the present application, the above-identified compound is referred to as Compound I.

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Example 30 - Preparation of bis(2,2'-bipyridine)[4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II) dihexafluorophosphate: Compound II

5 134 mg of 4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric
acid (0.52 mmol) were dissolved in 50 ml of water. The
solution was degassed with argon and 250 mg of
ruthenium bipyridyl dichloride dihydrate (Strem) (0.48
10 mmol) were added. The mixture was refluxed under argon
for 4 hrs. The water was rotevaporated and the residue
redissolved in the minimum amount of water and loaded
onto a SP-25-Sephadex ion-exchange column. After elut-
ing impurities with water, the compound was eluted as a
red band with 0.2 M NaCl solution and isolated as a
15 hexafluorophosphate by addition of a saturated aqueous
solution of NH_4PF_6 . The crude product was
reprecipitated twice from hot acetone with diethyl
ether. Anal: Calculated; C, 43.80%; H, 3.36%; N,
8.76%. Found; C, 43.82%; H, 3.54%; N, 8.55%.

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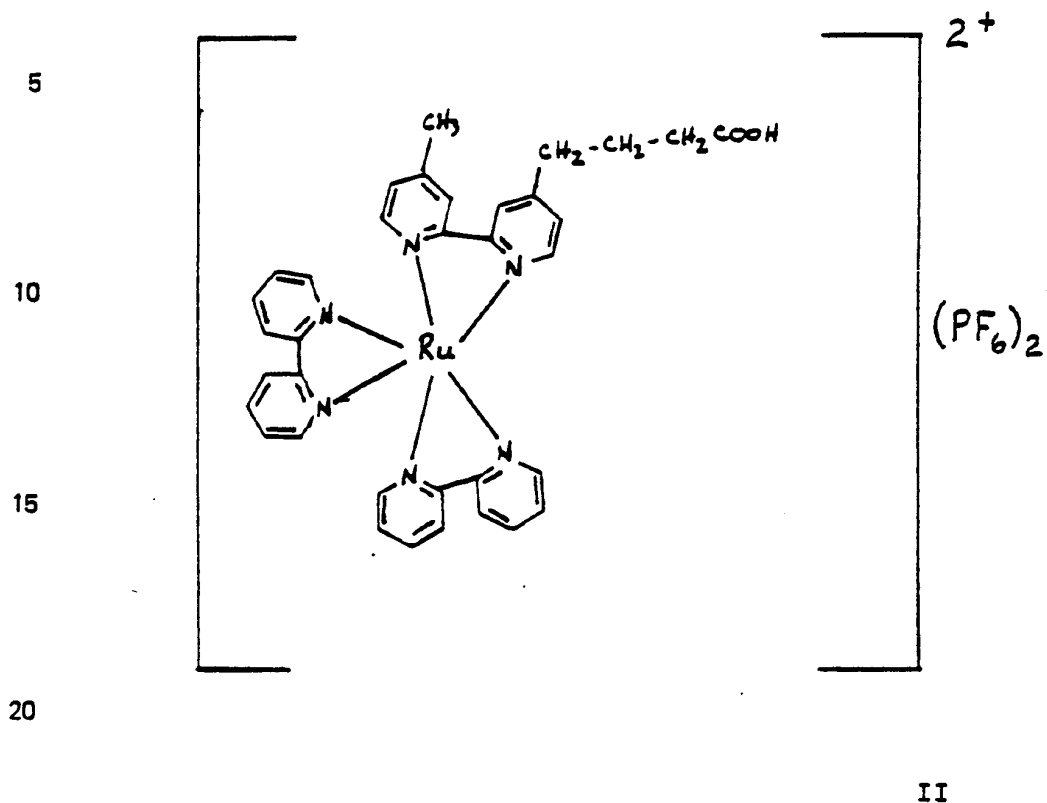
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Proton NMR analysis was consistent with the following structure



II

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Example 31 - Preparation of N-[4-(4-methyl-2,2'-bipyridine-4'-yl)-butyl]-phthalimide

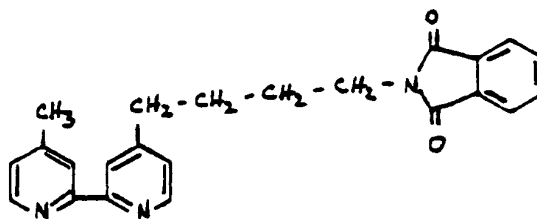
5 Under an inert atmosphere of argon, 30 ml of dry tetrahydrofuran (THF) and 7.65 ml of dry diisopropylamine (54.6 mmol) were added to a 3-neck, 600 ml flask via syringe with stirring. The solution was cooled to -78°C by immersing the flask in a mixture of dry ice-isopropanol in a low form beaker. 21.6 ml of 2.5 n-butyl lithium (54 mmol) were slowly added to 10 the flask. The resulting solution was stirred for 15 min and a solution of 9.58 g of 4,4'-dimethyl-2,2'-bipyridine (52 mmol) dissolved in 300 ml of dry THF was added dropwise by cannula with stirring over 1 hr.

15 The resulting brown mixture was further stirred at -78°C for 2 hrs, 100 g of 1,3-dibromopropane (0.495 mol) were rapidly added and the resulting mixture stirred at -78°C for 1 hr. The mixture was then 20 stirred at room temperature for 2 more hrs. The color of the mixture changed from brown to blue to yellow. Most of the solvent was rotoevaporated and 200 ml of water were added causing the formation of two phases. Concentrated HCl was added to lower the pH to 0. The 25 organic layer was discarded. The aqueous layer was washed twice with 100 ml of ether. The pH was raised to between 1 and 2. A red oil separated and was extracted into CH_2Cl_2 . After drying the solution with anhydrous Na_2CO_3 , most of the CH_2Cl_2 was rotoevaporated and the sample was loaded onto a silica gel column. 30 The sample was eluted with chloroform yielding a light yellow oil which crystallized after cooling overnight (12.37 g).

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4g of the crude 4-(4-bromobutyl)-4'-methyl-2,2'-bipyridine (13.3 mmol) thus prepared were then added to a suspension of 2.46g potassium phthalimide (13.3 mmol) in 60 ml of dimethylformamide (DMF). The mixture was then stirred at approximately 50°C for 2 hrs. 90 ml of CHCl₃ were added to the mixture, followed by 125 ml of water. The CHCl₃ layer was separated and the aqueous layer was extracted twice with 50 ml of chloroform. The combined CHCl₃ layers were washed with 50 ml of H₂O, dried over anhydrous Na₂SO₄, and rotoevaporated, leaving a pale orange oil which solidified overnight. The crude product was recrystallized from acetone/ethanol, yielding 2.21 g (44.5%) of white crystals (melting point, 114.5-117.8°C). Anal:Calculated; C, 74.38%; H, 5.70%; N, 11.31%. Found: C, 73.98%; H, 6.14%; N, 11.28%. IR: Phthalimide carbonyl stretch 1771 and 1704 cm⁻¹. Proton NMR analysis showed aromatic resonances (δ 7.75-7.85, m, 4H) in addition to the usual bipyridyl derivative signals. These data are consistent with the following structure



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Example 32 - Preparation of theophylline-8-butyric-[4-(4-methyl-2,2'-bipyridine-4'-yl)-butyl]amid : Compound III

5 370 mg (1 mmol) of N-[4-(4-methyl-2,2'-bipyridine-4'-yl)-butyl]-phthalimide were slurried in 10 ml ethanol, treated with hydrazine hydrate (720 mg, 1.4 mmol), stirred and refluxed for 4 hrs, during which time all the solid went into solution. Towards the end of the
10 reaction a white precipitate began to form.

After cooling, the reaction mixture was made basic with 50% NaOH and then poured into 100 ml of water. A solution resulted and Na_2CO_3 was added to salt out the
15 product as an oil. The amine was extracted using three 40 ml portions of CH_2Cl_2 . The extracts were dried with CaSO_4 , filtered and evaporated to give the amino-bipyridine derivative as a colorless oil (yield = 0.135 gm; 56%).

20 1.34 g (5.6 mmol) of 4-[4-(1-aminobutyl)]4'-methyl-2,2'-bipyridine and theophylline-8-butyric acid (1.18 g; 4.4 mmol) were dissolved at room temperature in 10 ml of dry pyridine. To the solution were added 1.16 g
25 (5.6 mmol) of dicyclohexylcarbodiimide. Stirring at room temperature was continued overnight. Thin layer chromatography on alumina (mobile phase = 15% methanol/chloroform) revealed one product spot with an R_f of 0.68. Precipitated dicyclohexylurea was removed
30 by filtration and the pyridine was stripped to give a solid, which was triturated in ether and filtered (yield = 2.13 g; 99%).

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Example 33 - Preparation of bis-(2,2'-bipyridine)[theophylline-8-butyric- 4-(4-methyl-2,2'-bipyridine-4'-yl)-butyl amide] ruthenium (II) dichloride: Ru(II)-Compound III Conjugate

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To a mixture of 154 mg (0.296 mmoles) of bis-(2,2'-bipyridine) ruthenium (II) dichloride dihydrate and 175 mg (0.357 mmoles) of Compound III described above were added 40 ml of ethanol/H₂O (1:1, v/v). This mixture was
10 argon degassed in the dark for 15 min and refluxed in the dark under argon for 3 hrs to produce a clear, cherry red solution. The resulting clear, cherry red solution was allowed to cool to room temperature and the solvent was stripped off using a rotary evaporator while
15 maintaining the solution in the dark under a temperature less than or equal to 37°C.

The resulting residue was dissolved in approximately 1-3 ml of methanol, loaded onto a Sephadex LH-20
20 chromatography column (75 cm x 3 cm) and eluted at a flow rate of about 0.4-0.7 ml/min. A bright red band (product) was closely followed by a brown, nonluminescent band (impurity) and two luminescent bands. The red product band was found to be contaminated with a small amount of the material from the
25 brown, nonluminescent band. This contaminating material was separated from the product by running the sample on a second Sephadex LH-20 column under similar conditions.

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The red product was obtained by stripping the solvent off by rotoevaporation. The resulting solid material was dissolved in approximately 1 ml of methanol and reprecipitated in approximately 75 ml of diethyl ether

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to yield an orange powder which was collected by filtration.

Anal: Calculated; C, 54.51%; H, 5.72%; N, 13.99%; O, 10.47%; Cl 6.44. Found; C, 55.04%; H, 6.35%; N, 13.18%; O, 10.62%; Cl, 6.68.

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Example 34 - Modulation of Electrochemiluminescent
Signal Generated By Ru(II)-Comp und III Conjugate Using
Antibodies Specific For Theophylline

5 The Ru(II)-Compound III Conjugate described in Example
33 was diluted to a final concentration of 150 nM using
0.1M phosphate buffer, pH 6.0, containing 0.35 M sodium
10 fluoride (PBF Buffer). Monoclonal antibody (clone
number 9-49, ascites lot number WO399, cat number 046)
specific for theophylline was obtained from Kallestad
Laboratories, Inc. (Chaska, MN). The monoclonal
antibody was diluted to different concentrations using
PBF Buffer (between 21.9 micrograms of protein/ml to
15 700 micrograms/ml).

Another monoclonal antibody (control MAB) that was not
reactive with theophylline was obtained from Sigma (St.
Louis, MO) and was diluted to different concentrations
between 21.9 micrograms of protein/ml to 700
20 micrograms/ml using PBF Buffer. A standard solution
of theophylline was prepared using theophylline
obtained from Aldrich Chemical Co., (Milwaukee, WI, cat
number 26-140-8, M.W. 180.17). Theophylline was dis-
solved in PBF Buffer to give a final concentration of
25 75 micromolar and was diluted with PBF Buffer to 6
micromolar for use in assays. Prior to making
electrochemiluminescence measurements a solution
containing 250 mM oxalic acid and 5% (v/v) Triton-X
100 (EC1 solution) was added to the reaction mixture.
30 Measurements were made using a Berthold luminometer
that was modified to allow the placement of two
platinum gauze electrodes into the test tube
containing the reaction solution. The electrodes were
connected to a potentiostat and the

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electrochemiluminescence measurement was made by sweeping an applied potential across the electrodes from 1.5 to 2.5 volts at a scan rate of 50 mV/sec. The Berthold luminometer used for the measurement had a high gain, red sensitive photomultiplier tube. The luminometer output to the recorder was adjusted to 10^5 counts/volt. The measurements were recorded on an X-Y-Y' recorder and the peak height was used as the measurement of electrochemiluminescence. The electrodes were cleaned between measurements by rinsing with a buffer at pH 4.2 containing 0.1 M phosphate, 0.1 M citrate, 0.025 M oxalic acid, and 1% Triton X-100; pulsing the electrodes in this solution between +2.2 to -2.2 volts for 60 sec; and followed by +2.2 volts for 10 seconds. Next the electrodes were removed from this solution, rinsed in distilled water and wiped dry. The experiment was carried out as outlined in Table XIV.

A solution of control monoclonal antibodies, antibodies to theophylline or PBF Buffer was added to a set of test tubes (Step 1). To the tubes, a solution of theophylline or PBF Buffer was added (Step 2). The solutions were mixed by briefly shaking the test tubes and allowed to react for 25 min at room temperature. Then a solution of Ru(II)-Compound III Conjugate was added to the tubes (Step 3). The test tubes were shaken and kept at room temperature for 15 min. Finally, 100 microliters of the ECL solution was added to each tube and electrochemiluminescence was measured as described above. The results are listed in Table XV.

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Table XIV

Experimental Design for Studying the Effect of
Antibody- Ru(II)-Compound III Conjugate
Interactions on Electrochemiluminescence

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	Step I 100 microliters of:	Step 2 200 microliters of:	Step 3 100 microliters of:
10	A. Control mono- clonal antibody (2.19 micrograms to 70 micrograms)	Buffer	Ru(II)-Compound III Conjugate
	or		
15	B. Anti-theophylline antibody (2.19 micrograms to 70 micrograms)	Buffer or Theophylline	Ru(II)-Compound III Conjugate
	or		
20	C. PBF Buffer	Buffer	Ru(II)-Compound III Conjugate or Buffer
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30			
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TABLE XV

Effect of Antibody and Theophylline
on Electrochemiluminescence of
Ru(II)-Compound III Conjugate

5	Antibody Protein Concentration (micrograms/tube)	Control MFB + Ru(II)-Compound III	Anti-Theophylline MFB + Ru(II)-Compound III	Anti-Theophylline MFB + Theophylline Ru(II)-Compound III
Luminescence Measurement				
10	2.19	55,000	40,000	43,000
		55,000	41,000	57,000
	4.38	57,000	22,500	37,000
		57,000	25,000	36,000
15	8.75	53,000	20,000	33,500
		50,000	22,000	30,500
	35.0	43,000	13,500	17,500
		41,000	14,000	16,000
	70.0	42,000	11,000	11,000
		37,500	12,000	12,500

The electrochemiluminescence of duplicate samples were measured as described above. The electrochemiluminescence of Ru(II)-Compound III Conjugate used in the above study was 57,200 when measured in buffer without the addition of antibody. The background for the buffer mixture was 5750.

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The data show that a monoclonal antibody which specifically recognizes theophylline, when contacted with an analog of theophylline to which a ruthenium compound is attached e.g., Ru(II)-Compound III, will decrease the electrochemiluminescence. The decrease in electrochemiluminescence is proportional to the antibody concentration when the Ru (II)-Compound III Conjugate concentration is held constant. When an antibody is used which does not react with theophylline, only a slight decrease in the electrochemiluminescence is seen at the highest concentration of antibody.

The data also show that when theophylline is contacted with the anti-theophylline antibody and then the Ru(II)-Compound III Conjugate is added to the mixture, the amount of electrochemiluminescence is greater. This demonstrates that theophylline competes for the binding of antibody to Ru(II)-Compound III Conjugate resulting in a greater amount of Ru(II)-Compound III Conjugate which can generate electrochemiluminescence.

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Example 35 - Assay for Theophyllin in Serum Based on a Homogeneous Electrochemiluminescent Immunoassay

5 Based on the results described Example 34, a homogeneous immunoassay for theophylline was developed using antibody to theophylline and the Ru(II)-Compound III Conjugate described in Example 33 in a competitive binding format. The materials used were described in Example 34 except the PBF buffer was 0.1M phosphate
10 buffer, pH 6.0, containing 0.1M sodium fluoride. For this assay, a specific concentration of monoclonal antibody to theophylline was chosen. The antibody concentration was 55 micrograms/ml. The Ru(II)-Compound III Conjugate concentration was adjusted to 175 nM. Theophylline was added to human serum to give
15 final concentrations of 2.5, 5, 10, 20 and 40 micrograms of theophylline/ml of serum.

The assay was performed by adding 10 microliters of
20 serum to 290 microliters of anti-theophylline monoclonal antibody and holding the solution at room temperature for 25 min. Then 100 microliters Ru(II)-Compound III Conjugate were added to each tube to give a final concentration of 35 nM and holding this
25 solution at room temperature for 15 min. 100 microliters of the ECL solution described in Example 34 were then added to each tube and electrochemiluminescent properties of the solutions were measured as previously described using a sweep
30 mode for 1.5 volts to 2.5 volts at 50 mV/sec. The data are shown in Figure 2 and demonstrate that there is a correlation between the concentration of theophylline in a serum sample and the amount of electrochemiluminescence that is emitted by the

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reaction mixture. This observation demonstrates that it is possible to develop an assay for theophylline.

Based on these results, one skilled in the art would be able to develop a homogeneous electrochemiluminescence immunoassay for detecting and quantifying an analyte of interest in a biological matrix.

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Example 36 - Assay for Theophylline in Hemolyzed, Lipemic, Icteric and Normal Serum Samples Based on a Homogeneous Electrochemiluminescent Immunoassay and Comparison to a Fluorescence Polarization Assay

5 The concentration of theophylline in different types of serum samples was determined using a homogeneous electrochemiluminescent immunoassay. The format for the assay was a competitive binding assay using a
10 monoclonal antibody specific for theophylline and the Ru(II)-Compound III Conjugate described in Example 33. The reagents and methods for electrochemiluminescence are described in the previous example.

15 The fluorescence polarization assay used to measure the concentration of theophylline in the different serum samples was carried out using an automated TDX instrument from Abbott Laboratories (North Chicago, IL). Hemolyzed, lipemic, icteric and normal sera were
20 used in the assays and data for the abnormal sera are listed in Table XVI below.

Table XVI

25 Homogeneous Theophylline Assay
Characteristics of Potentially Problematic Sera

	<u>Serum</u>	<u>Factor Concentration</u>	<u>Normal Range</u>
30	Hemolyzed	12.4 mg/dl hemoglobin	0-3.2 mg/dl
	Lipemic	405 mg/dl triglycerides 196 mg/dl cholesterol	10-190 mg/dl 120-200 mg/dl
	Icteric	10 mg/dl bilirubin	0-1.5 mg/dl

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Different amounts of theophylline were added to the serum samples to give final concentrations between 2.5 micrograms theophylline/ml and 40 micrograms theophylline/ml. The results for the homogeneous electrochemiluminescent immunoassay are displayed in Figure 3.

Each serum sample was also analyzed for the concentration of theophylline by a fluorescence polarization assay. The concentration of theophylline measured by the homogeneous electrochemiluminescence immunoassay and the fluorescence polarization assay were compared. The data were plotted as a scattergram and are shown in Figures 4A-D. The data points were analyzed by linear regression and the correlation coefficients were calculated. The analysis demonstrates an excellent correlation between the two assays. The correlation coefficients (r) were between 0.98 and 1.00. The slopes of the curves for normal, hemolyzed, and lipemic serum samples were between 0.8 and 1.2, demonstrating excellent recovery of theophylline from these serum samples.

Although the electrochemiluminescence emitted by the icteric serum samples containing theophylline was higher than for the other serum samples, it was proportionally higher at each theophylline concentration. This can be seen in Figure 4D. The correlation coefficient is 1.00 for the data points comparing electrochemiluminescence and fluorescence polarization; however, the slope is 2.14, demonstrating higher recovery for the theophylline in the icteric serum sample.

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Based on these results, the concentration of theophylline in an icteric sample may be determined by establishing a standard curve for the sample by adding known amounts of the Ru(II)-Compound Conjugate to aliquots of the icteric serum. These data demonstrate that a homogeneous electrochemiluminescent immunoassay may be used to measure the concentration of theophylline present in serum samples containing abnormal levels of hemoglobin, lipid and bilirubin.

A homogeneous electrochemiluminescent immunoassay offers advantages over a fluorescence polarization method because of the versatility of ECL detection, e.g., more sensitive detection at higher concentrations of biological molecules.

A homogeneous electrochemiluminescent immunoassay offers further advantages over a fluorescence polarization method because no incident light source is required; electrical excitation being the only requirement for efficient light-generation. Consequently, no sophisticated optics are necessary. Since the measurement principle is purely specific photon emission induced by electrochemical stimulation, the sensitivity of the system is potentially much greater than fluorescence polarization and a wider dynamic range will be achievable. Also, measurement of a much greater variety of analytes is possible with a homogeneous electrochemiluminescent immunoassay than is provided by the fluorescence polarization technique, due to the selective modulation of electrically-stimulated chemiluminescence by biomolecular recognition events, e.g., antibody-antigen interactions.

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Based on these results, one skilled in the art would know that homogeneous electrochemiluminescent immunoassays for detecting other analytes of interest in abnormal serum samples may be developed.

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Example 37 - Assay for Theophylline in Serum Based on a Homogeneous Electrochemiluminescence Immunoassay and Comparison to a High Pressure Liquid Chromatographic (HPLC) Method

5 Different amounts of theophylline were added to human serum samples to give final concentrations between 2.5 micrograms theophylline/ml and 40 micrograms theophylline/ml. Each sample was then divided into two aliquots and the concentration of theophylline in the sample was determined by a homogeneous electrochemiluminescence immunoassay and compared to the results obtained for the same serum samples using an HPLC method. The format for the homogeneous electrochemiluminescence immunoassay was a competitive binding assay using a monoclonal antibody specific for theophylline and the Ru(II)-Compound III Conjugate. The reagents and methods for this assay are described in a previous example. The HPLC method used to measure the concentration of theophylline in different serum samples is described as follows.

25 Theophylline (1,3-dimethylxanthine) was separated from serum proteins by precipitation of the latter with acetonitrile. The supernatant fluid containing theophylline was run on an HPLC system equipped with a Waters Associates Micro Bondapak C18 column, (3.9 mmx30 cm). The chromatogram was completely resolved in less than 10 min.

30 The following reagents were used: sodium acetate (reagent grade), deionized water (purified by the Millipore Milli Q system), acetonitrile (HPLC grade) and theophylline standard, (Sigma). The solvent used

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for precipitating the serum proteins was a 20 mM sodium acetate buffer, pH 4.0, containing 14% (v/v) acetonitrile. The HPLC mobile phase buffer was 10 mM sodium acetate buffer, pH 4.0, containing 7% (v/v) acetonitrile. The flow rate was 1.5 ml/min, and the eluant was monitored by a UV spectrophotometer set at 270 nm. The sensitivity of the UV absorbance detector was set at 0.02 Absorbance Units Full Scale (AUFS). The ambient temperature ranged typically between 22°C and 24°C.

The results for the homogeneous electrochemiluminescent immunoassay and the HPLC assay for determining the concentration of theophylline in serum are shown in Figure 5. The data were plotted as a scattergram and the data points were analyzed by linear regression. The correlation coefficient was calculated. The correlation coefficient (r) was 0.98, which demonstrates excellent correlation between the two assays.

The slope of the curve was 1.197, demonstrating excellent recovery of the theophylline from the serum sample for the homogeneous electrochemiluminescent immunoassay compared to a standard method based on HPLC. The homogeneous electrochemiluminescence assay offers advantages over the HPLC method because of the speed, sensitivity and ability to easily handle multiple samples. Based on these results, one skilled in the art would know that homogeneous electrochemiluminescent immunoassays for detecting analytes of interest, which may be detected by HPLC and similar methods, may be developed.

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Example 38 - Preparation of Theophylline-Bovine Serum
Albumin Conjugates

5 Theophylline-bovine serum albumin (BSA) conjugates were
prepared from theophylline-3-methyl-butyric acid,
theophylline-8-butyric acid and theophylline-7-acetic
acid by the following procedure. 50 mg of BSA were
dissolved in 1.5 ml of 0.15M NaHCO₃, pH 9.0.
10 Separately, 16 mg of ethyl 3'3-dimethyl amino propyl
carbodiimide hydrochloride (EDCI), 11 mg N-hydroxy
succinimide (NHS) and either 17.8 mg of theophylline-7-
acetic acid, 20 mg of theophylline-8-butyric acid or
20.9 mg of theophylline-3-methyl-butyric acid dissolved
15 in dimethylsulfoxide were added and the solution heated
at 45°C for 1-2 hrs. The solution was added to the BSA
solution dropwise and allowed to react for 1 hr.
Theophylline conjugates of BSA were purified by gel
filtration chromatography on a Sephadex G-25 column
(1.6 cm x 38 cm) using 0.15 M PBS/0.1% azide, pH 7.4,
20 as the mobile phase and a flow rate of 30 ml/hr.

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Example 39 - Preparation of Theophylline-BSA Biomag[•] Particles

5 A 4 ml volume of Biomag[•]-amine particles (Advanced Magnetic, Inc., Cambridge, MA) was washed 2-3 times in separate T-flasks with 20 ml of phosphate buffered saline (Sigma) (PBS), pH 7.4, containing 0.008% Nonidet P-40 (NP-40). To the Biomag[•] wet cake, 10 ml of 5% glutaraldehyde (Sigma) in PBS was added and activation
10 was allowed to proceed for 3 hrs using a rotary mixer. The activated Biomag[•] particles were washed as described above for a total of 4 washes and transferred to a T-flask.

15 6.8 mg of theophylline-BSA prepared as described in Example 38 in 10 ml of PBS/NP-40 were added to the activated Biomag[•] wet cake. The reaction was allowed to proceed overnight at 4⁰C with mixing.

20 The activated Biomag[•] wet cake was washed 3 times with 20 ml of 1% BSA/0.15M PBS/0.1% azide (pH 7.4), the first wash lasting for approximately 30 min using a rotary mixer.

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Example 40 - Preparation of Compound I-Anti-Theophylline Conjugate

5 1.1 ml of a mouse anti-theophylline monoclonal antibody (Kallestad, lot no. WO399; 4.6 mg/ml) were centrifuged at 10,000 g for 8 min. The buffer was exchanged with 0.2M NaHCO₃, 0.15 M NaCl, with azide, pH 9.0, using an Amicon Centricon 30 concentrator (centrifuged at 3000 x g). The antibody solution was diluted to 2 ml and 10 3.2 mg of Compound I were added. The reaction was allowed to proceed for 2 hrs at room temperature with stirring and 79 microliters of aqueous NaBH₄ at 1 mg/ml was added and the solution stirred for 30 min.

15 The resulting solution was loaded onto a Sephadex G-25 (1.0 cm x 18.0 cm) previously equilibrated with tris buffer and eluted at a flow rate of 15 ml/hr. The fractions containing the Compound I-anti-theophylline conjugate were pooled, transferred to dialysis tubing, 20 and dialyzed against 0.15M phosphate/0.15 M NaF (PBS), pH 6.9 (4 l).

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Example 41 - Assay for theophylline in serum based on a heterogeneous electrochemiluminescent assay

5 Using an immunometric assay format, a heterogeneous assay for theophylline was developed using a Compound I labeled anti-theophylline antibody and theophylline BSA immobilized on Biomag[®] magnetic particles. The antibody concentration was 20 micrograms/ml. The magnetic particle concentration was 1% solids (wt/vol).
10 Theophylline was added to a final concentration of 10 and 40 micrograms/ml of serum. The theophylline serum standards were diluted 1000 fold in PBF Buffer (sodium phosphate buffer, pH 7.0, 0.1 M sodium fluoride) containing 0.1% BSA.

15 The assay was performed by the addition of 75 microliters of the diluted serum standards to 75 microliters of antibody conjugated to Compound I and incubating the solution at room temperature for 20 min. Then 50 microliters of the theophylline-BSA-Biomag[®] particles were added and the suspension was allowed to stand for 5 min. The particles were separated magnetically and 100 microliters of the supernatant was measured for electrochemiluminescence
25 as described in Examine 48.

	<u>Theophylline Concentration</u> micrograms/ml ⁺⁺	<u>ECL*</u>	<u>SD</u>	<u>%CV</u>
	0	8,758	81	0.9%
30	10	11,078	368	3.0%
	40	14,106	674	4.8%

* ECL counts per 10 seconds

⁺⁺ corrected for dilution

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Based on these results, one skilled in the art would be able to develop a heterogeneous electrochemiluminescence immunoassay for other analytes of interest in a biological matrix.

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Example 42 - Preparation of a Compound II-Digoxigenin Conjugate

100.2 mg (0.104 mmol) of Compound II, 41 mg (0.105 mmol) of digoxigenin (Sigma) and 28 mg (0.136 mmol) of 1,3-dicyclohexylcarbodiimide (DCC) were added to a 50 ml round bottom flask equipped with a stirbar. To the resulting mixture were added 8-10 ml of anhydrous pyridine (Aldrich Sure-Seal) using a syringe with an 18 gauge needle. The flask was stoppered, sealed with Teflon tape and the contents gently stirred to yield a red solution. The flask was allowed to stand in the dark with stirring in the hood for 24 hrs, at which time 6 mg (0.015 mmol) of digoxigenin and 20 mg (0.097 mmol) of DCC were added. The solution was capped and allowed to stir at room temperature in the dark. The next day no evidence of dicyclohexyl urea precipitation was observed. An additional 18 mg (0.05 mmol) digoxigenin and 103 mg (0.50 mmol) DCC were added. The flask was resealed and stirring continued in the dark. After 72 hrs an additional 103 mg of DCC were added and the solution stirred in the dark for 3 hrs.

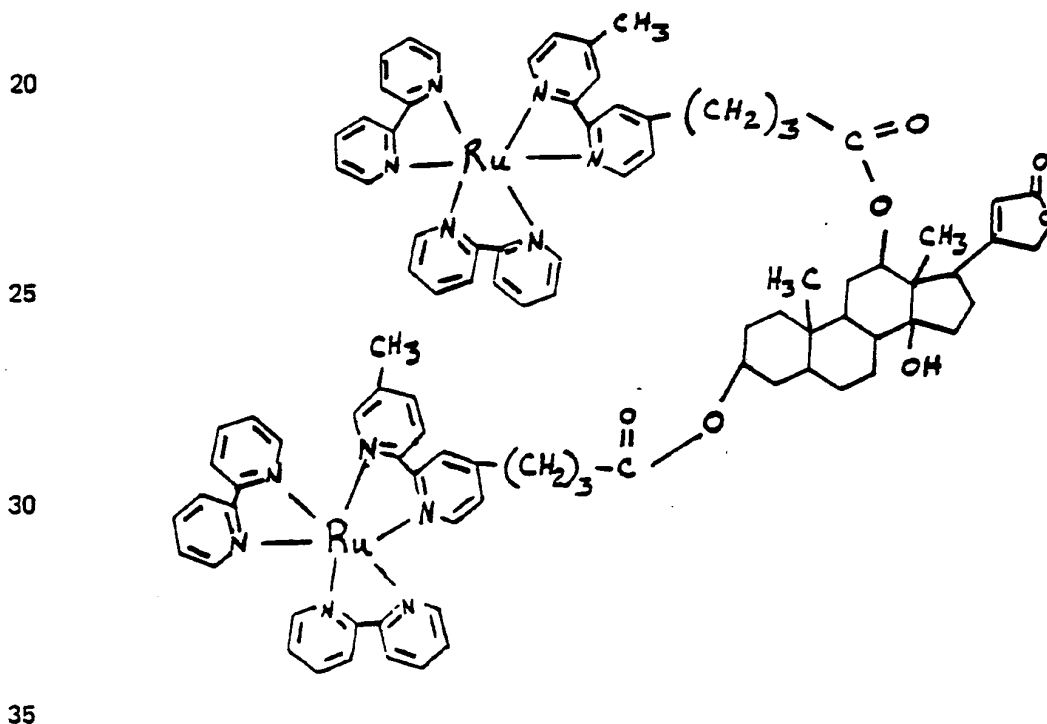
5-10 drops of H₂O were added to the solution, which was then stripped to dryness on a rotary evaporator in the dark to produce a red solid.

The resulting red solid was covered with aluminum foil and dried overnight under vacuum over CaSO₄ in a dessicator. The dried solid was then dissolved in 3-5 ml of methanol and approximately 0.25 g of anhydrous solid LiClO₄ were added to the mixture. Once the LiClO₄ was dissolved, the solution was loaded onto a Sephadex LH-20 column (75 cm x 19 mm) and eluted with methanol at a flow rate between 7-10 sec/drop.

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Three bands were noted as the chromatography proceeded; a pale orange (red luminescent) first band (Fraction 1); a dark red second band that represented the major portion of the reaction (Fraction 2); and a third band which trailed the first two bands (probably unreacted starting material) which was discarded.

Because bands 1 and 2 just barely separated on the column, any orange product in band 2 was isolated by dripping a methanol solution of Fraction 2 (15 ml) into approximately 300 ml of dry diethyl ether (stirred). The resulting precipitate was collected by suction filtration on a 15 ml medium frit and washed five times with 15 ml of diethyl ether. Residual ether was removed by drying the complex overnight over CaSO_4 in a vacuum desiccator. This solid material was determined to be



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Example 43 - Electrochemiluminescence (ECL) of Compound
II-Digoxigenin Conjugate: Modulation of ECL Signal by
anti-digoxin antibody

Monoclonal antibody to digoxin was diluted to the
following concentrations in immunoassay buffer:

1, 10, 50, 200, 400 micrograms/ml

Compound II-Digoxigenin Conjugate (50 micromolar) was
diluted to 150 nM in immunoassay buffer (0.1M phosphate
buffer, pH 6.0, containing 0.1M sodium fluoride).

To 200 microliters of immunoassay buffer in
polypropylene tubes (12 mm x 75 mm) were added 100
microliters of various concentrations of digoxin
antibody and 100 microliters of Compound II-Digoxigenin
conjugate (150 nM). Tubes were mixed on a vortex and
incubated at room temperature for 15 min. Following
incubation, 100 microliters of ECL solution (previously
described) were added and electrochemiluminescence was
measured.

Results:

	Specific Antibody Concentration microgram/tube	\bar{x} ECL Signal
25	0.1	108000
	1	114000
	5	82500
	10	64000
	20	52000
30	40	36000

Total ECL Counts for 30 nM Compound II-Digoxigenin Conjugate =
113666 (peak height)

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At a concentration of 40 micrograms/tube, non-specific antibody modulation of signal was 67,000 counts compared to 36,000 counts in the presence of a specific antibody to digoxin.

5 As can be seen from Figure 6, an increasing
concentration of anti-digoxin antibody when reacted
with a fixed concentration of Compound II-Digoxigenin
Conjugate showed increasing modulation of the
10 electrochemiluminescent signal. This characteristic
may be used advantageously to develop a homogeneous
electrochemiluminescence based assay for the
measurement of digoxin in serum or plasma.

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Example 44 - Homogeneous Digoxin Assay

Based on the results described in Example 43, a homogenous electrochemiluminescent immunoassay for digoxin may be developed using antibody to digoxin and the Compound II-Digoxigenin conjugate using a competitive binding assay format. The reagents which may be used have been described in Example 43. For this assay, a specific concentration of monoclonal antibody to digoxin would be chosen. The antibody concentration may be between 75 to 100 micrograms per ml. The Compound II-Digoxigenin Conjugate concentrations may be between 5-15 nM (Final Concentration).

Digoxin Standard would be added to human serum to give a final concentration of 0.1, 0.5, 1, 2, 4, 8 and 16 nanograms of digoxin per ml of serum.

The assay may be performed by adding 10-30 microliters of serum to 300 microliters of anti-digoxin monoclonal antibody and holding the solution at room temperature of 30 min. Then 100 microliters of the Compound II-Digoxigenin conjugate may be added to each tube to give a final concentration within the range of 5 to 15 nM and incubating the solution at room temperature for 20 min. 100 microliters of the ECL solution previously described may be added to each tube and ECL may be measured as previously described.

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Example 45 - Preparation of Oubain-Bovine Serum Albumin Conjugate

5 50 mg of oubain octahydrate (Aldrich) was dissolved in
5 ml of deionized H₂O. 81 mg of NaIO₄ was added to
the dissolved oubain and the mixture was incubated for
2 hrs at room temperature. The reaction was stopped by
10 passing the mixture over a Dowex 1X-8 ion exchange
resin column (5 ml) which had been equilibrated with
deionized H₂O until the pH was between 5 and 6. The
oxidized oubain fraction was collected when the drops
entering the waste container showed signs of mixing.

15 100 mg of crystalline, lyophilized bovine serum albumin
(Sigma) (BSA) was dissolved in 5 ml of 0.1 M potassium
phosphate buffered saline, pH 7.4, containing 0.05%
azide. The oxidized oubain solution was added dropwise
to the BSA solution with stirring. The resulting
20 solution was allowed to react for 1 hr at room
temperature before adding 30 mg of NaCNBH₄ (Aldrich).
The solution was then stirred overnight at room
temperature.

25 The solution was concentrated (11 ml to 4 ml) using
polyethylene glycol-8000 and free oubain and excess
borohydride were removed from the solution by gel
filtration on Sephadex G-25 (column = 0.6 cm x 37 cm;
eluant = 0.1 M K₂PO₄/0.15 M NaCl, pH 7.5, 0.05% NaN₃).
Fractions 11-17 were pooled and the protein
30 concentration determined by measuring absorbance at
280nm (after dilution).

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Example 46 - Preparation of Oubain-BSA-Sepharose

2 g of cyanogen bromide activated Sepharose 4B (Pharmacia) was washed with 400 ml of 1M HCl in 50 ml portions on a sintered disk funnel. The resin was then washed with 20 ml of 0.1M NaHCO₃, 0.5M NaCl buffer, pH 9.0 (coupling buffer). After transferring the resin to a polypropylene container, 30 mg of oubain-BSA dissolved in 15 ml of coupling buffer were added. The activated resin was allowed to react with the oubain-BSA for 2 hrs with rotary mixing. The remaining activated sites were reacted with 7 ml of 0.5M ethanolamine (pH 8.0) for 2 hrs at room temperature. Using a sintered disk funnel, the resin was subjected to 100 ml washes with each of the following solutions: coupling buffer; 0.15M PBS; 1M HCl; coupling buffer; 0.2% NP-40 in PBS; and PBS. The resin was resuspended to 11 ml and a sufficient amount of this suspension was added to a 1.0 cm inside diameter column (Pharmacia) to give a total bed volume of 3.5 ml.

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Example 47 - Preparation and Affinity Purification Of
An Anti-Digoxin-Compound I Conjugate

5 450 microliters of a 1 mg/ml stock solution of a mouse
anti-digoxin monoclonal antibody (Cambridge Medical
Diagnostics, cat. no. 200M-014, lot no. MA 2507F) were
concentrated to 100 microliters using an Amicon
Centricon 30[•] concentrating unit. To this concentrate
10 were added 900 microliters of 0.2 M sodium bicarbonate
buffer, pH 9.6 and 0.6 mg of Compound I. The reaction
(amidation) was allowed to proceed at room temperature
for 2 hours before 30 microliters of 1.0 M NaBH₄ (aq)
(Sigma) was added. The resulting solution was allowed
to stand for 1 hour.

15 Excess Compound I and other products were separated
from the antibody conjugate by Sephadex G-25
chromatography (column = 1.0 cm_{ID} x 18 cm; eluant = 0.1
M phosphate buffered saline). The sample was
20 fractionated in 1 ml portions at a flow rate of 20
ml/hr and the absorbance at 280 nm monitored at 2.0
AUFS and 0.5 AUFS.

25 Fractions 5, 6, 7 and 8 were pooled, loaded onto a
prewashed ouabain-BSA-Sepharose affinity column (3.5 ml
column with 1 cm diameter), and eluted at a flow rate
of 15 ml/hr. After unretained material was eluted, the
flow rate was increased to 30 ml/hr until 20 ml of
eluate were collected. The saline concentration of the
30 mobile phase were increased to 0.5M and another 20 ml
were eluted through the column.

Oubain specific anti-digoxin-Compound I conjugate was
removed by addition of 4 M and 6 M KSCN. Fractions
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corresponding to anti-digoxin-Compound I conjugate
wer pooled and dialyz d against 10 mM phosphate
buffered saline (4 l; pH 7.4) followed by 0.1 M
phosphate buffer (4 l; pH 7.0).

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Example 48 - Heterogeneous electrochemiluminescent immunoassay for digoxin

5 10 mg of solid digoxin were dissolved in 10 ml of DMSO:H₂O (8:2), to give a digoxin concentration of 1 mg/ml (hereinafter Stock Standard).

10 Working standards were prepared from the Stock Standard to the following concentrations in 0.15 M phosphate buffer, pH 7.0, containing 0.1% BSA and 0.15 M NaF (hereinafter ECL Buffer): 80 ng/ml, 40 ng/ml, 20 ng/ml, 10 ng/ml, 5 ng/ml and 0 ng/ml.

15 75 microliters of anti-digoxin-Compound I conjugate (diluted 1:90) and 75 microliters of the each standard were pipetted into a class tube, mixed on a vortex and incubated at room temperature for 20 min.

20 50 microliters of prewashed ouabain-BSA-Biomag particles were added to each tube, mixed on a vortex and incubated at room temperature for 5 min. Biomag particles were separated and supernatant was transferred to a separate tube.

25 100 microliters of supernatant were mixed with 400 microliters of 0.125 M potassium phosphate 0.125 M citric acid; 32 mM oxalic acid; 1.25% Triton X-100 in a tube.

30 The sample was placed into a Berthold instrument and the electrochemiluminescence was measured as previously described except the procedure was modified by stepping the applied potential from open circuit to 2.2V and integrating the photon counts for 10 sec.

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The electrode was cleaned between measurements using phosphate-citrate buffer as follows:

(a) Pulse electrode using 3 sec intervals alternating between -2.2V and +2.2V for 1 min.

(b) Poise the electrode at +2.2V for 10 seconds.

(c) Rinse electrode with deionized water H_2O and blot dry.

The results are shown in Figure 7.

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Example 49 - Labelling DNA with an electro-
chemiluminescent moiety

5 The following two methods have been used to label DNA
with an electrochemiluminescent moiety.

Synthesis A

10 1.0 A₂₆₀ of the custom synthesized 38 mer (MBI 38)

TCACCAATAAACCGCAAACACCATCCCGTCCTGCCAGT*

where T* is thymidine modified at carbon 5 with

15 $-\text{CH}=\text{CH}-\text{CO}-\text{NH}-(\text{CH}_2)_7-\text{NH}_2$

20 were dissolved in 100 microliters of 0.01 M phosphate
buffer, pH 8.7. 100 microliters of a solution of
bis(2,2'-bipyridine)[4-(butan-1-yl)-4'-methyl-2,2'-bi-
pyridine]ruthenium (II) diperchlorate (Compound I)
(2.3 mg in 300 microliters of 0.01 M potassium
phosphate buffer, pH 8.7). The contents were stirred
and allowed to stand at room temperature overnight.

25 100 microliters of a saturated aqueous solution of
sodium borohydride was added to the mixture to convert
the reversible imine Schiff's base linkage into non-
reversible amine linkage. The reaction was allowed to
30 run at room temperature for 2 hrs. The solution was
then treated carefully with a few drops of dil. acetic
acid to quench excess of sodium borohydride. The
reaction solution was loaded onto a P-2 gel filtration
column (18 inches x 1/2 inch) which had been

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preequilibrated with 0.1 M triethylammonium acetate, pH 6.77. The column was eluted with the same buffer and 2 ml fractions were collected at a flow rate of 20 ml/hr. DNA eluted in fractions 9 and 10 were well separated from unreacted ruthenium bipyridyl complex. The collected DNA sample exhibited typical UV absorption and additionally showed a fluorescent emission spectrum at 620 nm when excited at 450 nm. The fluorescent emission shows the presence of the ruthenium bipyridyl moiety in the DNA sample. The product travels as a single orange fluorescent band on polyacrylamide gel electrophoresis. The electrophoretic mobility of the labeled DNA (MBI 38-Compound I Conjugate) is approximately the same as the unlabeled DNA.

Synthesis B

The ruthenium complex was first converted into an N-hydroxysuccinimide derivative by dissolving 3 mg in 60 microliters of anhydrous dimethylformamide and treating it with a solution of N-hydroxysuccinimide (52 mg) in 200 microliters of anhydrous DMF in the presence of 94 mg dicyclohexylcarbodiimide (DCC). The reaction was allowed to proceed for 4 hrs at 0°C. Precipitated dicyclohexylurea was removed by centrifugation and the supernatant (200 microliters) was added to the solution of amino-linked DNA (described in Synthesis A) in .01 M phosphate buffer pH 8.77 (2A₂₆₀ in 100 microliters of buffer). The reaction was allowed to proceed overnight at room temperature. A considerable amount of solid appeared in the reaction which was removed by filtration through glass wool. The filtrate was concentrated and dissolved in 0.5 ml of 1 M triethylammonium acetate (pH 6.8). The reaction mixture was then chromatographed as described in

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Synthesis A. The labeled DNA exhibited all spectral and electrophoretic characteristics as discussed for the material prepared in Synthesis A.

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Example 50 - Electrogenerated Chemiluminescent
Properties of labeled DNA

5 The labeled DNA sample from Example 49, Synthesis A
(MBI 38-Compound I) was used to study its
electrochemiluminescent properties. Various concent-
rations of labeled DNA were dissolved in 0.5 ml of 0.1
M phosphate buffer, pH 4.2, containing 0.1 M citrate,
10 25 mM oxalate and 1.0% Triton X-100 and measured on a
modified Berthold luminometer. Figure 8 shows the
response of the electrochemiluminescent signal to
various DNA concentrations.

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Example 51 - Hybridization studies of Compound I-labeled oligonucleotide

5 The complementary strand to the 38 mer described in Example 50 was synthesized using the ABI model 380 B DNA Synthesizer and was designated MGEN-38.

10 To determine if the covalent attachment of Compound I to the oligonucleotide affected the hybridization properties of the MBI 38 oligonucleotide, the following experiment was devised. Various concentrations of the target fragment (MGEN-38) were spotted on a sheet of Gelman RP nylon membrane, fixed and probed with either MBI 38 or MBI 38-Compound I. Both fragments were
15 treated with T4 polynucleotide kinase and gamma ^{32}P [ATP] and labeled with ^{32}P at the 5' end. The hybridization sensitivities of DNA and Compound I-labelled DNA were then compared.

20 Concentrations of MGEN-38 DNA, ranging from 50 ng down to 0.05 ng, were spotted on a nylon membrane and allowed to air dry. Duplicate membranes were set up. The blots were treated for 2 min each in : 1.5M NaCl-0.5M NaOH to fully denature the DNA; 1.5M, NaCl-0.5M
25 TRIS to neutralize the blot, and finally in 2X SSC. The blot was baked in a vacuum oven at 80°C for 2 hrs.

The hybridization probe was prepared as follows: 3 micrograms of MBI 38 and MBI 38-Compound I were kinased
30 with 10 units of T4 kinase and 125 microcuries of gamma ^{32}P -ATP. The percentage of isotope incorporation into DNA was determined and shown below.

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MBI 38 total count 4.1×10^6 cpm/microliter
incorporated counts 3.1×10^5 cpm/microliter
% incorporation = 75.6%

5

MBI 38-Compound I total count 3.2×10^6 cpm/microliter
incorporated count 2.6×10^5 cpm/microliter
% incorporation = 81.2%

10

Prehybridization and hybridization solutions were prepared according to Maniatis (24). Blots were prehybridized for 4 hrs at 53°C with 50 micrograms/ml of calf thymus DNA. The blots were then placed in hybridization solution containing the respective probes at 10,000,000 cpm, and allowed to hybridize overnight (12 hrs) at 53°C. The following day, the blots were washed as follows:

15

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- twice with 2 X SSC + 0.1%SDS at 53°C for 15 minutes each wash

- twice with 0.2 X SSC + 0.1%SDS (same as above)

25

- twice with 0.16 X SSC + 0.1%SDS (same as above)

The blots were then air dried and exposed to Kodak X-omat[®] film at -70°C.

30

Analysis of the X-ray (see Figure 9) showed that very similar hybridization patterns were observed between the MBI 38 and MBI 38-Compound I probe. In both cases

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hybridization of prob to 0.5 ng of target was observed, and faint traces of hybridization were observed down to 0.05 ng of target DNA. No hybridization activity by the probe was detected for the negative control DNA (phage lambda DNA spotted at 50 ng).

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Example 52 - Compound I-labeled DNA probe hybridization
specificity study

5 Genomic DNA from several E. coli and non-E. coli
strains were isolated according to methods described by
Maniatis (24). The organisms used were:

E. coli strain EC8 - natural isolate
E. coli strain PC1A - enteropathogenic strain
10 E. coli strain 10H07 - enterotoxigenic strain
E. coli strain EC50 - natural isolate
E. coli strain B - lab strain
E. coli strain K12 - lab strain

15 Enterobacter aerogenes
Citrobacter freundii
Salmonella paratyphi B
Salmonella potsdam

20 DNA from the above-mentioned strains were spotted in
duplicate in 3 micrograms aliquots on a Gelman RP
nylon membrane and on a S & S nitrocellulose membrane.
For negative control, 50 ng of phage lamda DNA were
spotted. Positive control consisted of various
25 concentrations of the complementary strand, MGEN-38,
ranging from 50 ng down to 0.5 ng. The blots were
prepared and treated as described in Example 51.

30 The probing DNA consisted of 3.0 micrograms of the MBI
38-Compound I fragment which was T4 kinased with 125
microcuries of gamma ³²P-ATP to incorporate
radioactivity. The following amount of activity was
incorporated.

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MBI 38-Compound total counts - 3.35×10^6 cpm/microliter
incorporated counts - 1.50×10^6 cpm/microliter
% incorporation - 44.7%

5 For this assay, 2,600,000 cpm of activity was used for
probing each filter. The hybridization solutions,
conditions, washing solutions and protocols were the
same as those described in Example 51.

10 The X-ray film of the blot (Figure 10) shows that both
the positive and negative controls reacted accordingly.
No hybridization activity was detected with lambda DNA
(50 ng), and strong hybridization was observed for the
complementary sequence, MGEN-38, down to 0.5 ng
15 concentration. These findings are in total agreement
with the results of the sensitivity study.

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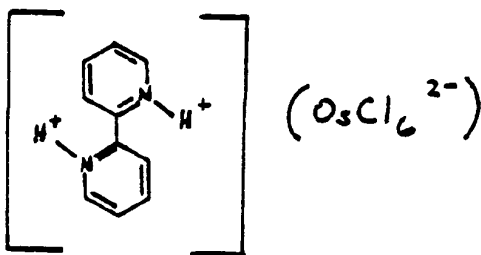
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Example 53 - Preparation of 2,2'-bipyridinium-hexachloroosmate (VI)

1.01 g of ammonium hexachloroosmate (VI), $(\text{NH}_4)_2\text{OsCl}_6$ (Alfa) (1.00 equivalents) were dissolved in 50 ml of 3N HCl (aq) at 70°C. To the hot, stirred solution was added dropwise a solution of 2,2'-bipyridine in 3N HCl (aq). A red salt,



(hereinafter $(\text{bpyH}_2^{2+})\text{OsCl}_6^{2-}$), with a molecular weight of 562.2g/mole, precipitated from the solution.

Precipitation was completed by cooling the solution at 0°C in an ice bath for 2 hrs. The product was collected by suction filtration on a glass fritted filter. The precipitate was washed successively with 5-10 ml portions of ice cold 3N HCl (aq) and water. After a final washing with 20 ml of anhydrous diethyl ether, the product was dried at 70°C under vacuum for 48 hrs. Yield = 1.01 g orange powder (78% based on $(\text{NH}_4)_2\text{OsCl}_6$). The product was used without further purification.

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Example 54 - Preparation of 2,2'-bipyridine-tetrachloroosmate (VI)

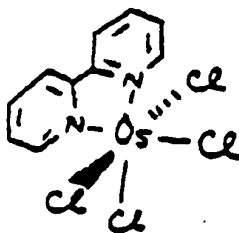
5 0.9g(1.60 mmol) of the $(bpyH_2^{2+})(OsCl_6^{2-})$ salt prepared in Example 53 was weighed into a pyrex test tube. The test tube was placed into a pyrex furnace tube fitted with an argon inlet and outlet and a thermometer (400°C max. reading).

10 The entire assembly was placed in a tube furnace and flushed with argon. The furnace was activated and brought to a temperature of 270-300°C. A slow stream of argon was passed through the tube throughout the reaction. The argon outlet was vented into the fume
15 hood to carry HCl produced during the reaction out of the laboratory. As the temperature approached 270°C, pyrolysis of the reactant began. HCl gas was observed to evolve from the solid. Vigorous evolution of HCl
20 was observed for 30 min and then slowly began to subside. After 6 hrs the oven was shut off and the sample cooled to room temperature. The product $(bpy)OsCl_4$, a finely divided red-brown solid, was purified by suspension in a stirred 3N HCl (aq) solution for 12 hrs followed by suspension for 6 hrs in
25 stirred anhydrous diethyl ether. Isolation of the product via suction filtration on a glass fritted filter gave 0.75 g (95%) yield of 2,2'-bipyridine-tetrachloroosmate (VI), hereinafter $(bpy)OsCl_4$. (MW=489.3 g/mole).

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The product was used without further purification.

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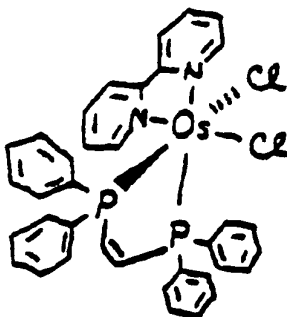
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Example 55 - Preparation of cis-(2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene]-dichloroosmium (II)

5 Into a 50 ml round bottom flask equipped with a stirbar and reflux condenser were added 230 mg (0.47 mmol) (bpy)OsCl₄ prepared as in Example 54 and 465 mg (1.17 mmol) cis-bis (1,2-diphenylphosphino)ethylene (DPP-ene). 25 ml diglyme were added and the contents were
10 refluxed under an argon atmosphere for 5 hrs. The dark green-black solution was cooled to room temperature under argon atmosphere and transferred to a 200 ml beaker. Addition of 80 ml anhydrous diethyl ether gave a dark green precipitate. The precipitate was
15 collected by suction filtration on a glass fritted filter (medium porosity) and washed with five 20 ml portions of anhydrous diethyl ether. The precipitate was then dissolved in 15 ml CH₂Cl₂ to give a forest green solution. The solution was allowed to gravity
20 drip through a glass fritted filter (medium porosity) into approximately 300 ml stirred anhydrous diethyl ether. Precipitation of the gray-green cis-(2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene-
25 dichloroosmium II complex, hereinafter cis-(bpy)(DPP-ene)OsCl₂), occurred. The complex was collected by suction filtration on a glass fritted filter (medium porosity). The product was washed quickly with 20 ml cold 1:2 v/v ethanol/water followed by seven 30 ml
30 portions of diethyl ether. The slate-gray powder was dried over CaSO₄ overnight in the vacuum desiccator.

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Yield: 240 mg (63% based on (bpy)OsCl₄; MW=814.4 g/mole. The product was used without further purification.

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Example 56 - Preparation of (2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene {2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane} osmium (II) dichloride

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Into a 100 ml round bottom flask equipped with stirbar and reflux condenser were added 129 mg (0.158 mmol) cis-(bpy)(DPPene)OsCl₂ prepared as in Example 55 and 0.3g (1.0 mmol) of 2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane, i.e., bpyoxal. To this was added 15 ml ethylene glycol. The suspension was refluxed under an argon atmosphere for 3 hrs. After cooling to room temperature, 10 ml H₂O were added to the contents of the flask.

15

A column (30 cm height x 19 mm i.d.) of SP-Sephadex C-25 ion exchange resin in water was prepared. The contents of the reaction flask were loaded onto the column. The column was eluted with H₂O (approximately 500 ml) to remove ethylene glycol and excess bpyoxal ligand. elution with 0.25 NaCl (aq) solution separated a small pale yellow band (orange fluorescence under long wave UV light) followed by a non-luminescent olive green band. These bands represented side-products of the reaction. No attempt was made to identify them and they were discarded. Once these bands were removed from the column, elution with 0.5M NaCl (aq) was begun. Three bands were eluted. The first, an orange band with orange luminescence, was identified as the chloride salt of the desired product moiety, (2,2'-bipyridine) [cis-bis] (1,2-diphenyl phosphino) ethylene] 2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3,-dioxolane osmium (II), hereinafter (bpy)(DPPene)(bpyoxal)Os(II)²⁺. It separated cleanly from a trailing

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yellow band (green luminescence) and brown band (non-luminescent).

The volume of solution in the fraction (approximately 400 ml) containing the (bpy)(DDPene) (bpyoxal)Os(II)²⁺ was reduced by evaporation to approximately 50 ml on a rotary evaporator. The aqueous NaCl concentrate was then extracted with three 50 ml portions of CH₂Cl₂. The CH₂Cl₂ extracts were combined, dried over anhydrous Na₂SO₄, and gravity filtered through a fluted filter paper. The red CH₂Cl₂ solution was concentrated to a volume of 10 ml by evaporation on the rotary evaporator. The complex was isolated as an orange solid by slowly dripping the CH₂Cl₂ solution into 200 ml of well-stirred petroleum ether. The precipitated complex was collected by suction filtration on a glass fritted filter (medium porosity). The product was washed three times with 15 ml portions of diethyl ether. The product was dried over CaSO₄ overnight in the vacuum desiccator.

Yield: 110 mg of (bpy)(DDPene)(bpyoxal)Os(II)²⁺(Cl⁻)₂ · 2H₂O (63% based on cis-(bpy)(DDPene)OsCl₂).

The product is a dihydrate and is analytically pure: C₅₇H₅₄N₄O₄P₂Cl₂Os · 2H₂O (MW = 1109.59 g/mole).

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Theory: C, 55.20%; H, 4.87%; N, 5.05%; O, 5.77%; Cl, 6.39%. Found: C, 56.03%; H, 5.18%; N, 4.88%; O, 6.87%; Cl, 7.07%.

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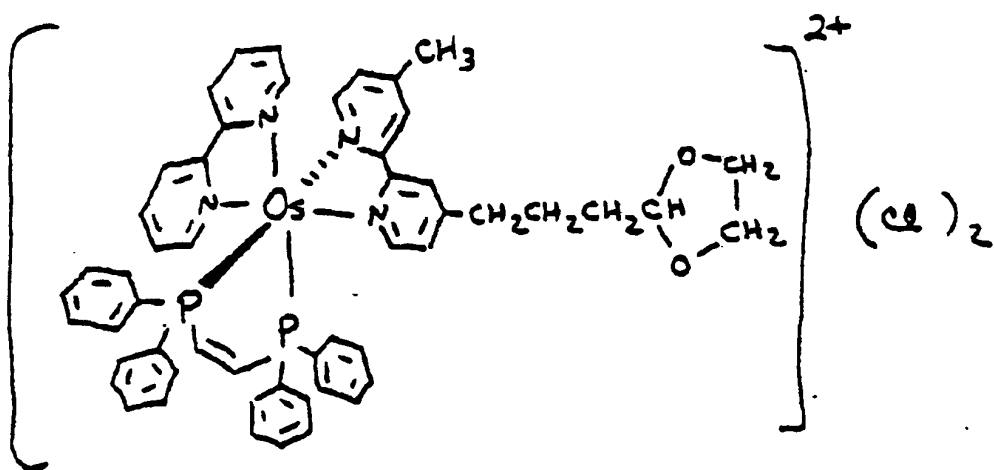
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Example 57 - Preparation of cis-dichloro-bis-[4,4'-carbomethoxy)-2,2'-bipyridin] ruthenium (II)

5 Into a 250 ml round bottom flash equipped with stirbar and reflux condenser were added 750 mg (1.55 mmol) of cis-tetra-(dimethylsulfoxide) dichlororuthenium (II) (Ru(DMSO)₄Cl₂) and 100 ml ethylene glycol. The contents of the flask were brought to a gentle boil under argon atmosphere and 756 mg (3.09 mmol) of (4,4'-carbomethoxy)-2,2'-bipyridine were added. Heating under argon was continued for 5 min. The orange solution became brown/black and 0.75 g of lithium chloride and 10 50 ml ethylene glycol were added. The solution was heated for another 10 min. After cooling to room temperature, approximately 100 ml H₂O were added to the mixture. The mixture was extracted with five 200 ml portions of CH₂Cl₂.

20 The CH₂Cl₂ extracts were washed with six 200 ml portions of water. The water layers were tested for fluorescence (red) after each washing and washing was continued if necessary until no fluorescence could be detected in the aqueous layer. The CH₂Cl₂ extracts were dried over anhydrous Na₂SO₄. The product was isolated by evaporation of the CH₂Cl₂ solution of the product into a stirred 10-fold volume excess of anhydrous diethyl ether. The precipitated product was collected by suction filtration, washed once with 30 ml diethyl ether and dried over CaSO₄ overnight in a vacuum desiccator. Yield = 25% dark metallic green crystals.

The product is analytically pure.

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Theory: C, 44.57; H, 4.01; N, 7.43; Cl, 9.40; O, 21.21.

Found: C, 44.16; H, 3.72; N, 7.11; Cl, 9.53; O, 20.15.

MW = 754.5 g/mole.

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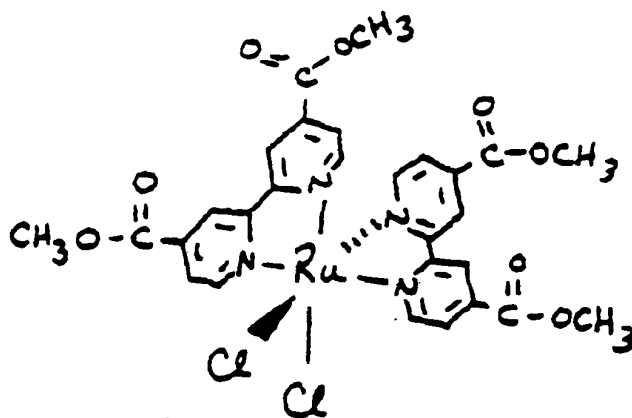
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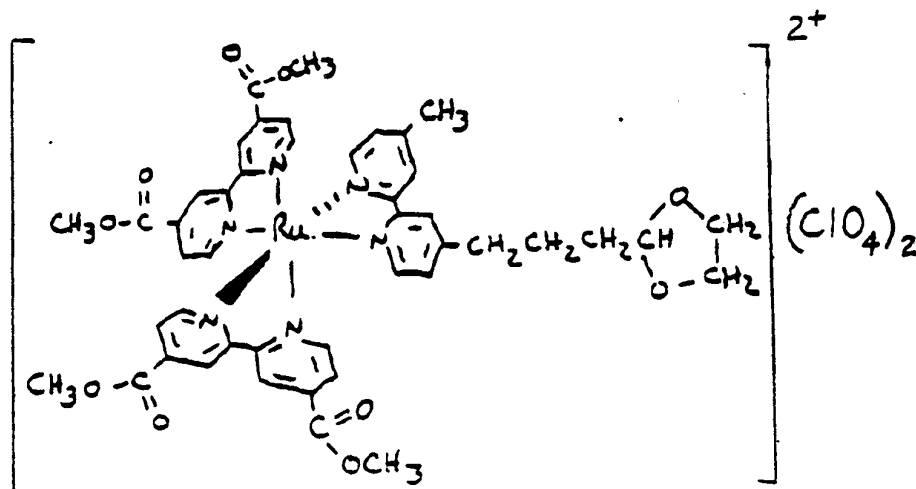
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Example 58 - Preparation of bis[(4,4'-carbomethoxy)-2,2'-bipyridine] 2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane ruthenium (II) diperchlorate

To 250 mg of bis(4,4'-dicarbomethoxy-2,2'-bipyridine) ruthenium (II) dichloride (0.33 mmol) in 50 ml methanol/water (1:1) were added 105 mg (0.37 mmol) of 2-[3-(4-methyl-2,2'-bipyridine-4'-yl)-propyl]-1,3-dioxolane (bpyoxal) and the mixture was refluxed for 12 hrs under an argon atmosphere. The solution was cooled and 0.5 ml of 70% HClO₄ were added. The methanol was slowly evaporated. Red crystals precipitated and were collected on a fritted funnel, washed with a small amount of cold water followed by ethanol and ether and dried to vacuo. Similar methods were used to prepare complexes from bis (4,4'-dicarbomethoxy-2,2'-bipyridine) ruthenium (II) dichloride and either 4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid or 4-4'-methyl-2,2'-bipyridine.



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Example 59 - Compound I Labeling of Human IgG

2 ml of human IgG (2.5 mg/ml) were dialyzed against 2
liters of 0.2M sodium bicarbonate buffer, pH 9.6,
5 overnight at 4°C with gentle stirring. Compound I was
prepared in a 100 molar excess to protein present (2.7
mg/100 microliters dimethylformamide) and allowed to
dissolve. The dialyzed protein was added dropwise to
the tag-aldehyde while gently stirring at room tempera-
10 ture for 2 hrs. A 100 molar excess (to protein) of
sodium borohydride (100 microliters of a 1.24 mg/ml
solution in deionized water) was added to the solution
and gently stirred for an additional 30 min at room
temperature. The conjugate was loaded onto a Sephadex
15 G-25 column (1.0 cm x 18.0 cm) equilibrated at room
temperature with 0.2M Tris, pH 8.0, and the eluant was
monitored at 280 nm. The void volume peak was
collected, pooled, and dialyzed against 2 liters of the
Tris buffer. The conjugate was tested for
20 immunological activity by standard ELISA methods, and
stored at 4°C until used.

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Example 60 - Biomag[®]/Goat-Anti-Human IgG Particle Preparation

5 A 2.5 ml aliquot of 5% Biomag[®]-amine terminated particles (Advanced Magnetics, Cambridge, MA) was transferred to a clean T-flask and washed 5 times with phosphate buffered saline (PBS). The wet cake was resuspended in 12.5 ml of 5% fresh glutaraldehyde (Sigma) and rotated end-over-end at ambient temperature
10 for 3 hrs. The wet cake was transferred to a second T-flask and washed 3 times with PBS. The activated particles were resuspended to 12.5 ml with PBS, and transferred to a 15 ml centrifuge tube. To this suspension were added 12.5 mg of goat anti-human IgG (H+L) (Jackson Labs) in 2 ml of PBS. The tube was rotated end-over-end for 3 hrs at room temperature and then overnight at 4°C. The next day the particles were washed 2 times with PBS containing 1% (w/v) bovine serum albumin (BSA) followed by storage in 12.5 ml of
15 PBS containing 0.1% BSA at 4°C until use.
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Example 61 - Pseudo-Homogeneous Human IgG
Electrochemiluminescence Assay Using Biomag[®] Magnetic
Based Particle Assay

Compound I-human IgG conjugate was diluted 1:50 in
0.15M phosphate buffer containing 0.1% BSA and
5 aliquoted at 250 microliters/tube. 150 microliters of
diluent (PB w/BSA as above) were added per tube,
followed by the addition of either various dilutions of
analyte (human IgG) or diluent (negative controls).
10 Additionally, a non-specific analyte (goat anti-rabbit
IgG) was used in some tubes to check for assay
specificity. 50 microliters of 1% Biomag[®] coupled with
goat anti-human IgG (H&L) were added to each tube. The
tubes were mixed on a vortex and incubated at room
temperature for 15 min with gentle shaking. The
15 particles were magnetically removed from solution and
100 microliters of the resulting supernatant were
transferred to a Berthold tube to which 400 microliters
of a citrate-oxalate-Triton X-100 solution were added.
The tube was mixed on a vortex and read in a modified
20 Berthold luminometer fitted with an R-268
photomultiplier tube and a 0.29 inch diameter circular
52 gauge double platinum mesh electrode supported by a
conductive paint covered polycarbonate support and
connected to the voltage source through a 0.01 inch
25 diameter platinum wire/silver paint contact. The
potential applied was stepped from +1.4 to +2.15 V
while a 10 second integration was done. Between
readings the electrode was electrochemically cleaned by
rinsing with deionized water, then immersing in 0.1M
30 phosphate-citrate buffer containing oxalic acid and
Triton X-100 and stepping the applied potential
from -2.2V to +2.2V (with 3 seconds at each potential)

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for 2 min and 20 s conds followed by pausing the potential at +2.2V for 10 seconds. The electrode was removed from the cleaning solution, rinsed with deionized water and blotted dry with an absorbent wiper. In this format the electrochemiluminescent signal was directly proportional to the analyte concentration.

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Example 62 - Preparation of bis(2,2'-bipyridine)maleimidohexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II) diperchlorate: Compound IV

5 4-[4-(1-aminobutyl)]-4'-methyl-2,2'-bipyridine, prepared as described in Example 32 from 500 mg (1.35×10^{-3} mol) of phthalimide, was dissolved in 5 ml of dry pyridine with 0.313 g (1.48×10^{-3} mol) of maleimide
10 hexanoic acid and 0.306 g (1.48×10^{-3} mol) of DCC. After storing overnight, the dicyclohexylurea was filtered off and the pyridine was stripped under vacuum. The residue was purified by column chromatography (activity II alumina, 5% MeOH/CH₂Cl₂) to
15 give the purified product. Yield: 0.56 g (95%).

100 mg (2.3×10^{-4} mol) of maleimido-hexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide and 100 mg (2.06×10^{-4} mol) of bis-(2,2'-bipyridine) ruthenium dichloride
20 dihydrate were dissolved in 50 ml of ethanol/water (1:1), degassed with argon and refluxed for 4 hr. The resulting clear orange solution was diluted with 25 ml of solid NaClO₄, 25 ml of ethanol and 25 ml of acetone, and slowly evaporated under vacuum. When dry, another
25 25 ml of water and 25 ml of acetone were added and the solution was rotoevaporated to about 15-20 ml. The precipitated solid was collected, washed with water and dried. The sample was purified by preparative TLC on
30 alumina using 1:9 methanol/chloroform. The fast running band was isolated by scraping off the plate and eluting the compound by stirring in methanol/chloroform (1:1). After filtration to remove the alumina, the orange solution was evaporated to dryness to give 86.7 mg of purified compound (72%). The
35 structure was confirmed by NMR.

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Example 63 - Labeling of hCG Peptide with Compound IV

2 mg of human chorionic gonadotropin (hCG) peptide (#109-145, JPl41, Vernon Stevens, Ohio State University) were suspended in 1 ml of 0.15M citrate buffer, pH 6.0, and 1.13 mg of Compound IV were dissolved in 300 microliters of dimethylformamide. The peptide solution was added dropwise to the Compound IV solution over a one minute period. The solution was stirred gently at room temperature for 1 hr. The sample was then loaded onto a Bio-Gel P-2 column (Bio-Rad; 1 cm X 45 cm) which was equilibrated at room temperature with 0.2M Tris-base, pH 8.5 at a flow rate of 15 ml/hr and the eluant was monitored at 280 nm. The void volume of the run was collected, pooled, and loaded onto a QAE-Sephadex A-25 column (Pharmacia; 1 cm x 10 cm) which was equilibrated at room temperature with 50 mM Tris-HCl, pH 7.0. This was done to remove any unlabeled peptides (which will adsorb to the positively charged resin; the positive charge of the labeled peptide allowed it to pass out of the column without further treatment). The eluant was monitored at 280 nm, and the first major peak was collected, pooled, and concentrated by lyophilization. The dried compound was resuspended in a minimal volume of PBF. The hCG peptide-Compound IV conjugate was stored at 4°C until used.

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Example 64 - Purification of Rabbit Anti-hCG Peptide By
DEAE AFFI-G 1 Blue Chromatography

4 ml of DEAE AFFI-Gel Blue (Bio-Rad) was poured into a
5 chromatography column (1 cm x 10 column size) and
equilibrated at room temperature with 0.2M Tris-HCl
containing 0.028M NaCl, pH 8.0, for 1 hr at a 40 ml/hr
flow rate. The flow rate was slowed to 20 ml/hr and 1
10 ml of rabbit anti-hCG peptide (anti 109-145, Vernon
Stevens, Ohio State University), which was predialyzed
against the column buffer, was loaded onto the column.
1 ml fractions were collected and the eluant was
monitored at 280 nm. The void volume peak was
collected, pooled from several runs, and concentrated
15 by placing the eluant in a 12K MWCO dialysis sack which
was surrounded with polyethylene glycol 6,000 (Sigma).
The antibody was tested for immunoreactivity using
standard ELISA techniques and for purity by HPLC, which
resulted in one major peak, indicating a pure, active
20 preparation.

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Example 65 - Titration of hCG Peptide-Compound IV
Conjugate Against Purified Rabbit Anti-hCG Peptide:
Electrochemiluminescence Signal Modulation

5 hCG peptide-Compound IV Conjugate was diluted in 0.1M
phosphate buffer containing 0.1M citrate, pH 6.2.
Aliquots of this mixture were placed in microfuge
tubes. The previously purified anti-peptide antibody
10 was diluted to various concentrations and added to the
tubes, which were mixed on a vortex and incubated for 1
hr at room temperature. Just before reading
electrochemiluminescence, an aliquot was transferred to
a Berthold tube to which oxalic acid and Triton X-100
(Sigma) were added. The tubes were mixed on a vortex
15 and read using sweep mode (+1.5 to +2.5V at 50mV/sec of
which 3 scans were done, taking the peak height of the
second scan as the value) on a Berthold luminometer
using an R-268 photomultiplier tube with a double
platinum mesh electrode. Between readings the
20 electrode was cleaned electrochemically by first
rinsing it with deionized water and then immersing it
in 0.1M phosphate-citrate buffer, pH 6.1, containing
25mM oxalate and 1% Triton X-100. The applied potential
was switched, with three seconds at each potential,
25 between +2.2V and -2.2V for 1 min and 50 sec and poised
at +2.2V for 10 sec. The power was then shut off and
the electrode removed from the cleaning solution and
rinsed with deionized water and blotted dry with
absorbent toweling.

30 The results showed a general trend in which the
electrochemiluminescent signal was directly proportion-
al to the concentration of antibody to the peptide.
This is in contrast to other experiments in which the

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electrochemiluminescent signal drops as the
electrochemiluminescent labelled analyte is contacted
with antibody.

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Example 66 - Electrochemiluminescence Signal Output:
Stability Data Of Ru(II)-Compound III Conjugate

Ru(II)-Compound III conjugate was diluted to 300 nM in 0.1M phosphate-citrate buffer, pH 6.1, either with or without 1% normal human serum (NHS). One of each buffer type was incubated at 4°, 20°, 37° and 55°C and sampled on incubation days 3, 4, and 5. The samples were equilibrated at room temperature and evaluated for their electrochemiluminescence signal output. 400 microliters of sample was mixed with 100 microliters of 125mM oxalic acid-5% Triton X-100 in a tube and placed in a modified Berthold luminometer with a Hamamatsu R-268 photomultiplier tube and a double mesh platinum gauze electrode. The reading was made by sweeping the applied potential from +1.5V to +2.5V at 50 mV/second for 3 cycles and the height of the second cycle peak was recorded and converted into electrochemiluminescent counts. Between readings the electrode was cleaned electrochemically by rinsing with deionized water, immersing the electrode in 0.1M phosphate-citrate buffer containing 25 mM oxalate and 1% Triton X-100 and pulsing the potential from -2.2V to +2.2V with a 3 sec pause at each potential for 1 min and 50 sec. The potential was poised at +2.2V for 10 sec and then removed. The electrode was removed from the cleaning solution, rinsed with deionized water and blotted dry with absorbent toweling. The results show that there was consistent signal in each buffer type over the course of the study. This indicates the stability of the reagent and its capability of generating an electrochemiluminescent signal.

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Example 67 - Immunoreactivity Testing of Ru(II)-
Compound III Conjugate: Stability Studies

5 The Ru(II)-Compound III conjugate was diluted and
incubated under the conditions as described in Example
66. Samples were taken on days 3, 4, and 5 cooled to
room temperature and tested for immunoreactivity by
Particle Concentration Fluorescent Immunoassay (PCFIA)
10 using a Pandex Screen Machine obtained from Pandex,
Inc., Mundelein, IL. The PCFIA was run in a
competitive assay format. The latex particles (Pandex)
were conjugated with theophylline-BSA. A constant
amount of these particles were mixed with the Ru(II)-
Compound III Conjugate test solution to final
15 concentrations of anti-theophylline monoclonal antibody
(ascites, Hyclone cat #E-3120M, lot #RD200) and a
constant amount of goat anti-mouse IgG-FITC conjugate
(Pandex, cat #33-020-1 lot # C01). After incubation,
the samples were processed and read on the Pandex
20 Screen Machine. The results showed that there was no
appreciable loss of activity even after 5 days
incubation at 55°C.

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Example 68 - Electrochemiluminescence of various ruthenium and osmium compounds

The electrochemistry of various osmium and ruthenium compounds was measured as 1 mM solutions in 10 ml of nitrogen-purged acetonitrile with 0.1 M tetrabutylammonium tetrafluoroborate as an electrolyte. The working electrode was a platinum disk electrode obtained from Bioanalytical Systems, Inc., West Lafayette, IN. A platinum wire counter electrode and a 1.0 mm silver wire was used as a reference electrode. Measurements were made by scanning from -2.2V to +2.2V (vs SCE) at a scan rate of 100 mV/second. After each electrochemical measurement the potential difference between a Saturated Calomel Reference Electrode (SCE) and the silver wire was determined. Thus, the values reported are corrected to the potential versus SCE.

Electrochemiluminescent (ECL) measurements were made in 0.5 ml aqueous solutions containing 0.1M phosphate-citrate buffer (pH 4.2), 25 mM oxalic acid, and 1% Triton X-100. The electrode system used consisted of two platinum gauze (52 gauge) electrodes connected to a Radio Shack transistor socket (#276-548) by a 0.1 mm platinum wire. The electrodes were mounted on the outside of a 60 ml thick piece of cellulose acetate plastic. This plastic was machined so that a 1/4 inch diameter hole allows solution to easily flow between the working and counter-reference electrodes. The electrodes were connected to the potentiostat so that one electrode functioned as a working electrode (which was closer to the photomultiplier tube) and one electrode functioned as the counter and reference electrode. Measurements were made by sweeping from

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1.5V to 2.5V (bias potential) at a scan rate of 50 mV/second. The ECL measurements are reported as the signal to noise ratio, i.e., or signal to background ratio for a given concentration of compound. Background is defined as the luminescent counts observed with buffer and no ECL compounds added. Luminescent measurements were the peak light output observed during the first or second linear sweep.

Both electrochemiluminescent (ECL) and cyclic voltammetric measurements of each solution were performed with either a EG&G Model 273 potentiostat or a bipotentiostat from Ursar Scientific Instruments, Oxford, England. The photon flux of each ECL measurement was monitored with a Berthold Biolumat LB 9500 luminometer from Wilebad, West Germany, modified so that either a two or three electrode system could be placed in the 0.5 ml measuring solution. Both electrochemical and electrochemiluminescent measurements were recorded on a Kipp & Zonen Model BD 91 X, Y, Y' recorder from Delft, Holland.

Fluorescence measurements were made with 50 micromolar solutions of the desired compound in 3.0 ml of ECL solution, or when insoluble in ECL solution, in acetonitrile. Measurements were made on a Perkin-Elmer LS-5 Fluorescence Spectrophotometer. Prescans of the solutions' excitation and emission spectra were performed before the excitation and emission spectra were recorded so that the emission spectrum could be measured while irradiating at the maximum excitation wavelength and conversely, the excitation spectrum could be recorded while monitoring the maximum emission wavelength.

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	Compound	E_{ox}	E_{red}	Fluorescence Emission Max	ECL(S/N) ¹ & Conc.
		vs. SCE			
	a) Ru(bipy) ₃	1.07 V	-1.52 V	625 nm	1 x 10 ⁻⁹ M (2.5)
5	b) Ru(4,4'- -CO ₂ -bipy) ₃	1.19 V	N.D.	628 nm	2 x 10 ⁻⁹ M (4.05)
	c) Ru(4,4'-CO ₂ - Et-bipy) ₃	1.54 V	-0.89 V	636 nm	1 x 10 ⁻¹⁰ M (2.01)
10	d) Ru(bipy) ₂ (C-8-theo- phylline C-4-bipy)	1.17 V	N.D.	624 nm	1 x 10 ⁻⁹ M (.97)
	e) Os(bipy) ₂ (CO) (py)	1.82 V	-0.99 V	585 nm	1 x 10 ⁻⁸ M
15	f) Os(DPPene) (bpy) (bpyoxal)	1.60 V	N.D.	630 nm	6 x 10 ⁻⁸ M (10.8)

¹(S/N) is the signal to noise ratio, where the signal is defined as the ECL output (luminescent counts) of a compound at a given concentration, and noise is the luminescent counts of the buffer in which the compound was dissolved.

Unlike the ECL of the other compounds which were measured at pH 4.2, compound C) also displays significant ECL at the physiological pH of 7.0.

- a) Tris (2,2'-bipyridine) ruthenium ²⁺
- b) Tris (4,4'-carboxylate-2,2'-bipyridine) ruthenium ²⁺
- c) Tris (4,4'-carboethoxy-2,2'-bipyridine) ruthenium ²⁺
- d) bis(2,2'-bipyridine)[theophylline-8-butyric-4-(4-methyl-2,2'-bipyridine)-4'-yl)-butyl]amide]ruthenium (II) dichloride

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-- e) [bis-(2,2'-bipyridine) {monocarbonyl} pyridyl] osmium (II)
dihexafluorophosphate

5 f) (2,2'-bipyridine[cis-bis(1,2-diphenylphosphine)ethylene]{2-
[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane}
osmium (II) dichloride

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Example 69 - Preparation of bis(2,2'-bipyridine)[4-(4'-methyl-2,2'-bipyridine)butyl amine] ruthenium (II) diperchlorate

5 1.29 g (2.48 mmol) of bis-(2,2'-bipyridine) ruthenium (II) dichloride dihydrate (Strem) and 0.719 g (2.98 mmol) of 4-[4-(1-aminobutyl)]4'-methyl-2,2'-bipyridine (described in Example 32) were suspended in 50 ml of 50/50 ethanol/H₂O and refluxed under argon for 3 hr.

10 The reaction solution was concentrated and subjected to chromatographic separation on Sephadex C-25, eluting first with distilled water and then with 0.25 M NaCl. Final elution was performed with 0.4 M NaCl. The fractions containing the product were concentrated to

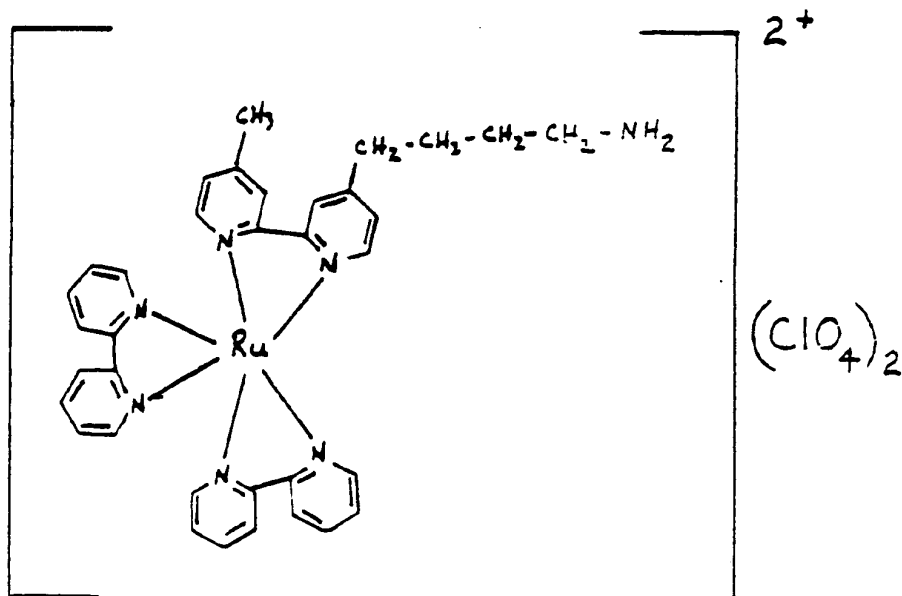
15 about 100 ml and perchloric acid was added until the solution became cloudy. Upon refrigeration overnight, red orange crystals of the product (1.215 g) were obtained. Elemental analysis confirmed the structure to be

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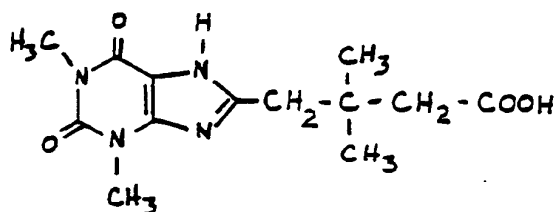


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Example 70 - Preparation of theophylline-8-(3,3-dimethyl) butyric acid

5 5,6-diamino, N¹, N³-dimethyl uracil hydrate (10 g, 0.059 mol) and 3,3-dimethylglutaric acid anhydride (12.6 g, 8.85 x 10⁻² mol) were dissolved in 100 ml N,N-dimethyl aniline and brought to reflux using a Dean-Stark trap. The contents of the reaction became solubilized after heating and turned orange red. After 4 hrs of reflux, 1
10 ml of water was collected in the trap, indicating the reaction had completed. The reaction was allowed to cool to room temperature, at which time the entire mass solidified. The solid was filtered on a fritted funnel and washed until it became pale yellow in color. After
15 two crystallizations from DMF, pure white crystals of the intermediate lactam (melting point 283.1-284.9°C) were obtained. The lactam was boiled in water for 8 hrs. Upon cooling, the solution yielded pure white crystals (melting point 218-220°C). Proton NMR and
20 elemental analysis were consistent with the following structure

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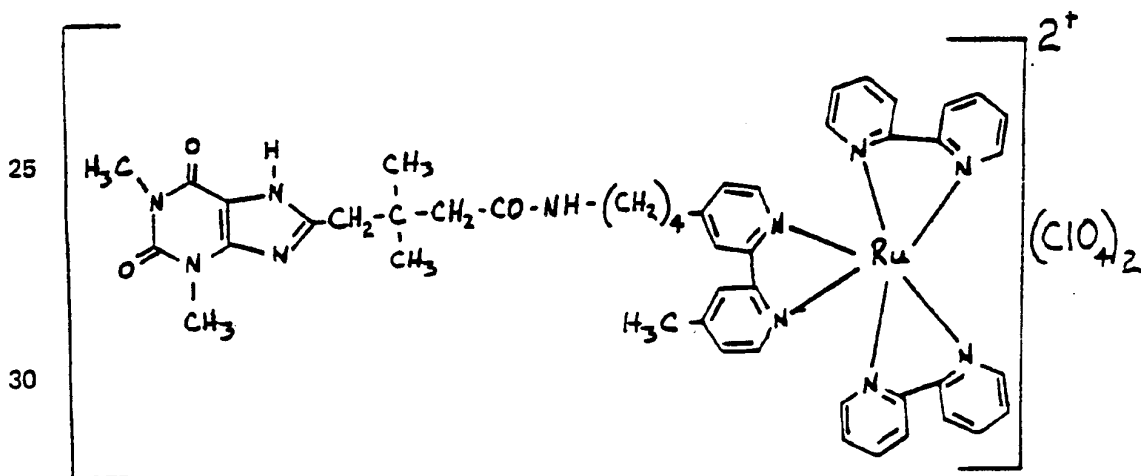
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Example 71 - Preparation of bis-(2,2'-bipyridine)[theophylline-8-(3,3-dimethylbutyric-{4-(4-methyl-2,2'-bipyridine-4'-yl)-butyl}amide]ruthenium (II) diperchlorate

100 mg (0.34 mmol) of theophylline-8-(3,3-dimethylbutyric acid and 0.325 g (0.34 mmol) of bis (2,2'-bipyridine) [4-(4'-methyl-2,2'-bipyridine) butyl amine ruthenium (II) diperchlorate were dissolved in 10 ml of dry pyridine along with 0.351 g (1.7 mmol) of dicyclohexylcarbodiimide. The reaction was allowed to stir for 2 days. Copious dicyclohexylurea precipitate was removed by filtration and the pyridine solution was concentrated under vacuum. The residue was dissolved in methanol and chromatographed on Sephadex LH-20. On concentration of the desired fractions the product was precipitated from ether to give an orange precipitate. Elemental analysis and proton NMR were consistent with the structure

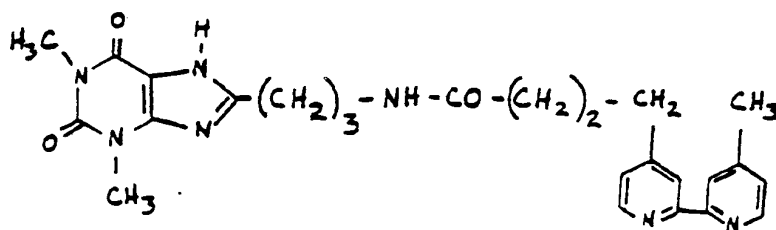
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Example 72 - Preparation of theophylline-8-propyl(4-methyl-2,2'-bipyridine-4'-butyl) carboxamide

61.8 mg (1.55×10^{-4} mol) of 8-(gamma-aminopropyl) theophylline phthalic acid hydrazide salt were dissolved in 1 ml H_2O and acidified with HCl. The precipitated phthalic acid hydrazide was filtered off and the aqueous filtrate was evaporated and dried under vacuum to give 34.5 mg (1.31×10^{-4} mol) of theophylline-8-propylamine hydrochloride. This material was dissolved in 5 ml dry pyridine and 36.9 mg (1.44×10^{-4} mol) of 4-(4-methyl-2,2'-bipyridine-4'-yl) butyric acid and 26.4 mg (2.62×10^{-4} mol) of triethylamine were added to it. Finally, 29.7 mg (1.44×10^{-4} mol) of dicyclohexylcarbodiimide were added to the solution. The resulting cloudy solution was allowed to stir overnight. The precipitated salt and dicyclohexylurea were filtered off and the solution was stripped to dryness to give a white solid which was chromatographed on alumina using 5% methanol in dichloromethane as the eluant to yield 44.2 mg (71%) of the product as a white powder (melting point 215° - $216.5^{\circ}C$). Proton NMR and elemental analysis were consistent with the following structure

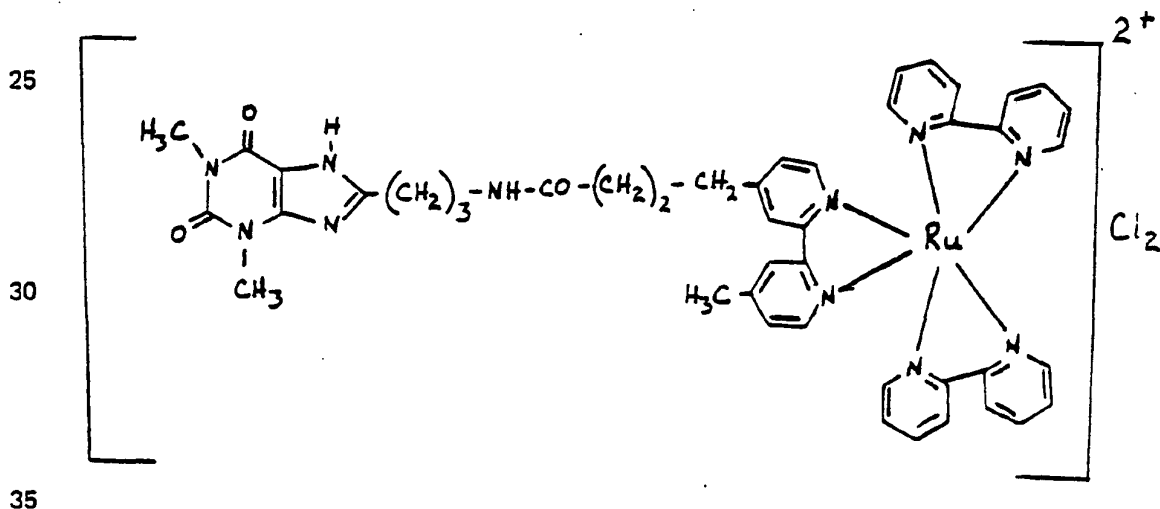


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Example 73 - Preparation of bis-(2,2'-bipyridine)[theophylline-8-propyl(4-methyl-2,2'-bipyridine-4'-butyl)carboxamide]ruthenium (II) dichloride

5 100 mg (2.06×10^{-4} mol) of bis(2,2'-bipyridine) ruthenium dichloride (Strem) and 108 mg (2.27×10^{-4} mol) of theophylline-8-propyl(4-methyl-2,2'-bipyridine-4'-butyl) carboxamide were added to 50 ml of 50% ethanol which had been degassed with argon. The solution was heated at reflux. The resulting cherry red solution was concentrated under high vacuum at room temperature. The residue was chromatographed on Sephadex LH-20 (75 cm x 19 mm I.D.) using methanol as eluent. The product band was isolated, evaporated and precipitated into anhydrous ether. The yield of the product was 156 mg (75%). Elemental analysis indicated the presence of 4 mols of methanol in the product.

20 Analysis: Calculated; C, 54.09; H, 5.65; N, 14.16; O, 10.29; Cl, 6.52; Found: C, 53.30; H, 5.22; N, 14.56; O, 10.63 and Cl, 6.95.



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Example 74 - Preparation of bis(2,2'-bipyridine)[1-bromo-4-(4'-methyl-2,2'-bipyridine-4-yl)butane]ruthenium (II) diperchlorate

5 0.29 g of the ligand 1-bromo-4-(4'-methyl-2,2'-
bipyridine-4-yl) butane prepared in Example 31 and 0.32
g of bis (2,2'-bipyridine) ruthenium (II) dichloride
were transferred to a flask in 60 ml of 1:1 v/v
ethanol/water mixture. The solution was degassed with
10 nitrogen for 15 min. The reaction was heated at reflux
under nitrogen for 3 1/2 hrs. An aqueous saturated
solution of NaClO₄ was added to the cooled red solution.
Solvents were removed under vacuum and the deep red
15 residue was chromatographed on an alumina column (20 cm
x 2.5 cm, Merck, neutral activity 3) using
acetonitrile/toluene 1:1 v/v as the eluant. The product
eluted as a dark red band (band #2) on the column. The
desired fraction was concentrated, redissolved in
approximately 10 ml volume of dichloro-
20 methane, precipitated from anhydrous ethyl ether and
finally dried under vacuum to yield 320 mg (53%) of the

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product. The structure of the product was confirmed by elemental analysis and spectral characteristics and is shown below.

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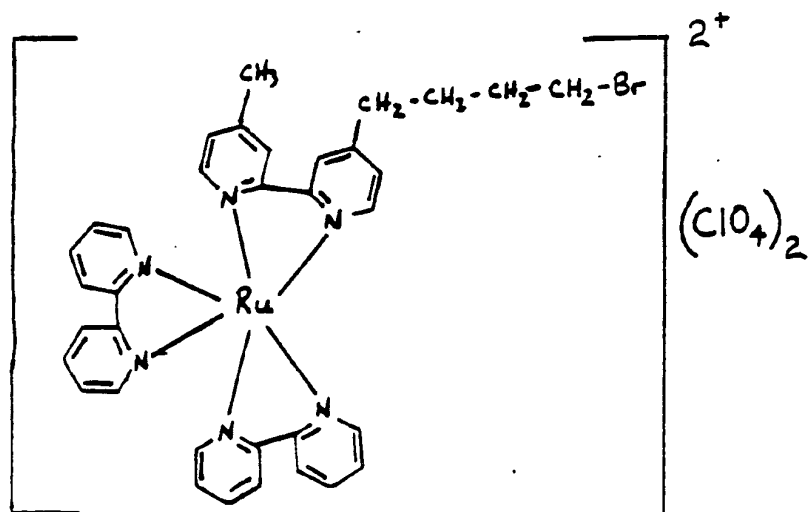
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Example 75 - Preparation of bis(2,2'-bipyridine)[theophylline-9-[4-(4-methyl-2,2'-bipyridine-4'-yl)butane] ruthenium (II) diperchlorate

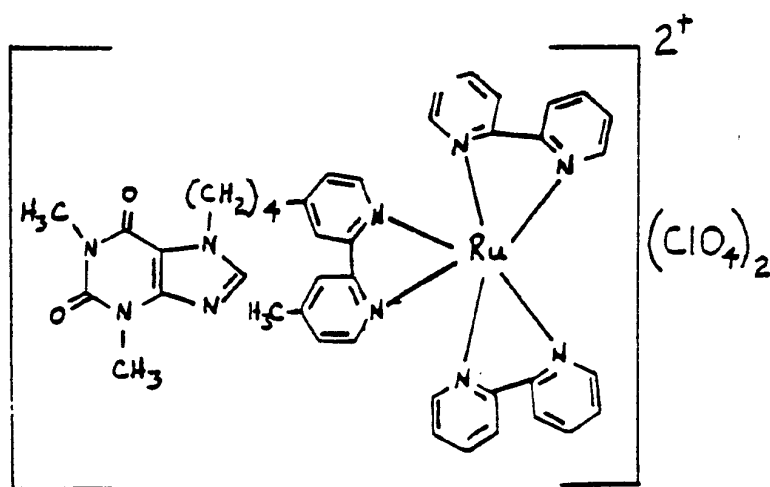
5 The silver salt of theophylline was prepared by mixing together an ammonia solution of theophylline (2.168 g in 25 ml conc. NH_4OH and 75 ml water) and an ammonia solution of silver nitrate (2.006 g in 10 ml conc. NH_4OH and 30 ml H_2O), the reaction being carried out in the dark. Shortly after mixing the two solutions, a white product appeared. Once the precipitation of the product was completed, the precipitate was collected by filtration on a 60 ml medium glass frit. The precipitate was washed with dilute ammonia and dried under vacuum. The structure of the silver salt of theophylline was supported by elemental analysis.

41 mg of the silver salt of theophylline and 122 mg of bis(2,2'-bipyridine)[1-bromo-4-(4'-methyl-2,2'-bipyridine-4-yl)butane] ruthenium (II) diperchlorate were dissolved in 15 ml of anhydrous dimethylformamide (DMF), the reaction being carried out in the dark. The resulting red solution was degassed with argon and heated at reflux for 24 hrs and then cooled to room temperature under argon. Cooling of the solution was continued down to ice temperature and then the black silver metal suspension was filtered off and washed with 5 ml acetone. The red filtrate was treated with 0.2g of lithium perchlorate and after dissolution of LiClO_4 , the solution was evaporated under vacuum and the red residue was dried overnight in the dark under vacuum. The sample was redissolved in 3-5 ml of methanol and 0.25 g of anhydrous LiClO_4 was added and dissolved. The resulting solution was chromatographed on a Sephadex LH-

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20 Column (75 cm x 19 cm i.d.) using methanol as the eluant. The major product eluted as fraction #2 (dark red band). Fraction #2 was concentrated, redissolved in methanol and precipitated into ethyl ether. The product was dried under vacuum to yield 80 mg (50% yield) of material, the structure of which was confirmed by elemental analysis, infra red and emission spectra.



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What is claimed is:

1. A method of detecting in a predetermined volume of a multicomponent, liquid sample an analyte of interest present in the sample at a concentration below about 10^{-3} molar which comprises:
 - a. contacting the sample with a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation and (ii) capable of combining with the analyte of interest, the contact being effected under appropriate conditions such that the analyte and reagent combine;
 - b. exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and
 - c. detecting electromagnetic radiation so emitted and thereby detecting the presence of the analyte of interest in the sample.
2. A method according to claim 1, wherein prior to step (b) any reagent which is not combined with the analyte of interest is separated from the sample resulting from step (a).
3. A method according to claim 1, wherein prior to contacting the sample with the reagent in step (a), the

sample is treated so as to immobilize the analyte of interest.

4. A method according to claim 1, wherein the analyte of interest is a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopoly-saccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule.

5. A method according to claim 4, wherein the pharmacological agent is theophylline.

6. A method according to claim 4, wherein the pharmacological agent is digoxin.

7. A method according to claim 4, wherein the hormone is human chorionic gonadotropin.

8. A method according to claim 4, wherein the analyte of interest is a whole cell, subcellular particle, virus, prion, viroid, nucleic acid, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, hormone, pharmacological agent, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule, and is present in the sample at a concentration below about 10^{-12} molar.

9. A method according to claim 8, wherein the analyte of interest is a whole cell, subcellular particle, virus, prion, viroid or nucleic acid, and is

present in the sample at a concentration below about 10^{-15} molar.

10. A method according to claim 1, wherein the reagent comprises an electrochemiluminescent chemical moiety conjugated to a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopoly-saccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, nonbiological polymer, synthetic organic molecule, organometallic molecule, inorganic molecule, biotin, avidin or streptavidin.

11. A method according to claim 1, wherein the reagent comprises an electrochemiluminescent chemical moiety conjugated to an antibody, antigen, nucleic acid, hapten, ligand or enzyme, or biotin, avidin or streptavidin.

12. A method according to claim 10 or 11, wherein the electrochemiluminescent chemical moiety comprises a metal-containing organic compound wherein the metal is selected from the group consisting of ruthenium, osmium, rhenium, iridium, rhodium, platinum, palladium, molybdenum and technetium.

13. A method according to claim 12, wherein the metal is ruthenium or osmium.

14. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is rubrene or 9, 10-diphenylanthracene.

15. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is bis[(4,4'-carbometh xy)-2,2'-bipyridine]2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-dioxolane ruthenium (II).
- 5 16. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is bis (2,2' bipyridine) [4-(butan-1-al)-4'-methyl-2,2'-bipyridine] ruthenium (II).
- 10 17. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is bis (2,2'-bipyridine) [4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II).
- 15 18. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is (2,2'-bipyridine) [cis-bis(1,2-diphenylphosphino)ethylene] {2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane} osmium (II).
- 20 19. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) [4-(4'-methyl-2,2'-bipyridine)butylamine] ruthenium II.
- 25 20. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) [1-bromo-4(4'-methyl-2,2'-bipyridine-4-yl)butane] ruthenium (II).
- 30 21. A method according to claim 12, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) maleimido hexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II) diperchlorate
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22. A method according to claim 1, wherein the sample is derived from a solid, emulsion, suspension, liquid or gas.

5 23. A method according to claim 1, wherein the sample is derived from water, food, blood, serum, urine, feces, tissue, saliva, oils, organic solvents or air.

10 24. A method according to claim 1, wherein the sample comprises acetonitrile, dimethylsulfoxide, dimethylformamide, n-methylpyrrolidinone or tert-butyl alcohol.

15 25. A method according to claim 24, wherein the sample additionally comprises a reducing agent.

26. A method according to claim 24, wherein the sample additionally comprises an oxidizing agent.

20 27. A method for detecting in a predetermined volume of a multicomponent, liquid sample an analyte of interest present in the sample at a concentration below about 10^{-3} molar which comprises:

25 a. contacting the sample with a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source
30 effective to induce the reagent to repeatedly emit radiation and (ii) capable of competing with the analyte of interest for binding sites on a complementary material not normally present in the sample, and with the complementary material, the

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contact being effected under appropriate conditions such that the analyte of interest and the reagent competitively bind to the complementary material;

5 b. exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and

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c. detecting electromagnetic radiation so emitted and thereby detecting the presence of the analyte of interest in the sample.

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28. A method according to claim 27, wherein the analyte of interest is theophylline, digoxin or human chorionic gonadotropin.

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29. A method according to claim 27, wherein the reagent is the analyte of interest conjugated to an electrochemiluminescent chemical moiety.

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30. A method according to claim 27, wherein the reagent is an analogue of the analyte of interest conjugated to an electrochemiluminescent chemical moiety.

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31. A method according to claim 29 or 30, wherein electrochemiluminescent chemical moiety is bis[(4,4'-carbomethoxy)-2,2'-bipyridine] 2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-dioxolane ruthenium (II).

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32. A method according to claim 29 or 30, wherein electrochemiluminescent chemical moiety is bis[(2,2'

bipyridine) [4-(butan-1-yl)-4'-methyl-2,2'-bipyridine] ruthenium (II).

5 33. A method according to claim 29 or 30, wherein electrochemiluminescent chemical moiety is bis (2,2'-bipyridine) [4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II).

10 34. A method according to claim 29 or 30, wherein the electrochemiluminescent chemical moiety is (2,2'-bipyridine) [cis-bis(1,2-diphenylphosphino)ethylene] {2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane} osmium (II).

15 35. A method according to claim 29 or 30, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) [4-(4'-methyl-2,2'-bipyridine)butylamine] ruthenium II.

20 36. A method according to claim 29 or 30, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) [1-bromo-4(4'-methyl-2,2'-bipyridine-4'-yl)butane] ruthenium (II).

25 37. A method according to claim 29 or 30, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine) maleimido hexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II).

30 38. A method according to claim 27, wherein the complementary material is a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent,

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tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule.

5 39. A method for quantitatively determining in a predetermined volume of a multicomponent, liquid sample the amount of an analyte of interest present in the sample which comprises:

10 a. contacting the sample with a known amount of a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly
15 emit radiation and (ii) capable of combining with the analyte of interest, the contact being effected under appropriate conditions such that the analyte and reagent combine;

20 b. exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to
25 repeatedly emit electromagnetic radiation; and

c. quantitatively determining the amount of radiation so emitted and thereby quantitatively determining the amount of the analyte of interest
30 present in the sample.

40. A method according to claim 39, wherein prior to step (b) any reagent which is not combined with the analyte of interest is separated from the sample
35 resulting from step (a).

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41. A method according to claim 39, wherein prior to contacting the sample with the reagent in step (a), the sample is treated so as to immobilize the analyte of interest.

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42. A method according to claim 39, wherein the analyte of interest is a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule.

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43. A method according to claim 42, wherein the pharmacological agent is theophylline.

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44. A method according to claim 42, wherein the pharmacological agent is digoxin.

45. A method according to claim 42, wherein the hormone is human chorionic gonadotropin.

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46. A method according to claim 39, wherein the reagent comprises an electrochemiluminescent chemical moiety conjugated to a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic

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organic molecule, organometallic molecule or inorganic molecule.

47. A method of claim 46, wherein the reagent comprises an electrochemiluminescent chemical moiety conjugated to an antibody, antigen, nucleic acid, haptan, ligand or enzyme, or biotin, avidin or streptavidin.

48. A method of claim 46, wherein the electrochemiluminescent chemical moiety comprises a metal-containing organic compound wherein the metal is selected from the group consisting of ruthenium, osmium, rhenium, iridium, rhodium, platinum, palladium, molybdenum and technetium.

49. A method of claim 48, wherein the metal is ruthenium or osmium.

50. A method of claim 46, wherein the electrochemiluminescent chemical moiety is rubrene or 9, 10-diphenylanthracene.

51. A method according to claim 46, wherein the electrochemiluminescent chemical moiety is bis[(4,4'-carbomethoxy)-2,2'-bipyridine] 2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-dioxolane ruthenium (II).

52. A method according to claim 46, wherein the electrochemiluminescent chemical moiety is bis (2,2' bipyridine) [4-(butan-1-yl)-4'-methyl-2,2'-bipyridine] ruthenium (II).

53. A method according to claim 46, wherein the electrochemiluminescent chemical moiety is bis (2,2'-

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bipyridine) [4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II).

54. A method according to claim 46, wherein the electrochemiluminescent chemical moiety is (2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene]{2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane} osmium (II).

55. A method according to claim 44, wherein the electrochemiluminescent chemical moiety bis(2,2'-bipyridine)[4-(4'-methyl-2,2'-bipyridine)butylamine] ruthenium II.

56. A method according to claim 44, wherein the electrochemiluminescent chemical moiety bis(2,2'-bipyridine)[1-bromo-4(4'-methyl-2,2'-bipyridine-4'-yl)butane] ruthenium (II)..

57. A method according to claim 46, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)maleimidoheptanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II).

58. A method according to claim 39, wherein the sample is derived from a solid, emulsion, suspension, liquid or gas.

59. A method according to claim 39, wherein the sample is derived from water, food, blood, serum, urine, feces, tissue, saliva, oils, organic solvents or air.

60. A method according to claim 39, wherein the sample comprises acetonitrile, dimethylsulfoxide,

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dimethylformamide, n-methylpyrrolidinone or tert-butyl alcohol.

61. A method according to claim 60, wherein the sample additionally comprises a reducing agent.

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62. A method according to claim 60, wherein the sample additionally comprises an oxidizing agent.

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63. A method for quantitatively determining in a predetermined volume of a multicomponent, liquid sample the amount of an analyte of interest present in the sample which comprises:

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a. contacting the sample with a known amount of a reagent (i) capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation and (ii) capable of competing with the analyte of interest for binding sites on a complementary material not normally present in the sample, and with a known amount of the complementary material, the contact being effected under appropriate conditions such that the analyte of interest and the reagent competitively bind to the complementary material;

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b. exposing the resulting sample to an amount of electrochemical energy from a suitable source effective to induce the reagent to repeatedly emit radiation, the exposure being effected under suitable conditions so as to induce the reagent to repeatedly emit electromagnetic radiation; and

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c. quantitatively determining the amount of radiation so emitted and thereby quantitatively determining the amount of the analyte of interest present in the sample.

5 64. A method according to claim 63, wherein the analyte of interest is theophylline, digoxin or human chorionic gonadotropin.

10 65. A method according to claim 63, wherein the reagent is the analyte of interest conjugated to an electrochemiluminescent chemical moiety.

15 66. A method according to claim 63, wherein the reagent is an analogue of the analyte of interest conjugated to an electrochemiluminescent chemical moiety.

20 67. A method according to claim 65 or 66, wherein the electrochemiluminescent chemical moiety is bis[4,4'-carbomethoxy)-2,2'-bipyridine]2-[3-(4-methyl-2,2'-bipyridine-4-yl)propyl]-1,3-dioxolane ruthenium (II).

25 68. A method according to claim 65 or 66, wherein the electrochemiluminescent chemical moiety is bis (2,2' bipyridine) [4-(butan-1-al)-4'-methyl-2,2'-bipyridine] ruthenium (II).

30 69. A method according to claim 65 or 66, wherein the electrochemiluminescent chemical moiety is bis (2,2'-bipyridine) [4-(4-methyl-2,2'-bipyridine-4'-yl)-butyric acid] ruthenium (II).

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70. A method according to claim 65 or 66, wherein the electrochemiluminescent chemical moiety is (2,2'-bipyridine)[cis-bis(1,2-diphenylphosphino)ethylene]{2-[3-(4-methyl-2,2'-bipyridine-4'-yl)propyl]-1,3-dioxolane} osmium (II).

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71. A method according to claim 65 or 66, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[4-(4'-methyl-2,2'-bipyridine)butylamine] ruthenium II.

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72. A method according to claim 65 or 66, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)[1-bromo-4(4'-methyl-2,2'-bipyridine-4-yl)butane] ruthenium (II).

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73. A method according to claim 65 or 66, wherein the electrochemiluminescent chemical moiety is bis(2,2'-bipyridine)maleimidohexanoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II).

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74. A method according to claim 57, wherein the complementary material is a whole cell, subcellular particle, virus, prion, viroid, lipid, fatty acid, nucleic acid, polysaccharide, protein, lipoprotein, lipopolysaccharide, glycoprotein, peptide, cellular metabolite, hormone, pharmacological agent, tranquilizer, barbiturate, alkaloid, steroid, vitamin, amino acid, sugar, non-biological polymer, synthetic organic molecule, organometallic molecule or inorganic molecule.

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75. A method for detecting the presence of different analytes of interest in a single liquid food or food homogenate sample which comprises:

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- 5 a. contacting the sample with a multiplicity of antibodies immobilized on a single support, each antibody being capable of forming a complex with a different analyte of interest so as to form complexes between the immobilized antibodies and the analytes of interest;
- 10 b. removing the immobilized antibodies from the sample;
- 15 c. treating the immobilized antibodies so as to remove unbound analytes;
- 20 d. contacting the complexes between immobilized antibody and analyte of interest with labeled antibodies capable of combining with the complexes so as to form a combination between the complexes immobilized antibody and analyte of interest and the labeled antibodies; and
- e. detecting the presence of labeled antibodies and thereby the presence of the of analytes of interest.
- 25 76. A method of claim 75 wherein the analytes of interest are microorganisms.
- 30 77. A method of claim 76, wherein the microorganisms are bacteria.
78. A method of claim 75, wherein the analytes of interest are antigens.

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79. A method of claim 78, wherein the antigens are enterotoxins.

80. A method of claim 78, wherein the antigens are aflatoxins.

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81. A method of claim 75, wherein the labeled antibodies are each labeled with the same detectable marker.

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82. A method of claim 81, wherein the detectable marker is an enzyme conjugate, a fluorescent moiety or a chemiluminescent moiety.

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83. A method of claim 81, wherein the detectable marker is an electrochemiluminescent chemical moiety capable of being induced to repeatedly emit electromagnetic radiation upon exposure to an amount of electrochemical energy from a suitable source effective to induce the moiety to emit radiation.

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84. A method of claim 83, wherein the electrochemiluminescent chemical moiety comprises ruthenium or osmium.

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85. A method of claim 75, wherein the immobilized antibodies are immobilized on a single nitrocellulose membrane.

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86. A kit for detecting different enterotoxins in a single sample which comprises:

a diagnostic reagent holder comprising a lower portion having a surface for immersion into the sample, at least one membrane attached to the

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surface, a multiplicity of monoclonal antibodies specific for different antigenic determinants on the enterotoxins immobilized to separate regions of the membrane;

5 a first rinsing solution comprising a buffered-aqueous solution containing a surfactant;

10 a detection solution comprising a monoclonal antibody conjugated to an enzyme and specific for a different antigenic determinant on each enterotoxin for which a monoclonal antibody immobilized on the membrane is specific;

15 a second rinsing solution comprising a buffered-aqueous solution;

a solution comprising a substrate of the enzyme which is converted by the enzyme into a product;

20 a solution comprising a dye which produces a visibly colored precipitate upon the reaction with the product formed by conversion of the substrate by the enzyme.

25 87. A kit of claim 86, wherein the membrane is nitrocellulose and the antigenic determinants are on Staphylococcal enterotoxins.

30 88. A kit of claim 87, wherein four monoclonal antibodies specific for Staphylococcal enterotoxins A, B, D and E are attached to the nitrocellulose membrane.

89. A kit of claim 88, wherein a monoclonal antibody which is specific for an antigenic determinant on more

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than one Staphylococcal enterotoxin is attached to the nitrocellulose membrane.

90. A kit of claim 89, wherein the monoclonal antibody is specific for antigenic determinants on Staphylococcal enterotoxins C₁, C₂ and C₃.

91. A kit of claim 90, wherein the enzyme is alkaline phosphatase, the substrate is 5-bromo, 4-chloro indolyl phosphate and the dye is nitroblue tetrazolium.

92. A method for detecting in a sample the presence of microorganisms comprising:

a. placing the sample in the growth zone of a unitary reactor, the reactor being provided with an immobilization zone and a growth zone;

b. treating the sample in the growth zone so that any microorganisms present in the sample multiply;

c. moving the sample to the immobilization zone so that microorganisms present in the sample are immobilized therein;

d. separately recovering the immobilized microorganisms;

e. treating the immobilized microorganisms with labeled antibody specific for the microorganisms so that complexes form between the antibody and the microorganisms; and

f. detecting the presence of complexes containing labeled antibody and thereby detecting the microorganism.

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93. A method of claim 92, wherein the labeled antibody is labeled with a detectable marker.

5 94. A method of claim 93, wherein the detectable marker is a detectable enzyme, a fluorescent moiety, or a chemiluminescent moiety conjugated to the antibody.

10 95. A method of claim 93, wherein the detectable marker is an electrochemiluminescent moiety conjugated to the antibody.

96. A method of claim 93, wherein the electrochemiluminescent moiety comprises ruthenium or osmium.

15 97. A method of claim 92, wherein the sample is water or food.

20 98. A method of claim 92, wherein the microorganism is a coliform bacterium.

25 99. A unitary reactor comprising a growth zone for treating a sample containing microorganisms so that the microorganisms grow and an immobilization zone for immobilizing microorganisms.

30 100. A reactor of claim 99, comprising a glass or plastic vial containing growth media as the growth zone and a membrane coated vial cap as the immobilization zone.

35 101. A reactor of claim 99, wherein the membrane is coated with an antibody specific for the microorganism.

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102. A reactor of claim 99, wherein the membrane is polystyrene, nitrocellulose or nylon.

103. A kit for detecting in a water sample the presence of coliform bacteria which comprises:

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a. at least one vial containing a non-selective lactose medium;

b. at least one Durham vial;

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c. at least one vial cap provided with a polystyrene insert;

d. a solution of bovine serum albumin;

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e. an anti-coliform bacteria monoclonal antibody-enzyme conjugate;

f. a buffered-aqueous solution; and

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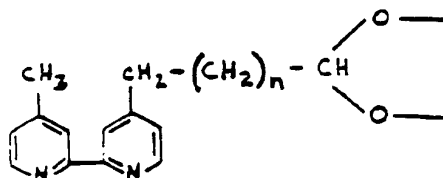
g. a solution of enzyme substrate.

104. A kit of claim 103, wherein the enzyme conjugated to the anti-coliform bacteria monoclonal antibody is calf intestinal alkaline phosphatase and the enzyme substrate is p-nitrophenyl phosphate disodium.

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105. A compound having the structure

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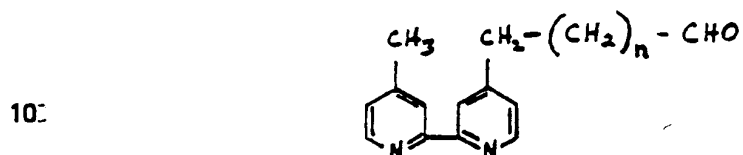
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wherein n is an integer.

106. A compound of claim 105, wherein n is 2.

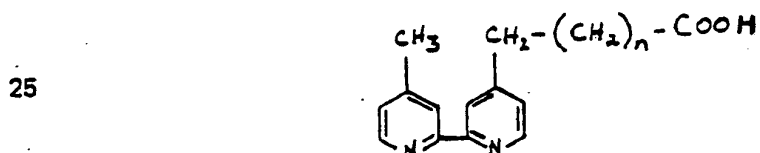
5: 107. A compound having the structure



15: wherein n is an integer.

108. A compound of claim 107, wherein n is 2.

20 109. A compound having the structure

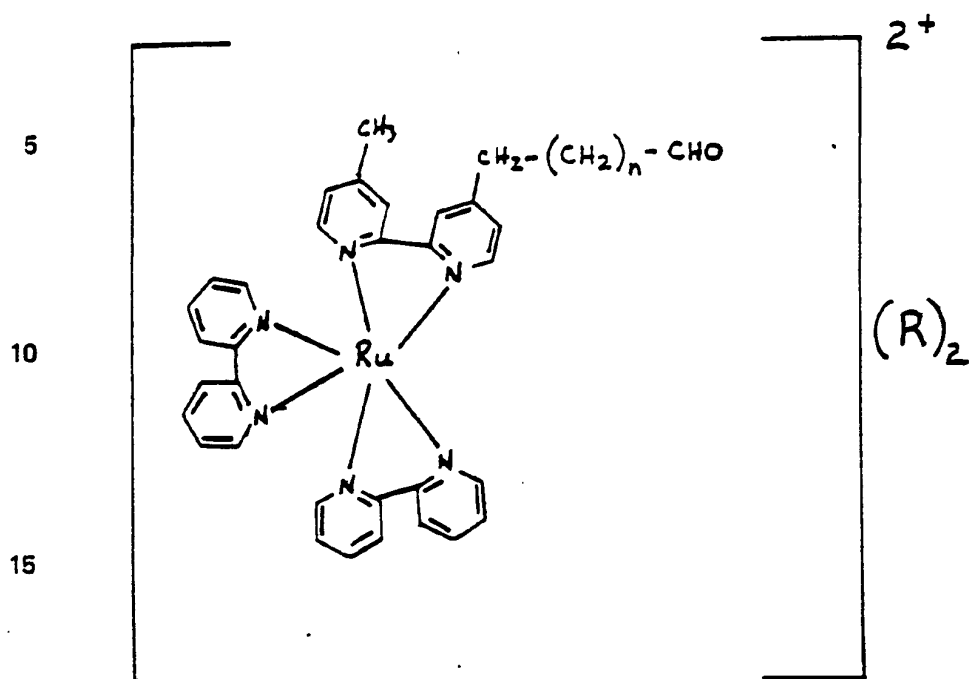


30: wherein n is an integer.

110. A compound of claim 109, wherein n is 2.

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111. A compound having the structure



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wherein R is an anion and n is an integer.

112. A compound of claim 111, wherein n is 2.

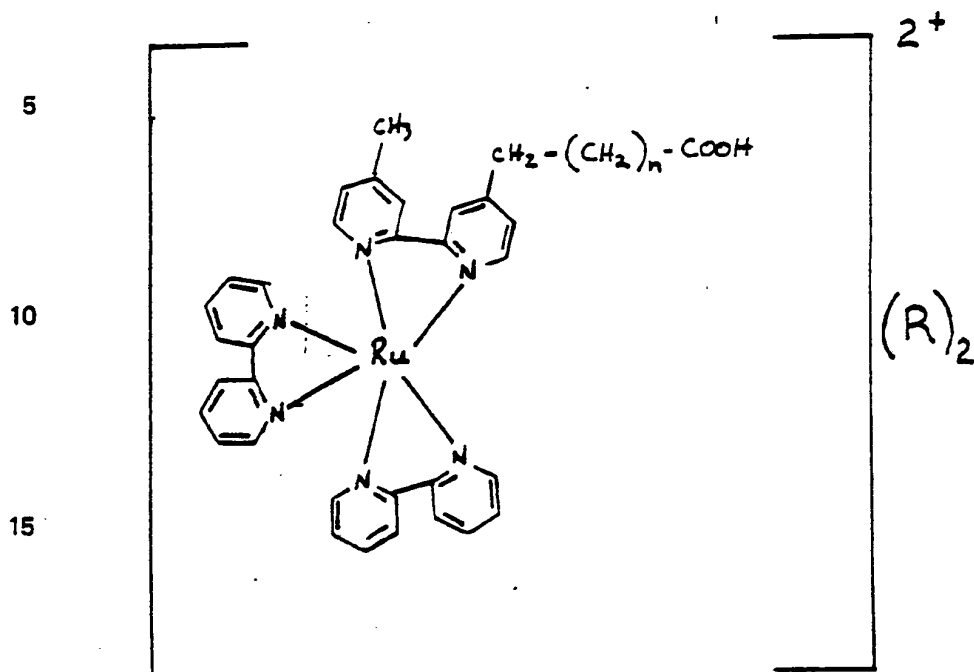
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113. A compound having the structure



wherein R is an anion and n is an integer.

114. A compound of claim 113, wherein n is 2.

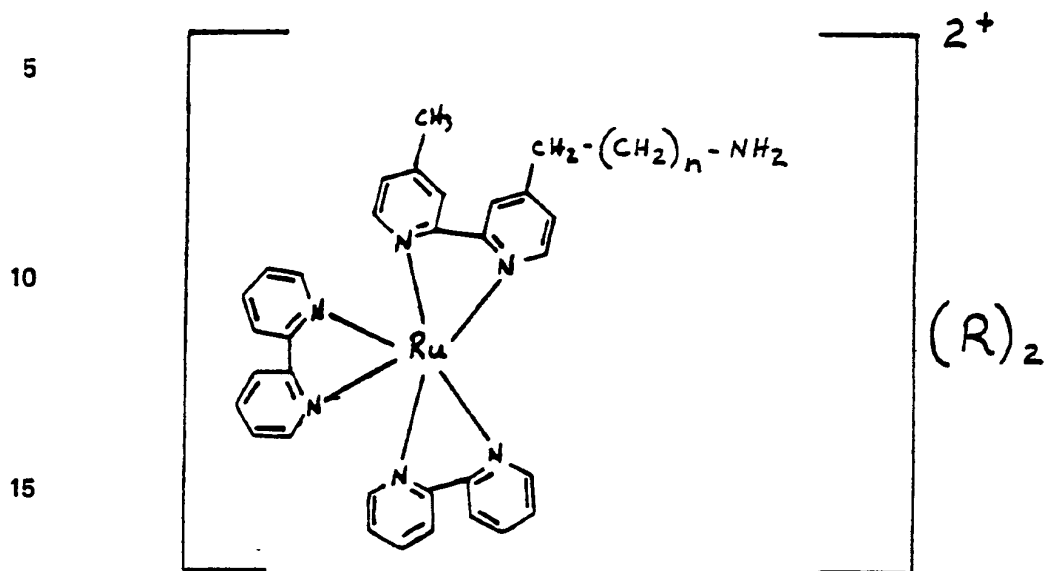
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115. A compound having the structure



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wherein R is an anion and n is an integer.

116. A compound of claim 115, wherein n is 3.

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117. A compound having the structure

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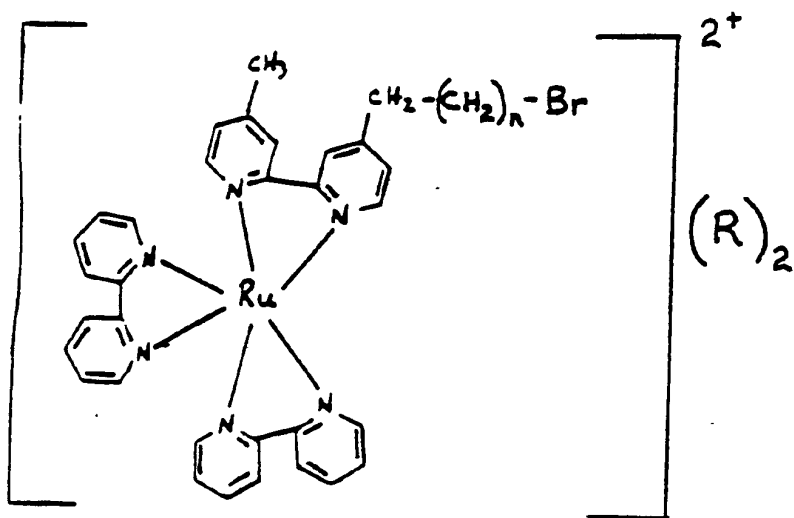
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wherein R is an anion and n is an integer.

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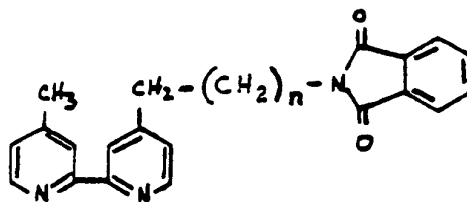
-227-

118. A compound of claim 117, wherein n is 3.

119. A compound having the structure

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15

wherein n is an integer.

20 120. A compound of claim 119, wherein n is 3.

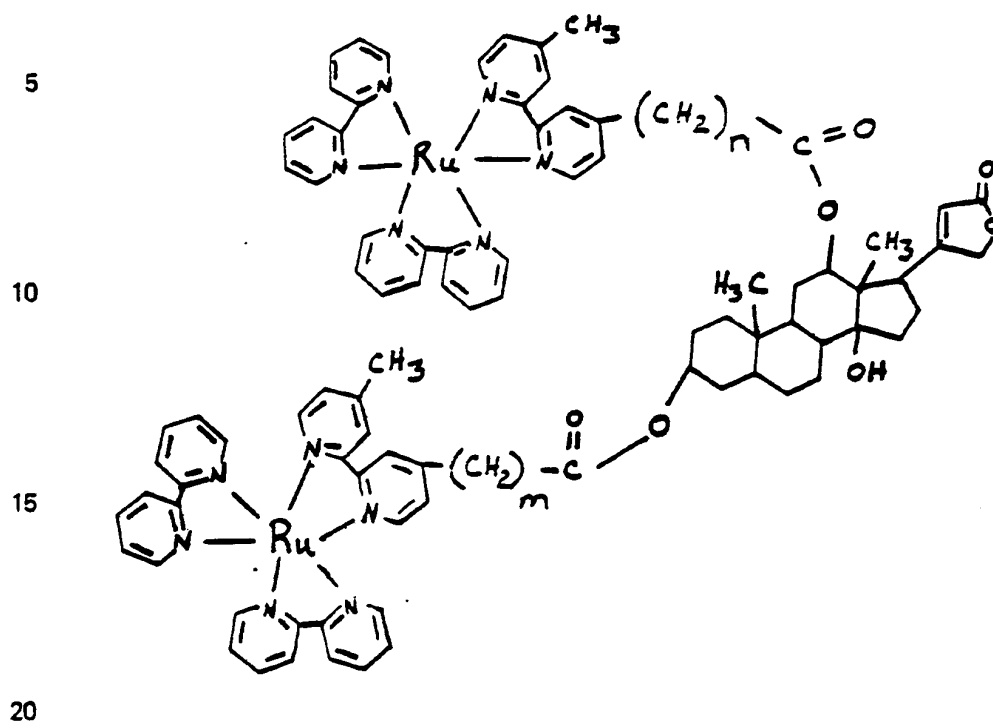
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-228-

121. A compound having the structure



wherein m and n are each integers which may be the same or different.

122. A compound of claim 121, wherein m and n are both 3.

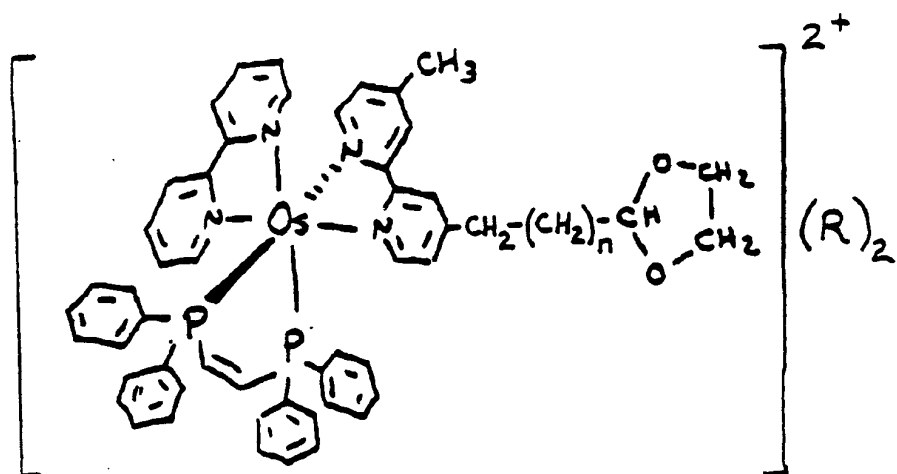
123. A compound having the structure

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wherein R is an anion and n is an integer.

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30

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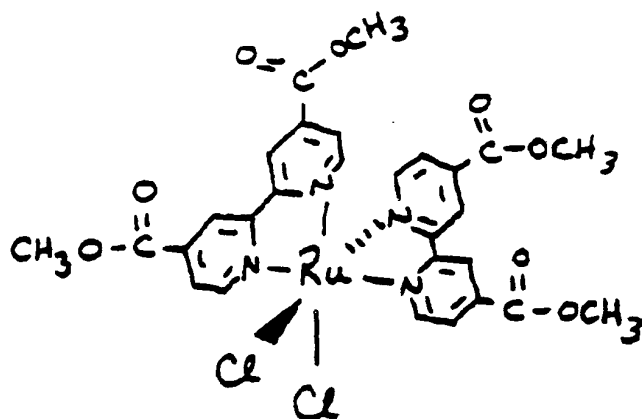
124. A compound of claim 123, wherein n is 2.

125. A compound having the structure

5

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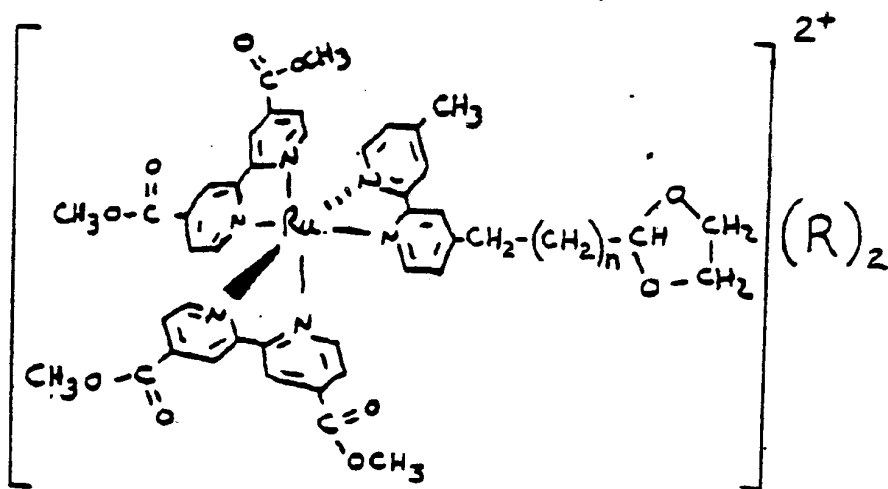


126. A compound having the structure

20

25

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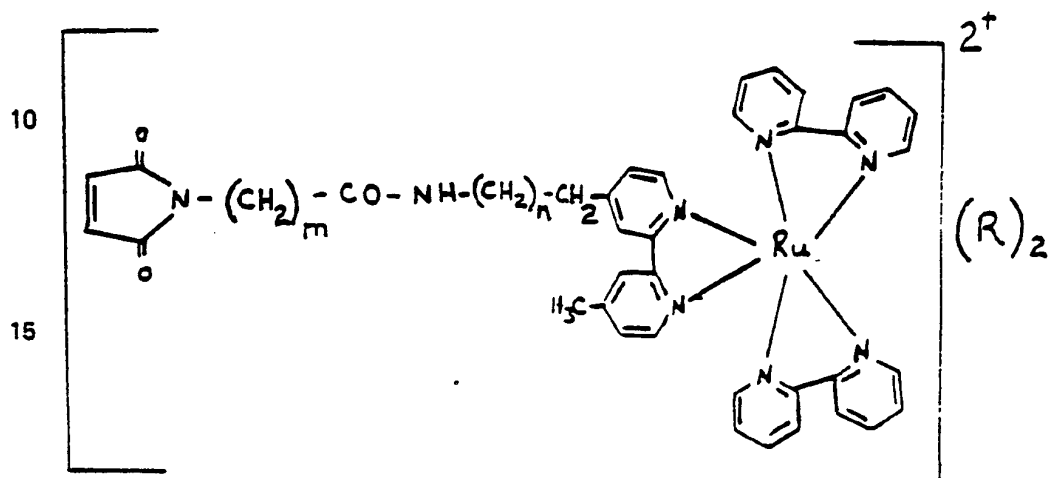
wherein R is an anion and n is an integer.

127. A compound of claim 122, wherein n is 2.

35

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128. A compound having the structure



wherein R is an anion and m and n are each integers which may be the same or different.

129. A compound of claim 128, wherein m is 5 and n is 3.

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130. A composition of matter having the structure

wherein:



5

X represents one or more nucleotides which may be the same or different, one or more amino acids which may be the same or different, an antibody, an analyte of interest or an analogue of an

10

Y represents a linker group having at least one carbon atom and attached to X and Z; and

15

Z represents a compound of claim 111, 113, 115, 117, 123, 126 or 128.

131. A composition of matter of claim 130, wherein X is theophylline.

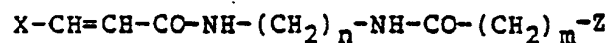
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132. A composition of matter of claim 130, wherein X is digoxigenin.

133. A composition of matter of claim 130, wherein X is a peptide derived from human chorionic gonadotropin.

25

134. A composition of matter having the structure



30

wherein:

X represents one or more nucleotides which may be the same or different;

35

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Z represents an electrochemiluminescent chemical moiety;

5 n represents an integer greater than or equal to 1; and

m represents an integer greater than or equal to 1.

10 135. A composition of matter of claim 134, wherein X is thymidine attached to CH at carbon 5, n is 7 and m is 3.

136. A composition of matter of claim 135, wherein Z is bis (2,2' - bipyridine) [4-(butan-1-yl)-4' methyl-2,2'-
15 bipyridine] ruthenium (II).

137. A composition of matter of claim 136, wherein the thymidine nucleotide is a 3' terminal nucleotide attached to the nucleotide sequence

20

TCACCAATAAACGGCAAACACCATCCGGTCCTGCCAG

138. A composition of matter having the structure

25 $[T-Y-Z]^{2+}(R)_2$

wherein:

T represents theophylline;

30 Y represents a linker group attaching T to Z;

Z represents bis-(2,2'-bipyridine) [4-methyl-2,2'-bipyridine-4'-yl] ruthenium (II); and

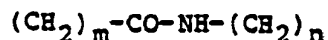
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R represents an anion.

139. A composition of matter of claim 138, wherein Y is attached to the carbon at position 8 of T.

5 140. A composition of matter of claim 139, wherein Y has the structure

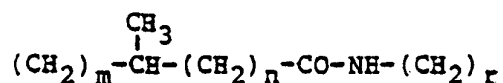


10 and wherein m and n represent an integer, which may be the same or different, greater than or equal to 1.

141. A composition of matter of claim 140, wherein m is 3 and n is 4.

15 142. A composition of matter of claim 140, wherein m and n are both 3.

20 143. A composition of matter of claim 139, wherein Y has the structure



25 and wherein m, n and r represent an integer, which may be the same or different, greater than or equal to 1.

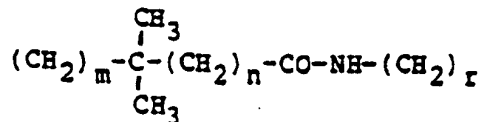
144. A composition of matter of claim 143, wherein m is 1, n is 1 and r is 4.

30

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145. A composition of matter of claim 139, wherein Y has the structure



5

and wherein m, n and r represent an integer, which may be the same or different, greater than or equal to 1.

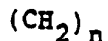
146. A composition of matter of claim 145, wherein m is 1, n is 1 and r is 4.

10

147. A composition of matter of claim 138, wherein Y is attached to the nitrogen at position 7 of T.

148. A composition of matter of claim 147, wherein Y has the structure

15



20 and wherein n is an integer greater than or equal to 1.

149. A composition of matter of claim 148, wherein n is 4.

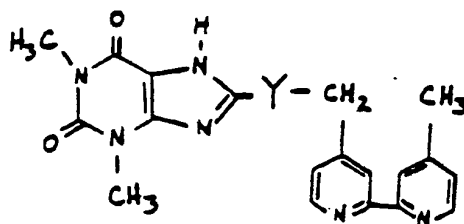
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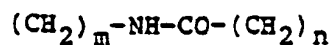
-236-

150. A compound having the structure



wherein Y is a linker arm.

151. A compound of claim 150, wherein Y has the structure

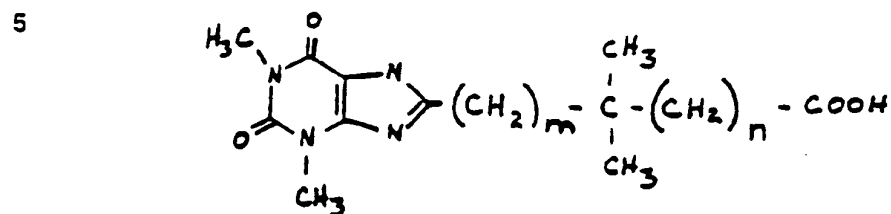


and wherein m and n are integers, which may be the same or different, greater than or equal to 1.

152. A compound of claim 151, wherein m is 3 and n is 2.

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153. A compound having the structure



15 wherein m and n are integers which may be the same or different.

154. A compound of claim 153, wherein m and n are both 1.

20

155. A composition of matter having the structure

X-Z

25 wherein:

X represent one or more amino acids which may be the same or different, comprising at least one amino acid which is cysteine or methionine; and

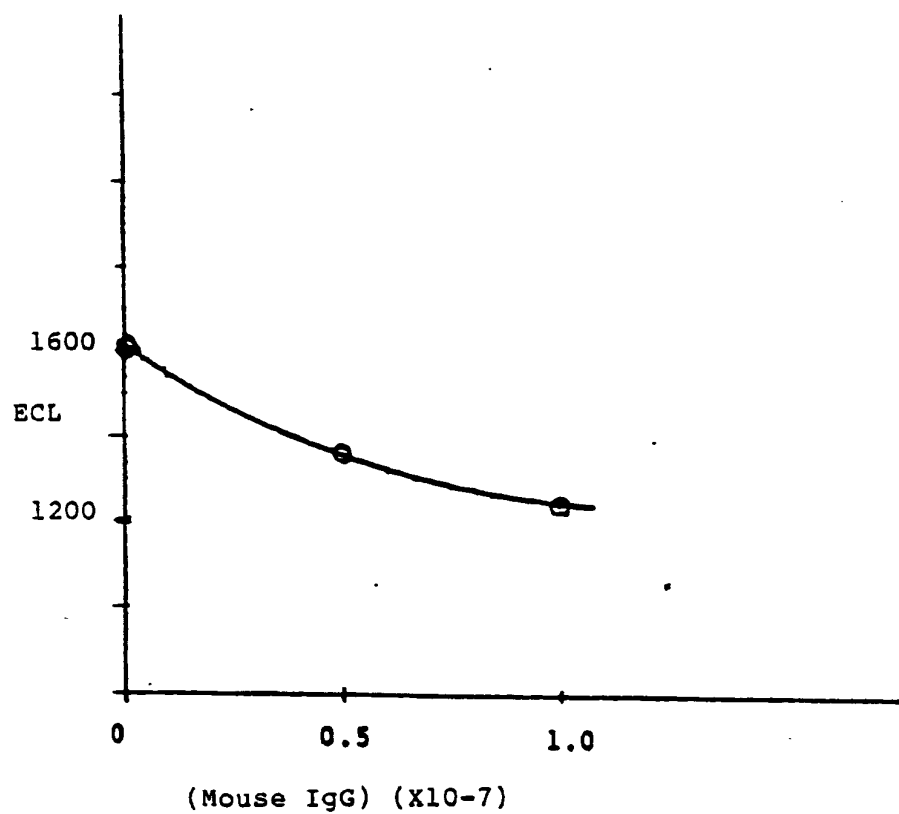
30

Z is bis(2,2'-bipyridine)maleimidohehexonoic acid, 4-methyl-2,2'-bipyridine-4'-butylamide ruthenium (II) attached by the carbon at position 3 or 4 of the maleimide to a sulfur substituent of cysteine or methionine.

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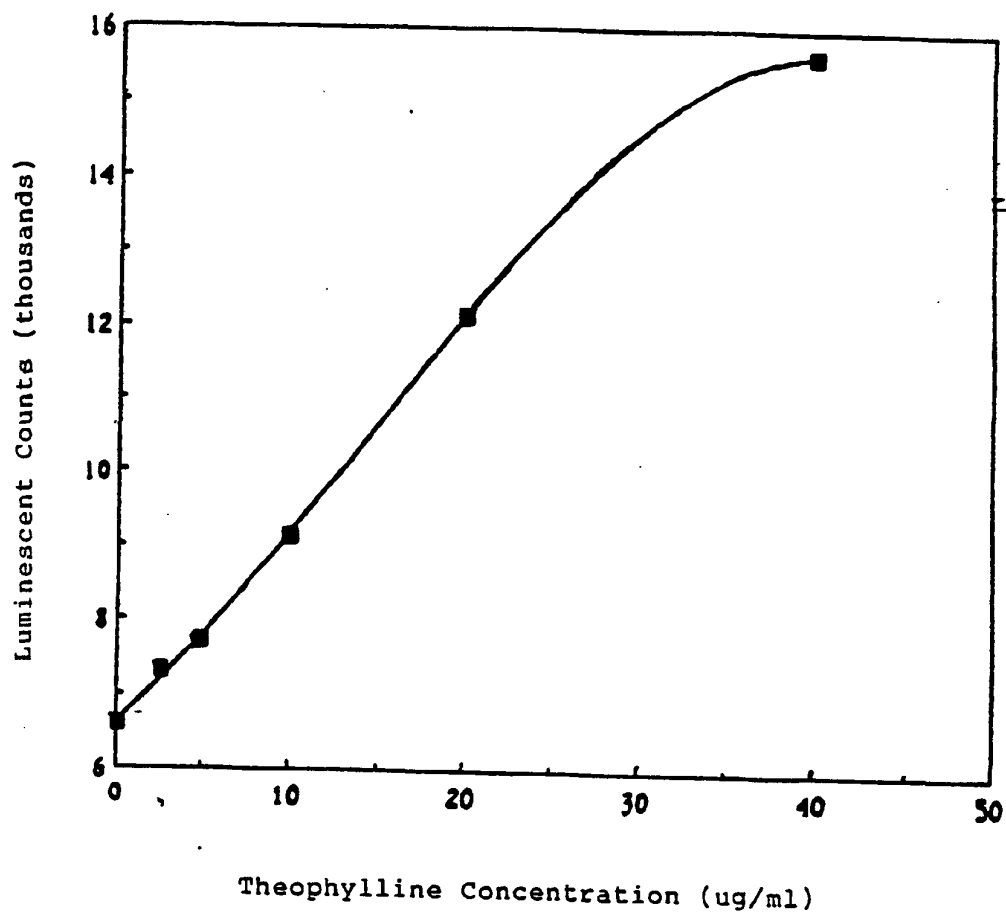
Figure 1



SUBSTITUTE SHEET

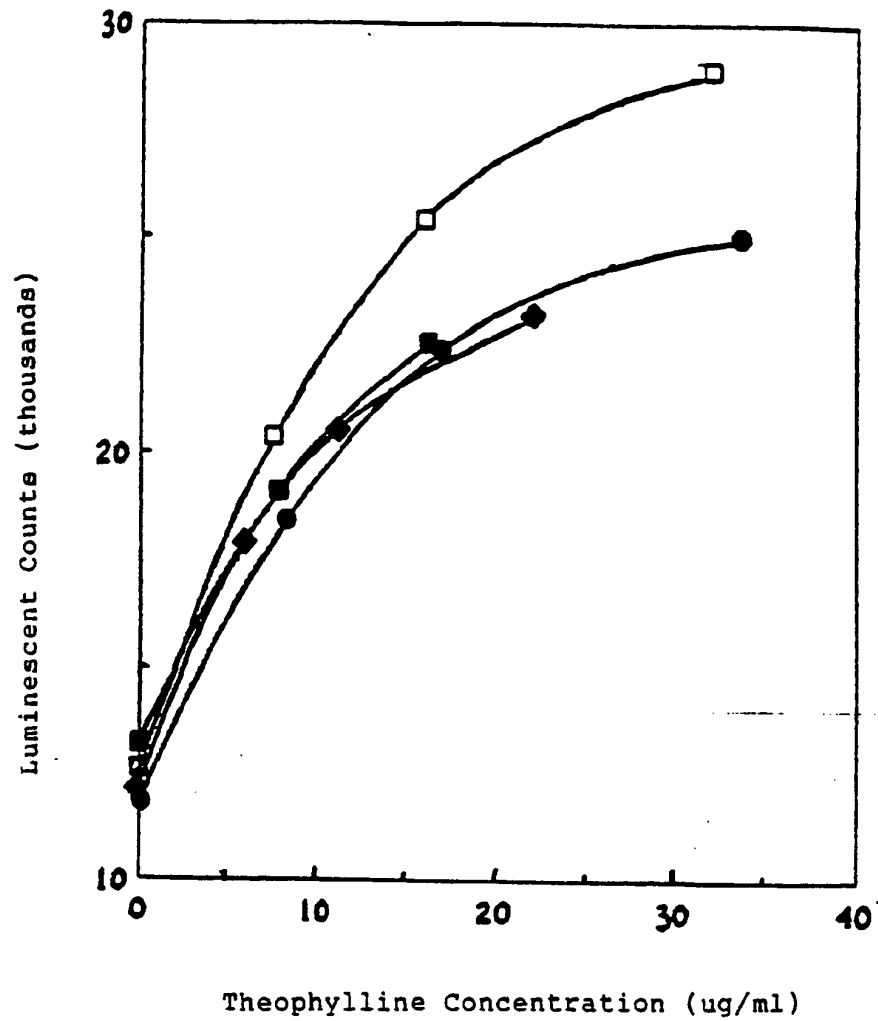
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Figure 2



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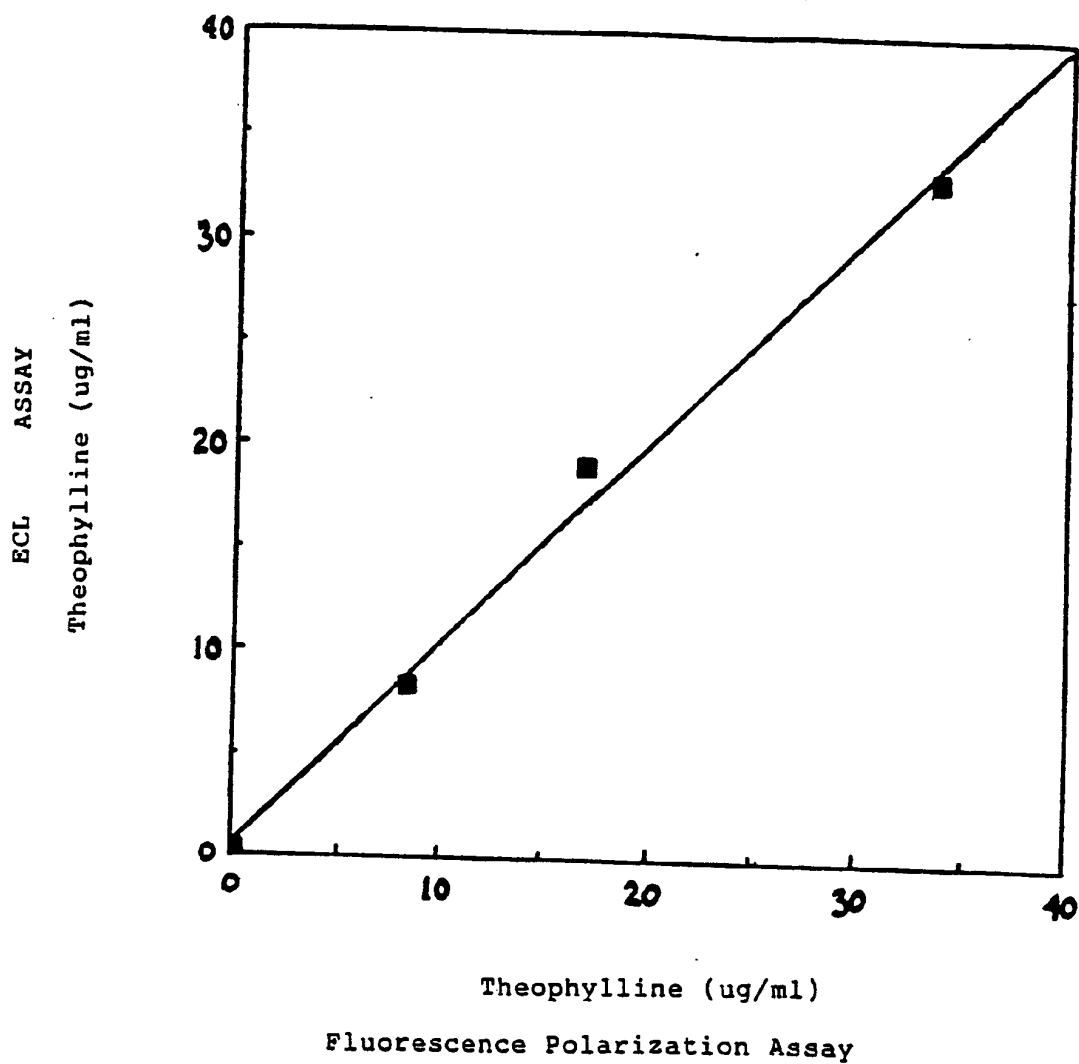
Figure 3



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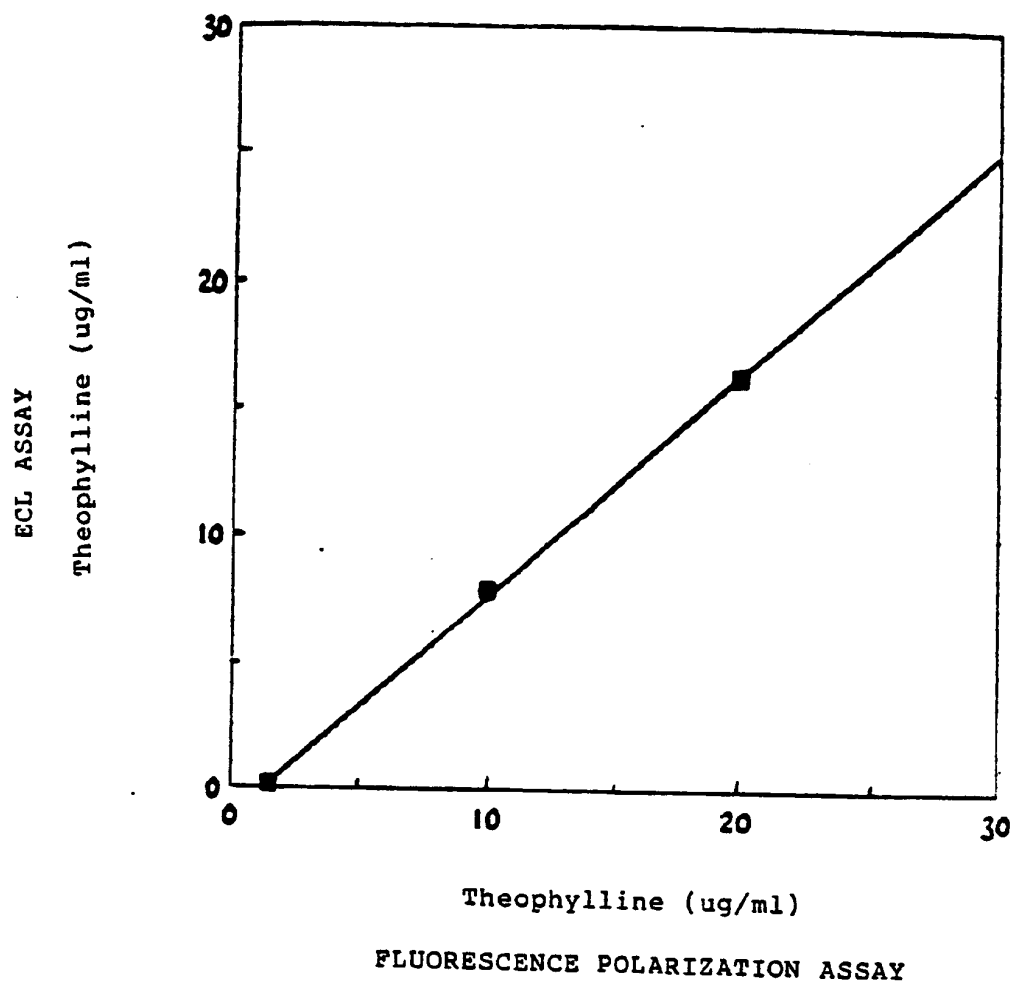
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Figure 4A



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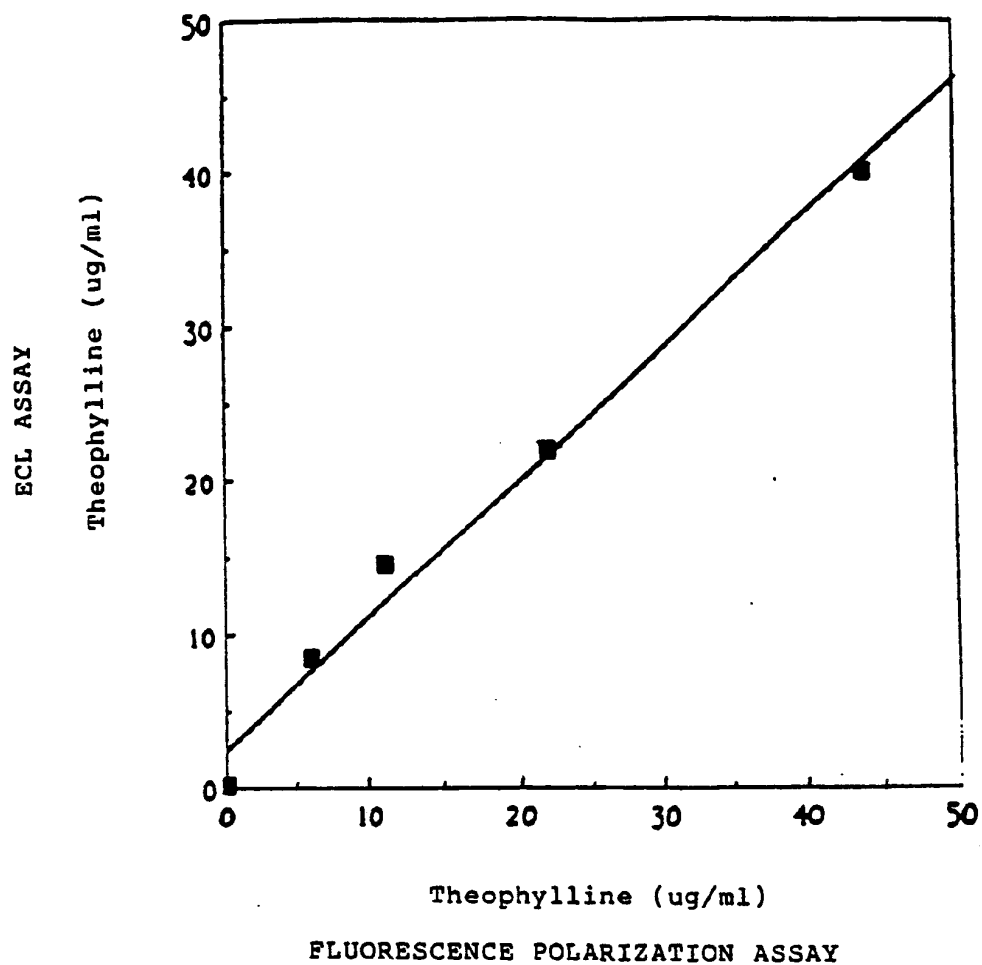
Figure 48



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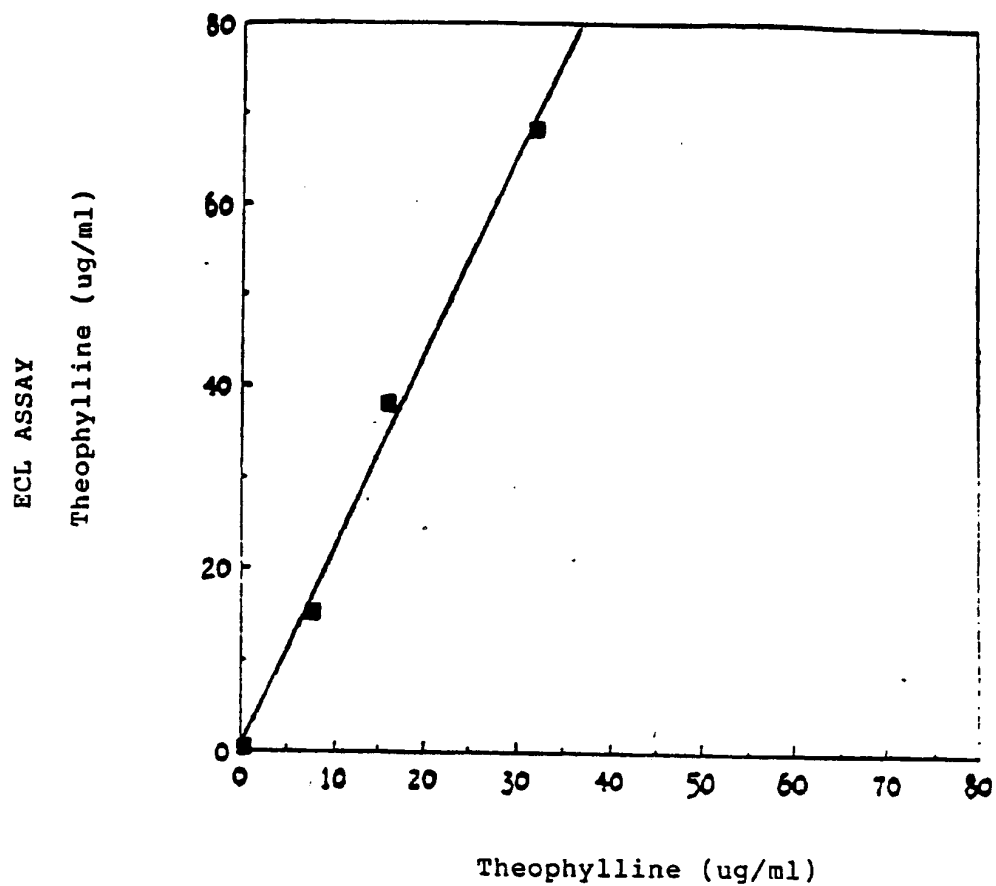
Figure 4C



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Figure 4D

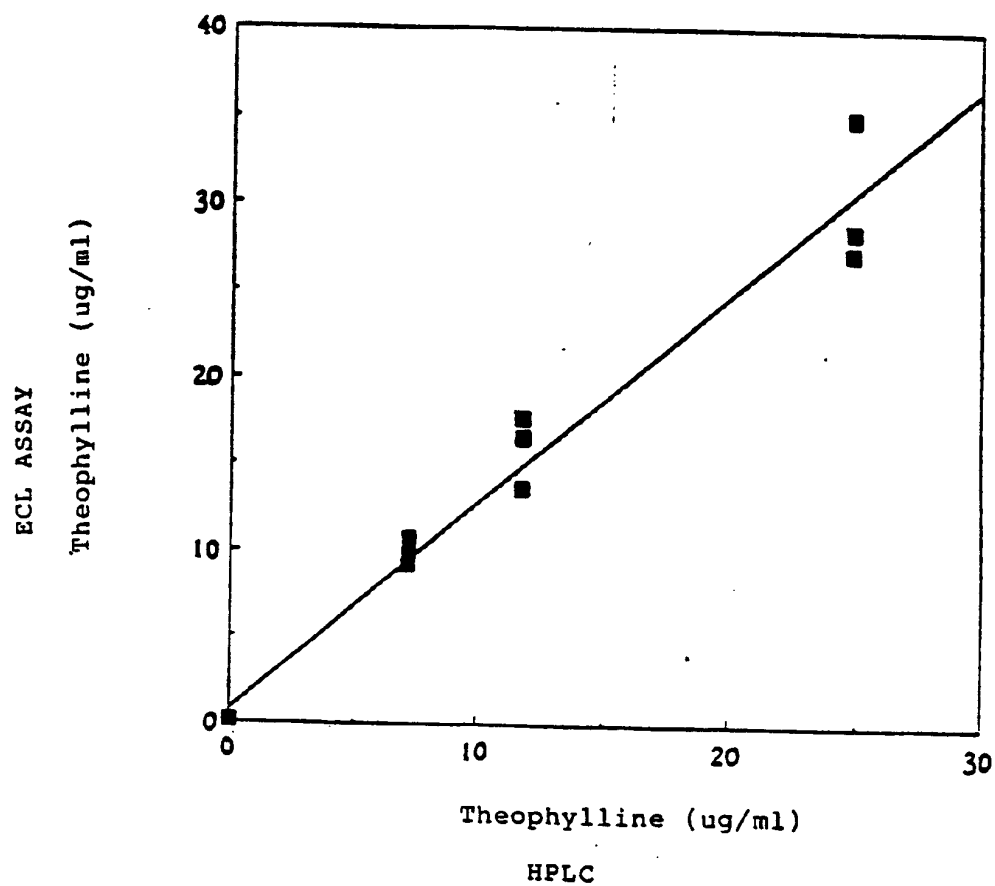


FLUORESCENCE POLARIZATION ASSAY

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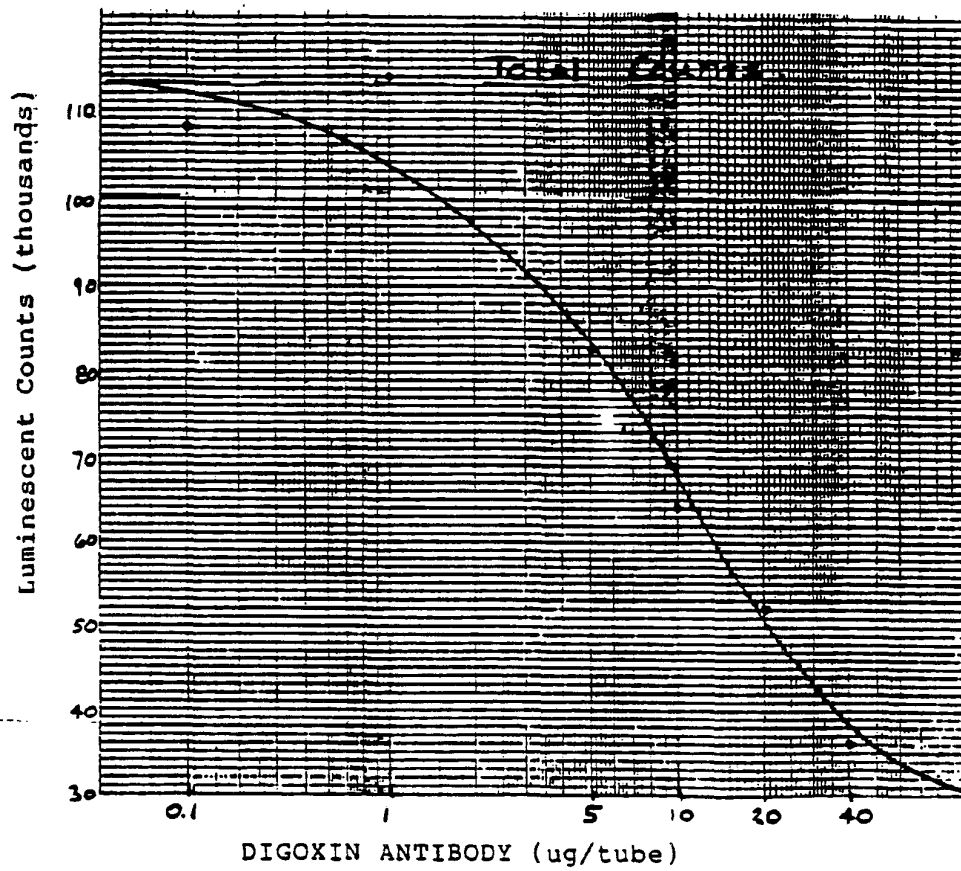
Figure 5



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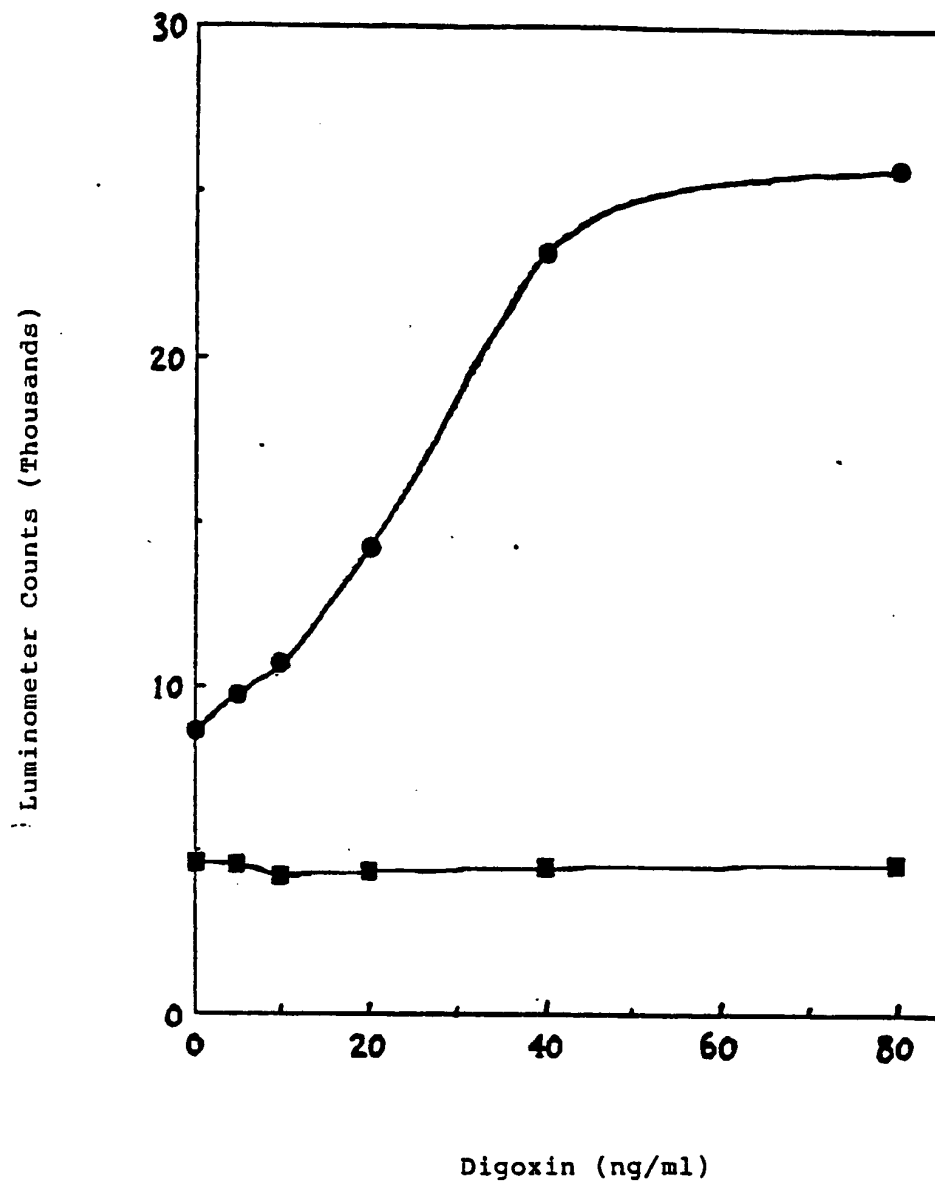
Figure 6



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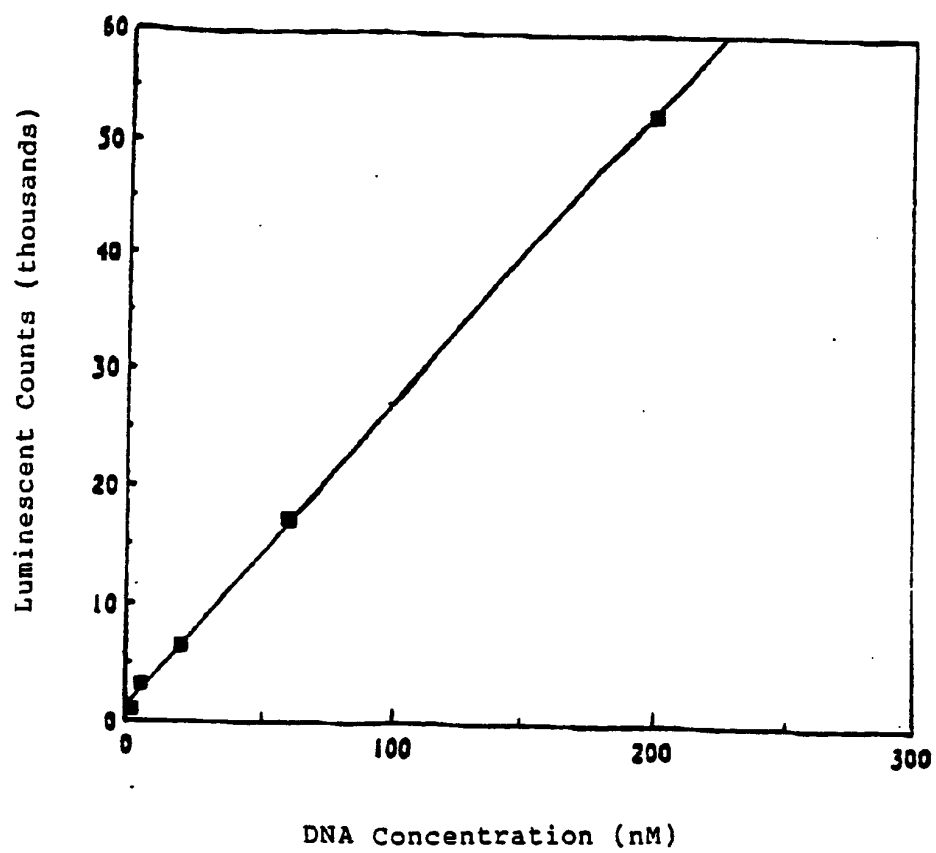
Figure 7



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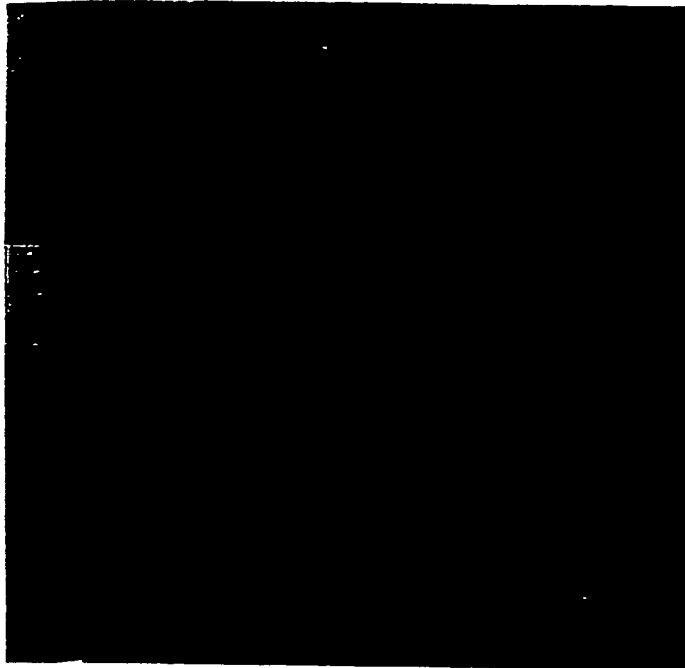
Figure 8



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Figure 9



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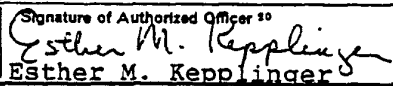
Figure 10



SUBSTITUTE SHEET

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US87/00987

I. CLASSIFICATION & SUBJECT MATTER (If several classification symbols apply, indicate all) ¹		
According to International Patent Classification (IPC) or to both National Classification and IPC IPC(4): G01N 33/532, 33/569, 33/76 U.S. Cl: 435/5,7; 436/501		
II. FIELDS SEARCHED		
Minimum Documentation Searched ²		
Classification System	Classification Symbols	
U.S.	435/5,6,7,808 436/501,518,519,536,544,547,805,806,815,818 935/78	
Documentation Searched other than Minimum Documentation to the Extent that such Documents are Included in the Fields Searched ³		
III. DOCUMENTS CONSIDERED TO BE RELEVANT ¹⁴		
Category ⁵	Citation of Document, ¹⁶ with indication, where appropriate, of the relevant passages ¹⁷	Relevant to Claim No. ¹⁸
Y, P	WO, A, 86/02734 (BARD) 09 May 1986, see page 13, line 23 - page 14, line 5; page 15, lines 3-29; page 18, line 23 - page 19, line 35; page 32, lines 13-22.	1-74,83, 84,130
Y	U.S., A, 4,280,815 (OBERHARDT) 28 July 1981, see column 1, lines 51-64, column 6, lines 19-23 and column 8, lines 23-33.	1-74,83, 84,130
Y	U.S., A, 4,233,144 (PACE) 11 November 1980, see column 1, lines 43-51, column 2, lines 57-68 and column 4, lines 61-68.	1-74,83, 84,130
A	U.S., A, 4,293,310 (WEBER) 06 October 1981, see column 4, lines 3-43.	
A	U.S., A, 4,205,952 (CAIS) 03 June 1980, see column 4, lines 39-50, column 5, lines 39-43 and column 10, line 56 - column 11, line 36.	
<div style="display: flex; justify-content: space-between;"> <div style="width: 45%;"> <p>¹⁵ Special categories of cited documents:</p> <p>"A" document defining the general state of the art which is not considered to be of particular relevance</p> <p>"E" earlier document but published on or after the international filing date</p> <p>"L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)</p> <p>"O" document referring to an oral disclosure, use, exhibition or other means</p> <p>"P" document published prior to the international filing date but later than the priority date claimed</p> </div> <div style="width: 45%;"> <p>"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention</p> <p>"X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step</p> <p>"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.</p> <p>"&" document member of the same patent family</p> </div> </div>		
IV. CERTIFICATION		
Date of the Actual Completion of the International Search ¹	Date of Mailing of this International Search Report ²	
26 AUGUST 1987	17 SEP 1987	
International Searching Authority ¹	Signature of Authorized Officer ¹⁰	
ISA/US	 Esther M. Kepplinger	

PCT/US87/00987

VI. OBSERVATIONS WHERE UNITY OF INVENTION
IS LACKING Form PCT/ISA210

- I. Claims 1-74, 83, 84 and 130 (where X is an analyte) drawn to an immunoassay using a label which limits radiation; Class 435 subclass 5.
- II. Claims 75-82 and 87-91 drawn to an immunoassay and kit; Class 435 subclass 7.
- III. Claims 92-104 drawn to a reactor, kit and immunoassay for microorganisms; Class 435, subclass 7.
- IV. Claims 105-110, 119 and 120 drawn to bipyridine compounds; Class 546, subclass 256.
- V. Claims 111-118, 123-129 and 155 (where X is one amino acid) drawn to ruthenium and osmium compounds; Class 546, subclass 2.
- VI. Claims 121-122 and 132 drawn to steroid compounds; Class 540, subclass 3.
- VII. Claims 130 and 134-137 drawn to nucleotide compounds; Class 536, subclass 27.
- VIII. Claims 130, 133 and 155 drawn to peptides; Class 530, subclass 300.
- IX. Claims 131 and 138-154 drawn to theophylline compounds; Class 544, subclass 225.

FURTHER INFORMATION CONTINUED FROM THE SECOND SHEET

V. ☐ OBSERVATIONS WHERE CERTAIN CLAIMS WERE FOUND UNSEARCHABLE ¹⁰

This International search report has not been established in respect of certain claims under Article 17(2) (a) for the following reasons:

1. ☐ Claim numbers because they relate to subject matter ¹² not required to be searched by this Authority, namely:

2. ☐ Claim numbers because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out ¹³, specifically:

VI. ☒ OBSERVATIONS WHERE UNITY OF INVENTION IS LACKING ¹¹

This International Searching Authority found multiple inventions in this international application as follows:

See Attachment Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims of the international application.

2. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims of the international application for which fees were paid, specifically claims:

3. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claim numbers: 1-74, 83, 84 and 130 (where X is the analyte).

4. ☐ As all searchable claims could be searched without effort justifying an additional fee, the International Searching Authority did not invite payment of any additional fee.

Remark on Protest

☐ The additional search fees were accompanied by applicant's protest.

☐ No protest accompanied the payment of additional search fees.